

# CO<sub>2</sub>:O<sub>2</sub> balance in boreal freshwaters in a changing climate

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Paper I

Paper II

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# List of Papers

## Paper I.

Alleson, L., Koehler, B., Thrane, J.E., Andersen, T. and Hessen, D.O., 2021. The role of photomineralization for CO<sub>2</sub> emissions in boreal lakes along a gradient of dissolved organic matter. *Limnology and Oceanography*, 66(1), pp.158-170.

## Paper II.

Alleson, L., Andersen, T., Dörsch, P., Eiler, A., Wei, J. and Hessen, D.O., 2020. Phosphorus availability promotes bacterial DOC-mineralization, but not cumulative CO<sub>2</sub> production. *Frontiers in microbiology*, p.2272.

## Paper III.

Alleson, L., Ström, L. and Berggren, M., 2016. Impact of photochemical processing of DOC on the bacterioplankton respiratory quotient in aquatic ecosystems. *Geophysical Research Letters*, 43(14), pp.7538-7545.

## Paper IV.

Alleson, L., Valiente, N., Dörsch, P., Eiler, A., Andersen, T., and Hessen, D.O. Drivers and variability of CO<sub>2</sub>:O<sub>2</sub> along a gradient from boreal to Arctic lakes. Submitted to *Biogeochemistry* in May 2022.





## Summary

Browning of lakes due to increasing concentration of coloured dissolved organic matter (CDOM), has become an increasingly studied topic over the past few decades. A large share of the CDOM consists of carbon (dissolved organic carbon; DOC), making browning of lakes associated with intensified carbon cycling in freshwater ecosystems. Affecting both microbial and photochemical mineralization of DOC and causing increasing light attenuation, lake browning may therefore regulate net heterotrophy by both inhibiting photosynthesis and stimulating DOC mineralization.

Using a combination of chemical, physical, and spectral data from lake surveys and in vitro measurements, this thesis presents an addition to the understanding of lake carbon cycling and the mechanisms behind the increasing CO<sub>2</sub> supersaturation in northern lakes. The research mainly focuses on DOC mineralization: both photochemical and microbial, and how this may vary with DOC quantity and quality, and nutrient availability.

In Paper I of this thesis, we study the contribution of photochemical DOC mineralization to total lake CO<sub>2</sub> production in Scandinavian lakes of different DOC concentration and colour. Although lakes differed substantially in color, depth-integrated photomineralization estimates were similar in all lakes, regardless of DOC concentrations and relative contribution of photochemical mineralization to total in-lake CO<sub>2</sub>-production was small (about 3 %). DOC concentrations were positively related to CO<sub>2</sub>-efflux and total in-lake CO<sub>2</sub> production but negatively related to primary production. We therefore conclude that enhanced rates of photochemical mineralization will be a minor contributor to increased heterotrophy under increased browning.

In vitro microbial DOC mineralization was studied in Paper II and III. In Paper II, we find cell specific respiration to be dependent on the carbon to phosphorus (P) ratio while total CO<sub>2</sub> production depended on DOC concentration and seemed to be unaffected by P additions.

P availability yields enhanced bacterial growth and lower cell specific respiration than under phosphorus limitation where the bacteria get rid of excess carbon via enhanced respiration. In addition to these P and DOC level effects, respiration rates responded in a non-monotonic fashion to temperature with increasing respiration rates from 15 to 25°C and a decrease above 30°C. This study highlights DOC as the major determinant of CO<sub>2</sub> production in boreal lakes, with P and temperature as significant modulators of respiration kinetics

Many studies assume a respiratory quotient (RQ = molar ratio of CO<sub>2</sub> produced to O<sub>2</sub> consumed) close to 1 when calculating bacterioplankton respiration. In Paper III, we find that the RQ is systematically higher than 1 when the bacterial metabolism in large part is based on photoproducts. By assuming an RQ of 1, bacterioplankton respiration in freshwater ecosystems may be greatly underestimated.

In Paper IV, we assess the drivers of CO<sub>2</sub> saturation in three groups of lakes separated by steps of approximately ten degrees latitude. CO<sub>2</sub> saturation was coupled to DOC concentrations and net heterotrophy in the southernmost boreal lakes with catchments dominated by coniferous forests. Contrastingly, further north in the arctic regime with sparsely- to unforested catchments, CO<sub>2</sub> saturation was instead tightly coupled to conductivity. This suggests that in the arctic, dissolved inorganic carbon inputs from the surrounding ecosystems and from carbonate weathering dominate as sources to lake CO<sub>2</sub> saturation. The results point to fundamentally different links between DOC and CO<sub>2</sub> in boreal and Arctic catchments, pinpointing the fundamental role of coniferous forests for organic and inorganic carbon dynamics in lakes.

# Introduction

Over the past few centuries, the anthropogenic impact on Earth has grown increasingly heavy, making the footprint we leave on Earth's surface irreversible. The current interval of time on Earth is therefore commonly denoted the Anthropocene (Crutzen, 2006). The industrial revolution called for increased coal burning, marking the start of serious human influence on the environment. Since the end of the Second World War, both industry and the world's population have increased close to exponentially, resulting in a concomitant increase in human impact on the environment.

One notable anthropogenic change is the exponential increase in atmospheric CO<sub>2</sub> concentration. These concentrations are today more than 50 % higher than pre-industrial levels (NOAA, 2022) and they keep increasing, with the major share of the increase taken place since the mid 1900's (Change, 2018). As a direct result of these increases in atmospheric greenhouse gases, the global average temperature has increased with roughly 1 °C during the industrial era. At high latitudes, the increase in surface temperatures is about double that of the global mean (Change, 2018, Masson-Delmotte et al., 2021).

In addition to CO<sub>2</sub>, burning of fossil fuels emits sulphur dioxide and nitrous oxides that, through chemical transformations in the atmosphere, form sulphuric acid and nitrous acid (Odén, 1968, Grennfelt and Hov, 2005, Reis et al., 2009). This has led to acid rains and thus a lowering of pH in soils and inland waters. Such acidification may cause severe damage to ecosystems with loss of acid-sensitive species and enhanced toxicity because of leaching metals (Raddum and Fjellheim, 1984, Lydersen et al., 2002). However, since the 1980s, the emissions of sulphur dioxide and nitrous oxides have decreased as the result of a number of successful measures taken. This reduction in emissions has led to a recover from acidification in northern soils and lakes over the past 30-40 years (Monteith et al., 2007).

Although we commonly speak about global warming; depending on geographic location, the ongoing climate change also leads to changes in precipitation patterns, and both intensified and increased numbers of extreme weather events and drought. In northern Europe, precipitation has increased during the past century (Dore, 2005), affecting the run-off from precipitation into the ground and within and between catchments.

Lakes are regarded as reflections of their catchments and react fast to changes in catchment characteristics such as catchment size, topography, geology, land use, vegetation, and soil properties. Lakes thus integrate catchment responses, serving as “sentinels” of changes at the catchment scale (Adrian et al., 2009). Additionally, lakes are sensitive to changes in temperature, precipitation patterns, cloudiness, and atmospheric composition, making lake properties respond rapidly to climate change (Assessment, 2004, Rosenzweig et al., 2007).

Higher temperatures and increased precipitation may be beneficial to plant growth. In fact, a so-called “greening” has been observed at high latitudes of Europe with increased forest biomass and an increased fraction of vegetation cover in the landscape (Tømmervik et al., 2009, Harsch et al., 2009, Larsen et al., 2011b, Myers-Smith et al., 2011, Berner and Goetz, 2022). However, enhanced movement of water leads to enhanced input of soluble minerals and substances from the terrestrial to the aquatic ecosystems. The enhanced biomass production together with enhanced movement of organic substances have led to increased inputs of terrestrially derived (allochthonous) dissolved organic matter (DOM), in northern inland waters (Larsen et al., 2011b, De Wit et al., 2016, Finstad et al., 2016). This DOM consists to a large part of dissolved organic carbon (DOC) and organic nutrients that may be recycled in the aquatic ecosystem by biogeochemical processes (Hessen and Tranvik, 1998).

Over the past few decades the view on lakes has shifted from being mere pipelines, transporting DOC from terrestrial ecosystems to the oceans, to active spots of carbon cycling,

and today we know lakes are a major component of the global carbon cycle (Cole et al., 2007, Tranvik et al., 2009). In lakes with high terrestrial influence, allochthonous DOC is a major source of energy to heterotrophs such that DOC increase has the potential to increase carbon mineralization rates (Hessen et al., 1990, Karlsson et al., 2007). Further, the brown colour of allochthonous DOC leads to lake browning and increased light attenuation (Kritzberg et al., 2020). Enhanced concentrations of allochthonous DOC thus has the potential to considerably perturb biogeochemical balances in lake ecosystems.

Changes in DOC and organic nutrient mineralization rates together with changes in light availability may alter the balance between autotrophic and heterotrophic processes and by extension also production further up the food web (Finstad et al., 2014).

This thesis revolves around the effects of enhanced inputs of allochthonous DOC on lake CO<sub>2</sub> and O<sub>2</sub> production and consumption. The results can contribute to predicting changes in lake ecology and biogeochemistry in a changing climate and environment.

### *Thesis Outline*

Lake CO<sub>2</sub>:O<sub>2</sub> balance depends on in-lake CO<sub>2</sub> and O<sub>2</sub> production and consumption. The main processes involved are DOC mineralization through heterotrophic respiration and photochemical processing on the one hand and primary production on the other. Further, DOC mineralization in the catchment may result in inputs of CO<sub>2</sub> to lakes via run-off and groundwater flow.

In this thesis, I focus on each mineralization process and aim to disentangle their relative importance to total lake CO<sub>2</sub> production and how this may change under an increasing DOC regime. In four chapters, I shed light on the different sources of CO<sub>2</sub> saturation in lake ecosystems, aiming to add to the knowledge of how northern lakes may

respond to the ongoing climate and environmental change. Although there is an overall agreement about the broad picture of how terrestrial DOC is cycled in lakes, there are still gaps to fill. Some of these gaps revolve around the relative importance between microbial and photochemical mineralization of DOC, and the relative importance of in-lake to CO<sub>2</sub> production to allochthonous CO<sub>2</sub> loadings for lake CO<sub>2</sub> saturation and evasion. Others revolve around how interacting factors such as substrate quantity and quality, nutrient availability, and temperature affect bacterial respiration, both on a cell-specific and on a community level. For a deeper understanding and to make reliable predictions of how future climate will affect aquatic ecosystems and, by extension, how to mitigate possible negative effects, more research on lake C cycling is needed.

In Paper I, we look at photochemical DIC production in boreal lakes of different DOC concentrations and colour. A few studies have estimated the relative importance of photomineralization to total lake DIC production and CO<sub>2</sub> evasion, mainly focusing on a single, or a limited set of lakes and the resulting estimates differ significantly (Granéli et al., 1996, Jonsson et al., 2001, Cory et al., 2014, Vachon et al., 2016, Groeneveld et al., 2016). One reason of the scarce number of large-scale studies is the need for specialized equipment and time-consuming experiments. We combined spectral data from the lakes with regression estimates between optical parameters and wavelength specific photochemical reactivity (Koehler et al., 2016) to estimate rates of photochemical DOC mineralization in 70 boreal lakes along an ecosystem gradient of DOC concentration. Further, we estimated total in-lake CO<sub>2</sub> production and efflux from lake chemical and physical data.

The aim is to find the relative importance of photochemical DOC mineralization to total in-lake CO<sub>2</sub> production in the study lakes. Shortwave radiation attenuates quickly in the water column and we expect all photo-reactive photons to be absorbed in all lakes. Although

the lakes differ substantially in DOC content and colour, we therefore hypothesise that the total amount of photochemical mineralization of DOC are similar in all lakes.

In Paper II, we focus on the biological mineralization of DOC via bacterioplankton respiration. The C to nutrient ratio of the substrate may have great implications on bacterioplankton metabolism (Hessen et al., 1994, Elser et al., 2009). At high ratios, the bacteria may dispose of excess C via increased CO<sub>2</sub> production (Hessen and Anderson, 2008), while at lower ratios, more C is allocated to growth (Hessen, 1992, Smith and Prairie, 2004). Furthermore, the temperature dependency of bacterioplankton metabolic rates interacts with the substrate quantity and quality (Apple et al., 2006). Microbial mineralization of DOC thus depends on several interacting factors. Although we can expect increases in bacterial growth and metabolism with increased temperatures and loadings of terrestrially DOC and nutrients, how the different environmental factors interact is still not fully elucidated.

Here we aim to fill in some of these knowledge gaps, studying heterotrophic respiration's dependence on additions of allochthonous DOC and P. We use chemical and physical data from 75 boreal lakes to estimate in-lake CO<sub>2</sub> production. The lakes span close to orthogonal ecosystem gradients in DOC and TP, allowing us to assess the interactive effects of these two parameters. To test for dynamic responses of bacterioplankton respiration to allochthonous DOC concentrations, nutrient availability, and temperature, we additionally performed experimental incubations. We hypothesize that while the cell-specific respiration and growth depends on the nutrient availability, respiration on the community level may not differ substantially under nutrient poor and nutrient rich circumstances. The major difference being the number of respiring cells.

In Paper III we continue focusing on bacterioplankton respiration, studying biological DOC mineralization of photochemically altered humic substances. Bacterial respiration is

often measured as O<sub>2</sub> consumption and converted to CO<sub>2</sub> production via a conversion factor. This conversion factor is the respiratory quotient (RQ = unit CO<sub>2</sub> produced per unit O<sub>2</sub> consumed) and is often assumed to be one (1). However, true bacterioplankton RQ can vary greatly (Cimblaris and Kalff, 1998, Berggren et al., 2012, Romero-Kutzner et al., 2015). Especially importantly, RQ is strongly dependent on the composition of the assimilated organic substrate (Berggren et al., 2012, Romero-Kutzner et al., 2015). Assimilation of organic substrates with large O content and high O:H ratio requires less O<sub>2</sub> from the surroundings, and hence, respiration is performed at a relatively high RQ (Dilly, 2001). Several of the most common LMW compounds that are released from reactions between DOC and ultraviolet (UV) sunlight are theoretically oxidized at RQs between 1.3 and 4. Although photooxidation is known to be an important process in natural freshwater systems, its potential role as RQ regulator has to our knowledge never been addressed.

We therefore tested the impact of UV radiation on bacterioplankton RQ, hypothesizing that photo-chemically processed DOC is used at a higher RQ than non-irradiated DOC because bacteria assimilate highly oxidized photoproducts such as organic acids. To test this hypothesis, we perform biological incubations of both irradiated and non-irradiated samples of natural lake water, simultaneously measuring the changes in O<sub>2</sub> and CO<sub>2</sub>.

Finally, in Paper IV, we take a step back and look at the broader picture, aiming to provide a better understanding of net heterotrophy and gas balance at the catchment scale. We aim to gain understanding on whether lake CO<sub>2</sub> saturation is dominated by in-lake processes or by allochthonous inputs and whether there is a latitudinal difference in drivers of lake CO<sub>2</sub>:O<sub>2</sub> ratio, notably related to forested or unforested catchments. There are conflicting reports of whether the main regulator of lake CO<sub>2</sub> saturation is in-lake DOC mineralization or export from terrestrial environments (Larsen et al., 2011a, Weyhenmeyer et



al., 2015, Nydahl et al., 2020, Jonsson et al., 2001). Some of the contradictory findings likely depend on climate, local hydrology, catchment slopes, water retention time, and not the least catchment properties like lake size or fraction and type of forest, bogs and wetlands (Puts et al., 2022, Valiente et al., 2022). We couple surface CO<sub>2</sub> and O<sub>2</sub> concentrations in 103 Norwegian lakes to environmental variables along a geographical gradient ranging from the boreal zone in southern Norway (58° N) through sub-Arctic northern Norway (69° N) to the high Arctic at Svalbard (79° N). The gradient reflects different catchment properties varying from dense spruce forest, via open birch forest to totally unforested catchments with thin soils in the high Arctic.

## Lake Browning

Lake browning has been attributed to a number of drivers. One commonly suggested driver is the decline in atmospheric sulphur deposition. As acid rains have decreased during the last few decades, soil pH has increased, increasing the solubility and thereby mobility of soil organic matter (Monteith et al., 2007, Finstad et al., 2016, De Wit et al., 2016). If recovery from acidification were the only driver of browning, it would eventually level off and return to a state similar to before acidification started (De Wit et al., 2016).

However, there is a lack of long-term records to statistically confirm this return to previous water colour, and other factors should also be considered important drivers of browning. According to some existing historical records from southern Sweden, water colour was not primarily suppressed in response to increasing S deposition, suggesting that the change was rather a result of the transition in land use from agriculture to forestry (Kritzberg, 2017).

Forest type also has a key role in the level of browning. Litter from conifers is generally of lower quality for microbial processing and thereby more recalcitrant compared

to that from deciduous trees (Duan et al., 2014). More DOC thus leaches from coniferous forests than from deciduous forests, explaining higher levels of browning in boreal compared to temperate forests. Moreover, iron (Fe) is known to contribute to light absorption in freshwater systems and thus add to the water colour (Canfield Jr et al., 1984). Complexes between Fe and humic substances may in turn affect water colour more than DOC and Fe alone (Maloney et al., 2005, Kritzberg and Ekström, 2011, Weyhenmeyer et al., 2014). Increase in Fe concentrations are generally higher in lakes in forested areas dominated with coniferous trees than in lakes in deciduous forests (Li and Richter, 2012). Transition towards dominance of coniferous forests over deciduous forests due to forestry practices may thus also have an impact on browning.

Climate and land use change over the last century have caused an increasing trend in vegetation cover. Increased terrestrial biomass renders an increased source of organic matter to be exported to inland waters (Larsen et al., 2011b, Finstad et al., 2016). Time series analyses have shown that wetter periods may give rise to increased water colour compared to dryer periods (Erlandsson et al., 2008). Increased precipitation due to climate change may thus be a driver of increased browning of inland waters as it results in enhanced run-off and by that enhanced transport of DOC. However, in high precipitation areas a further increase in precipitation may likely result in shorter retention times and enhanced dilution of terrestrial DOC (De Wit et al., 2016).

The individual importance of each mechanism behind browning is difficult to disentangle and therefore also predicting future water colour. Climate and environmental change will however promote high concentrations and movement of DOC and Fe in the aquatic continuum, having the potential to further promote browning.

## Lake CO<sub>2</sub> production

## *Light Attenuation and photochemical mineralization of DOC*

Terrestrial DOC is high in aromatic humic compounds and therefore efficient in absorption of ultraviolet (UV) and visible light (Lindell et al., 1995). Several studies have shown that increased concentrations of terrestrial DOC in lakes leads to increased light attenuation in the water column and thus a decreased euphotic zone, constraining primary production to a thinner surface layer of the water body (Williamson et al., 1996, Thrane et al., 2014, Seekell et al., 2015). Consequently, browning regulates lake heterotrophy and net CO<sub>2</sub> efflux to the atmosphere.

Besides being an essential source of energy for bacterioplankton (Hessen, 1992), the chromophoric terrestrial DOC is highly photo-reactive, especially in the UV waveband (Lindell et al., 1995). Photochemical processing of DOC might therefore be a significant part of carbon cycling in humic lakes. This processing includes both photomineralization of DOC to dissolved inorganic carbon (DIC) and partial photooxidation of high molecular weight humic compounds to low molecular weight compounds available for bacterial metabolism (Kieber et al., 1989, Bertilsson and Tranvik, 1998, Berggren et al., 2012).

DOC photo-reactivity is highest at the shortest wavelengths and decreases close to exponentially towards longer wavelengths (Koehler et al., 2016). The apparent quantum yield (AQY) of photomineralization is the mole of photochemically produced DIC per mole of light quanta absorbed by the DOC pool (Miller et al., 2002) The AQY is thus highest in the UV – blue area of the spectrum which are also the wavelengths that are attenuated fastest in the water column (Koehler et al., 2016). In lakes with high concentrations of coloured DOM (CDOM) the majority of the photomineralization takes place near the surface, while in clearer lakes with lower CDOM concentrations, light penetrates further down in the water column, letting photochemical mineralization take place at greater depths. A few studies have estimated the relative importance of photomineralization to total lake DIC production and

CO<sub>2</sub> evasion. Most of the studies focus on a single, or a limited set of lakes and the resulting estimates differ significantly (Granéli et al., 1996, Cory et al., 2014, Vachon et al., 2016, Dempsey et al., 2020). One reason of the scarce number of large-scale studies is the need for specialized equipment and time-consuming experiments (Koehler et al., 2016). In a first global estimate, Koehler et al. (2014) found the annual photochemical mineralization to account for about 10 % of the total CO<sub>2</sub> emissions from inland waters. As loadings of photo-reactive DOC to lakes increase, the relative importance of photomineralization to lake C cycling may change (Cory et al., 2014, Dempsey et al., 2020).

To Paper I, we estimated the DIC photoproduction in 70 boreal lakes along an ecosystem gradient in DOC concentration and colour. The range in DOC concentrations spanned a wide range and the results could give a hint of what may be the role of photomineralization for CO<sub>2</sub> emissions in lakes undergoing browning. According to regression estimates from Koehler et al. (2016), we used optical parameters to estimate AQY spectra. Together with site-specific irradiance spectra we could estimate photochemical DIC production in the lakes (Figure 1).

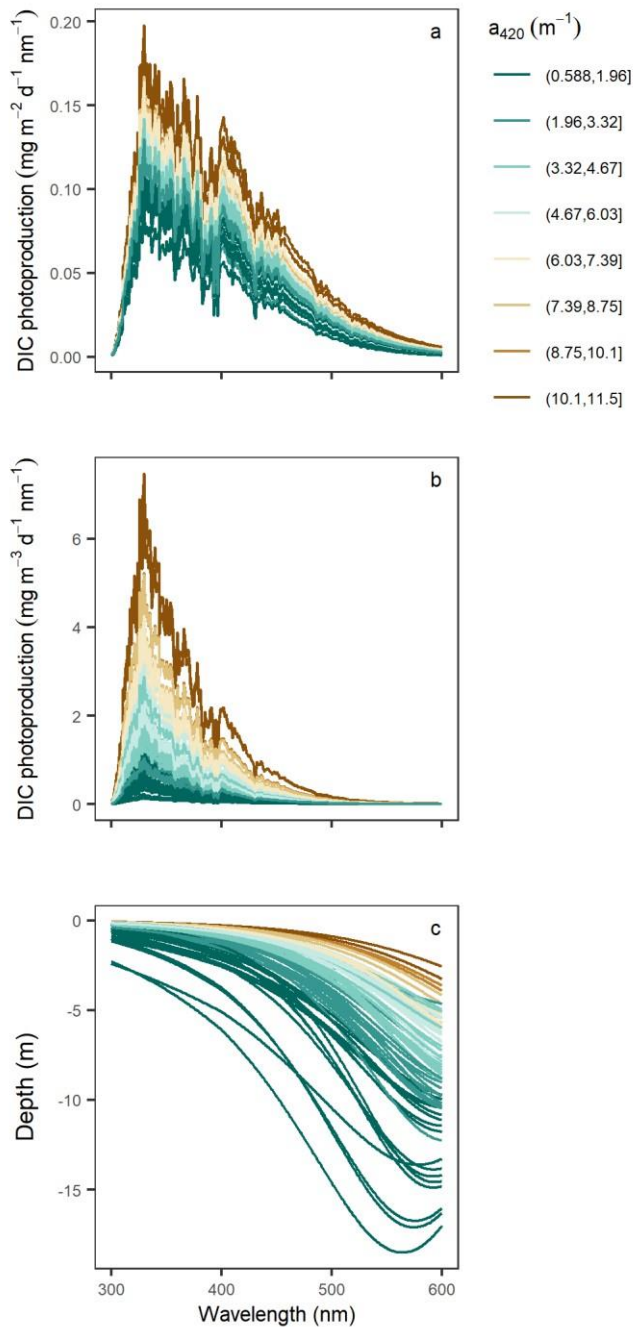


Figure 1. Estimated photoproduction spectra of dissolved inorganic carbon (DIC) from all 70 study lakes. In (a), the estimated areal DIC photoproduction ( $\text{mg C m}^{-2} \text{d}^{-1} \text{nm}^{-1}$ ) spectra are shown; and (b) shows the estimated volumetric DIC photoproduction ( $\text{mg C m}^{-3} \text{d}^{-1} \text{nm}^{-1}$ ) spectra just below the surface. In (c) the depth at which the volumetric DIC photoproduction ( $\text{mg C m}^{-3} \text{d}^{-1} \text{nm}^{-1}$ ) is 1 % of that just below the surface is shown, indicating also the depth that receives 1 % of incoming radiation. The color gradient goes from dark blue for clear lakes with low  $a_{420}$  to brown for brown lakes with high  $a_{420}$ .

We found that close to all of the short wavelengths are absorbed within the top meters of the water column in most lakes regardless of water colour (Paper I; Figure 1c). The

browner the lake, the faster the wavelengths are absorbed. However, in lakes deeper than about five meters, all photo-reactive wavelengths are absorbed. This indicates that further browning would not result in higher amounts of photochemical DIC production but rather move the photochemical reactions further up in the water column. Further, we found depth-integrated photomineralization estimates to be similar in all lakes. There was thus little difference in the amount of photochemical DIC production per lake area among lakes of different colour. Rather, the difference was at which depth in the water column the photochemical reactions took place.

The relative contribution of DOC photomineralization to in lake carbon cycling has been shown to vary, both between systems (Granéli et al., 1996, Molot and Dillon, 1997, Cory et al., 2014), and temporally within the same system (Vachon et al., 2016, Groeneveld et al., 2016), depending mainly on DOC quantity and quality. However, in our study, where the studied lakes spanned a wide range in DOC concentration, the relative contribution of photochemical mineralization to total in-lake CO<sub>2</sub>-production was about  $3 \pm 0.2\%$  in all lakes with no significant difference between systems (Paper I; Figure 2). Rather than photochemical mineralization of DOC, heterotrophic respiration was thus the major CO<sub>2</sub> source in all lakes, regardless of water colour.

In the boreal biome, photochemical DIC production has been estimated to correspond to about 9-12% of the annual lake CO<sub>2</sub> emissions of 47 – 59 Tg C yr<sup>-1</sup> (Kortelainen et al., 2006, Koehler et al., 2014). In accordance with these numbers, we estimated photochemically produced DIC to correspond on average to  $9 \pm 1\%$  of the total CO<sub>2</sub>-evasion from the study lakes. CO<sub>2</sub> evasion is generally higher in brown than in clear lakes (Sobek et al., 2003, Humborg et al., 2010, Larsen et al., 2011a) while the contribution of photochemically produced DIC to CO<sub>2</sub> evasion was highest in clear lakes (Paper I).

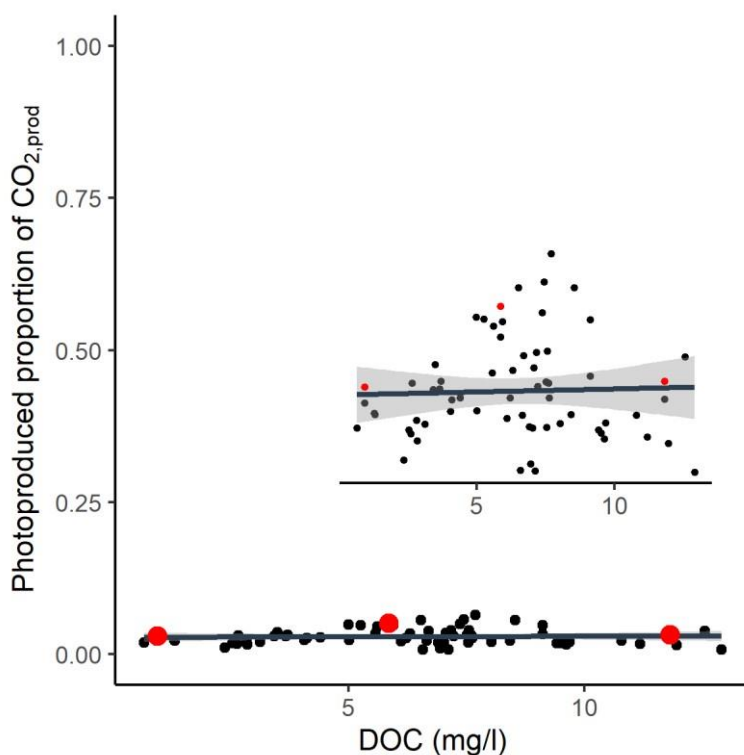


Figure 2. The relative proportion of photoproducted DIC to total  $\text{CO}_2$  production in the 70 study lakes in Paper I (the inset figure is zoomed in on the y-axis). There was no significant difference between lakes of different DOC concentrations.

The lakes in Paper I differed widely in colour from clear to dark brown and both  $\text{CO}_2$  efflux and total in-lake  $\text{CO}_2$  production were positively related to DOC concentration.

Although browner lakes most likely will result in enhanced  $\text{CO}_2$  evasion, the contribution of enhanced rates of photochemical DIC production will presumably be minor.

Photochemical processing of DOC may however give other end-products than  $\text{CO}_2$ . Organic nutrients associated to DOC are photo-mineralized and made available for consumption by both aquatic autotrophs and heterotrophs. Further, partial photooxidation of DOC transforms recalcitrant DOC to more biologically available organic compounds such as carboxylic acids (Bertilsson and Tranvik, 1998, Bertilsson and Tranvik, 2000). Aquatic heterotrophs often preferentially consume such partially photooxidized compounds over the high molecular weight non-photodegraded compounds (Berggren et al., 2012). Although

most photochemical processes occur in the surface layer, downward mixing makes photochemically produced carboxylic acids a potential source of labile DOC in the entire mixed layer. Indirect microbial mineralization of partial photooxidation products may be as important as direct photomineralization (Bertilsson and Tranvik, 1998).

Finally, light absorption by DOC may be beneficial to aquatic organisms as it may protect from harmful UV-radiation (Williamson et al., 1996, Dillon and Molot, 2005). However, as the coloured DOC absorbs light, it re-emits heat. This may modify the stratification properties of the water column (Williamson et al., 2015). Shallower mixed layers and stronger and longer periods of stratification yields colder temperatures and anoxia in the hypolimnion (Couture et al., 2015). Anoxia in the hypolimnion controls lake productivity (Craig et al., 2015) and CH<sub>4</sub> production, which can be released to the atmosphere (Vuorenmaa et al., 2014). Moreover, a darker environment may disturb predator-prey interactions, especially for visual predators, potentially having great impacts on the ecosystem and its populations and individuals (Ranåker et al., 2012).

### *Heterotrophic respiration*

Heterotrophic bacteria consume DOC and convert it to CO<sub>2</sub> through bacterial respiration (BR) (del Giorgio et al., 1997, Duarte and Prairie, 2005) and to biomass through bacterial production (BP) (Jansson et al., 2006, Berggren et al., 2010, Del Giorgio and Cole, 1998). Oceanic and freshwater BR together may represent the single largest sink of organic carbon worldwide and DOC constitutes a major part of the bulk organic carbon globally (Del Giorgio and Williams, 2005, Drake et al., 2018). Aquatic bacteria are thus an essential part of the global carbon cycle.

Most lakes on Earth are supersaturated with and net emitters of CO<sub>2</sub> to the atmosphere (Cole et al., 2007, Raymond et al., 2013). Several studies have reported a positive correlation



between lake CO<sub>2</sub> emission and DOC concentration (Sobek et al., 2003, Larsen et al., 2011a, Bogard and del Giorgio, 2016), indicating that bacterial respiration dominate as main CO<sub>2</sub> source. Autochthonous (in-lake) production is however not the only source of DIC. Lateral flux of inorganic carbon produced in the catchment may account for a sizeable share of lake CO<sub>2</sub>, especially in small lakes with short retention times and long ice-free seasons (Weyhenmeyer et al., 2015).

Whether a lake is a net conduit of CO<sub>2</sub> is not necessarily a sign of net heterotrophy but could also reflect that catchment derived CO<sub>2</sub> exceeds photosynthetic uptake. Likewise, the magnitude of atmospheric CO<sub>2</sub> uptake in a net autotrophic system may be reduced by inputs of terrestrial CO<sub>2</sub>. There are conflicting reports of whether CO<sub>2</sub> produced in aquatic environments via DOM mineralization or exported from terrestrial environments is the main regulator of lake CO<sub>2</sub> flux (Larsen et al., 2011a, Weyhenmeyer et al., 2015, Nydahl et al., 2020, Jonsson et al., 2001). Some of the contradictory findings likely depend on climate, local hydrology, catchment slopes, water retention time, and not the least catchment properties like lake size or fraction and type of forest, bogs and wetlands (Puts et al., 2022, Valiente et al., 2022).

In Paper I and II, we estimated in-lake CO<sub>2</sub> production in boreal lakes using chemical and physical data. The lakes spanned close to orthogonal ecosystem gradients in DOC and total phosphorus (TP), allowing us to assess the interactive effects of these two parameters on CO<sub>2</sub> production and on CO<sub>2</sub> and O<sub>2</sub> saturation deficits. We found a strong negative correlation between O<sub>2</sub> and CO<sub>2</sub> saturation deficits such that lakes saturated with O<sub>2</sub> were also saturated with CO<sub>2</sub> (Figure 3). Further, we saw no correlation between total inorganic carbon (TIC) and CO<sub>2</sub> deficit. This together indicates that in the sampled boreal lakes, processes within lakes, especially microbial respiration was the predominant source of CO<sub>2</sub> supersaturation.

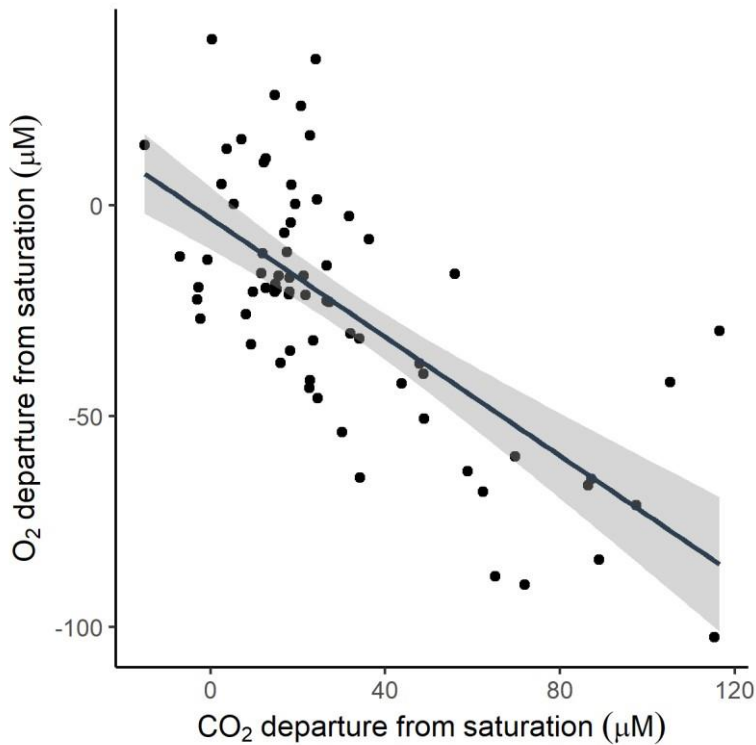


Figure 3. Lake O<sub>2</sub> departure from saturation with the atmosphere vs CO<sub>2</sub> departure from saturation with the atmosphere in 70 boreal lakes along an environmental gradient in DOC concentrations (Paper I; CI slope: (-9, -5); CI intercept: (-10, 4); R<sup>2</sup> = 0.49; p << 0.001).

There, however, seems to be a difference in dominating CO<sub>2</sub> source between systems of differing catchment type. In Paper IV, we coupled surface CO<sub>2</sub> and O<sub>2</sub> concentrations in 103 Norwegian lakes to environmental variables along a geographical gradient ranging from the boreal zone in southern Norway (58° N) through sub-Arctic northern Norway (69° N) to the high Arctic at Svalbard (79° N). The gradient reflects different catchment properties varying from dense spruce forest, via open birch forest to unforested catchments with thin soils in the high Arctic.

We saw a clear distinction in drivers of CO<sub>2</sub> saturation between different regions. Boreal lakes, with catchments dominated by coniferous forests followed the expected pattern with both CO<sub>2</sub> and O<sub>2</sub> saturation being largely dependent on DOC concentrations and negatively related to one another, suggesting enhanced net heterotrophy with increased DOC inputs.

In Arctic lakes, despite differences between sub-Arctic and high-Arctic sites, we saw no correlation between DOC and CO<sub>2</sub>, yet these sites were also to a large degree supersaturated with CO<sub>2</sub> and could be considered net heterotrophic. However, most Arctic lakes were also saturated or supersaturated with O<sub>2</sub>, indicating low respiratory activity (Figure 4). This is in agreement with generally nutrient-poor conditions and low levels of primary production. We also saw a positive correlation between CO<sub>2</sub>:O<sub>2</sub> ratio and conductivity, while the influence of DOC concentration was weak or non-significant. This may suggest that the major share of CO<sub>2</sub> in the Arctic lakes is of allochthonous origin, likely from organic carbon mineralization and carbonate weathering in the catchment soils, entering via groundwater flow (Rodríguez-Rodríguez et al., 2018, Puts et al., 2022). This points to fundamentally different links between DOC and CO<sub>2</sub> in boreal and Arctic catchments, pinpointing the role of coniferous forests for organic and inorganic carbon dynamics in lakes.

Although there may be pronounced regional differences within this vast area, the wide spatial gradient across climatic regions and catchment properties could provide insights relevant to larger parts of both the boreal and the arctic biome.

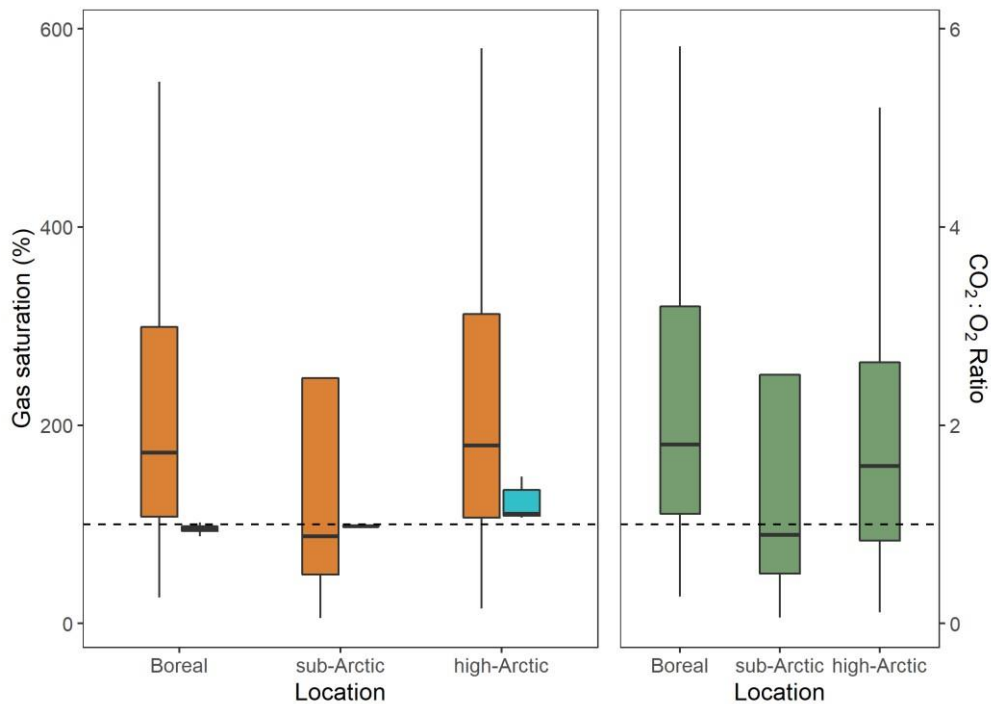


Figure 4. Box plot of lake CO<sub>2</sub> saturation (red), O<sub>2</sub> saturation (blue), and CO<sub>2</sub>:O<sub>2</sub> ratio (green) in three different geographical regions. Dashed line represents the 100 % saturation (to the left) and a CO<sub>2</sub>:O<sub>2</sub> ratio of 1 (to the right).

### *Substrate stoichiometry: nutrients*

Heterotrophic respiration has a key role in C cycling in boreal lakes and will most likely increase with enhanced loadings of allochthonous DOC and enhanced temperatures (Apple et al., 2006, Larsen et al., 2011a, Lapierre et al., 2013). However, questions remain about bacterial metabolism's response to enhanced loadings of allochthonous DOC and nutrients and to increasing temperatures in a changing climate.

Current knowledge shows that nutrient stoichiometry, quantity and quality of the substrate, and temperature are main factors, governing bacterioplankton metabolism. Bacteria have a high nutrient demand and the C to nutrient ratio of the substrate and the availability to inorganic nutrients governs the carbon use efficiency (Hessen, 1992, Hessen et al., 1994, Elser et al., 2000). Low such C:nutrient regimes promote biomass production while under

high C:nutrient regimes, bacteria may dispose of excess C via enhanced respiration (Hessen, 1992, Hessen and Anderson, 2008).

The efficiency of bacterial carbon use can be approximated as the share of the total assimilated organic carbon used for BP (bacterial growth efficiency:  $BGE = BP / (BP + BR)$ ). This determines to what degree bacterial metabolism results in bacterial biomass production or in mineralization of organic carbon (Del Giorgio and Cole, 1998).

The carbon to nutrient ratio therefore has great impact on the cycling and fate of carbon in a planktonic habitat. Phosphorus (P) is the most commonly reported limiting nutrient for BP and P availability thus regulates the use of DOC for growth (Vadstein, 2000). Planktonic DOC is generally more bioavailable and has a lower C:P ratio than allochthonous DOC. When DOC of both autochthonous and allochthonous origin is available, heterotrophic bacteria use the planktonic DOC for catabolic processes (Kritzberg et al., 2004).

Microorganism's activity and growth are constrained by temperature (Farrell and Rose, 1967, Madigan et al., 1997) with a general increase in metabolism with increased temperatures. However, studies show that the temperature dependency is stronger for bacterial respiration than for bacterial production (Rivkin and Legendre, 2001, Apple et al., 2006, Berggren et al., 2010, Kritzberg et al., 2010), and that this dependency may be constrained by nutrient availability. Furthermore, metabolic rates have been shown to be less temperature dependent for heterotrophic bacteria growing on labile autochthonous DOC than when growing on complex and recalcitrant allochthonous DOC (Ylla et al., 2012, Jane and Rose, 2018). Microbial mineralization of DOC thus depends on several interacting factors. Although we can expect an overall increased bacterial metabolism with increased temperatures and terrestrially derived DOC and nutrient loadings in lakes, there are still knowledge gaps of how the different environmental factors interact.

In Paper II, we coupled environmental factors to in situ CO<sub>2</sub> production rates in 75 Scandinavian lakes. The lakes spanned close to orthogonal ecosystem gradients in DOC and TP, allowing us to assess the interacting effect of these two parameters on CO<sub>2</sub> production. We found CO<sub>2</sub> production to have a unimodal response to increased DOC concentrations with a minimum around 5 mg DOC L<sup>-1</sup>. This indicates a shift in DOC pool from primarily autochthonous at lower DOC concentrations to primarily allochthonous at higher DOC concentrations. After the turning point and as the concentration of allochthonous DOC increases, the CO<sub>2</sub> production rates increased linearly with increased DOC. This turning point corroborates with earlier observations of primary production rates increasing with increased DOC concentrations until around 5 mg C L<sup>-1</sup>, after which they decline (Karlsson et al., 2007, Seekell et al., 2015, Tanentzap et al., 2017). The DOC concentration-specific CO<sub>2</sub> production, i.e., the rates of CO<sub>2</sub> production per unit of DOC concentration, was positively related to primary production rates. Further, the DOC:TP ratio had a negative effect on primary production, and consequently, the DOC:TP ratio also had a negative effect on the DOC concentration-specific CO<sub>2</sub> production. This implies a more bioavailable DOC pool in productive than in unproductive lakes (Søndergaard et al., 1995) and could also suggest that this is explained by a lower C:P ratio of the substrate.

To further assess the dependency of bacterial respiration on DOC and P, we monitored CO<sub>2</sub> production in incubations of water with a gradient of DOC crossed with two levels of inorganic P. Finally, we crossed DOC and P with a temperature gradient to test the temperature dependency of respiration rates. While the total amount of CO<sub>2</sub> produced during the incubations increased with DOC concentration, P additions had little to say for the cumulative CO<sub>2</sub> production (Figure 5a). On the other hand, respiration rates and bacterial growth yield were higher in P-spiked than in P-limited samples (Figure 5b), suggesting increased bacterial growth and decreased cell-specific respiration under non-limited P conditions. This is in accordance with earlier findings of negative relations between P supply

and cell-specific respiration (Smith and Prairie, 2004, Vikström and Wikner, 2019). DOC concentration hence regulates the overall respiratory output of CO<sub>2</sub> (and consumption of O<sub>2</sub>), while additions of P changes the dynamics by boosting respiration.

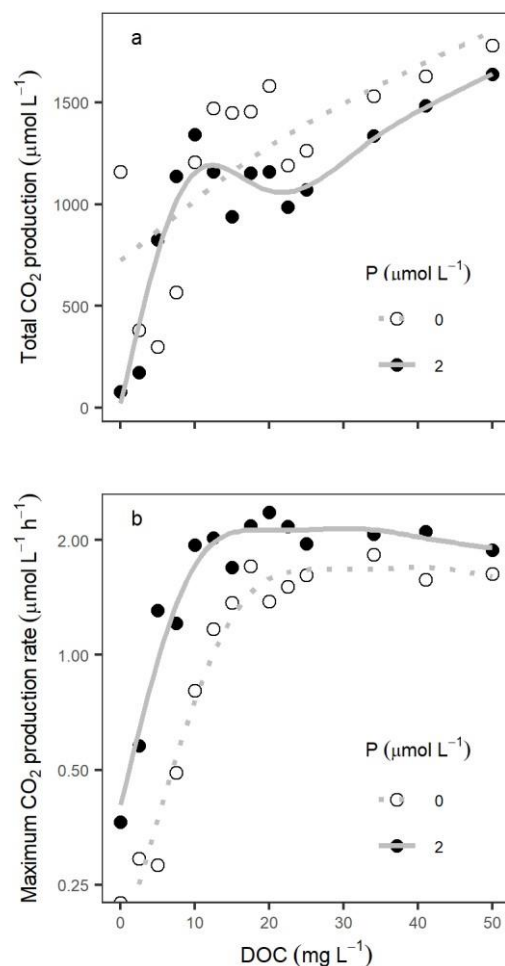


Figure 5. The response of a) total CO<sub>2</sub> production (μmol L<sup>-1</sup>) during the entire incubation (240 h) and b) maximum CO<sub>2</sub> production rates (μmol L<sup>-1</sup> h<sup>-1</sup>) to increased DOC concentrations (mg L<sup>-1</sup>). The solid and dotted lines are fitted gam curves ( $y \sim s(x)$ ) to samples with and without P additions respectively.

Respiration rates showed a sigmoid response to increasing DOC availability reaching a plateau at about 20 mg C L<sup>-1</sup> of initial DOC concentrations. This increase in respiration with DOC was in accordance with the increase in total CO<sub>2</sub> production rates at increased DOC concentration >5 mg L<sup>-1</sup> found in the lake survey. Many boreal lakes have DOC

concentrations below 20 mg L<sup>-1</sup> (e.g. the lake survey of Paper I and II had a maximum value of 12.9 mg C L<sup>-1</sup> and Paper IV had a median value of 7.7 mg C L<sup>-1</sup>) and a continued increase in BR with increased terrestrially derived DOC up to about 20 mg L<sup>-1</sup> could be expected. In addition to these P and DOC level effects, respiration rates responded in a non-monotonic fashion to temperature with an increase in respiration rates by a factor of 2.6 from 15 to 25°C and a decrease at higher temperatures. This further reinforces the idea that net heterotrophy in lakes will increase with increasing temperatures, which also would lead to increased emissions of CO<sub>2</sub> (Sobek et al., 2003). The combined results from the survey and experiments highlight DOC as the major determinant of CO<sub>2</sub> production in boreal lakes, with P and temperature as significant modulators of respiration kinetics.

### *Substrate stoichiometry: oxygen*

The DOC source we used in Paper II was a naturally isolated DOC, added at the beginning of the experiment. In a natural environment, the DOC pool would be constantly refreshed by new additions from the surrounding, not considered in our experiment. Further, recalcitrant DOC would undergo photochemical processing, breaking down high molecular weight humic acids to more bioavailable substrates (Bertilsson and Tranvik, 1998). When available, bacterioplankton preferentially use smaller fractions, easier to assimilate before the more recalcitrant humic fractions (Berggren et al., 2012).

Besides regulating bacterial respiration and growth, substrate composition may also regulate heterotrophic O<sub>2</sub> consumption. Assimilation of organic substrate of high O content and high O:H ratio requires less O<sub>2</sub> from the surroundings (Dilly, 2001). Under such circumstances, respiration is performed at a relatively high respiratory quotient (RQ = unit CO<sub>2</sub> produced per unit O<sub>2</sub> consumed).



High molecular weight humic substrates generally have low O:H ratios and respiration is performed at low RQ, requiring a relatively large amount of O<sub>2</sub> from the surroundings. However, partial oxidation of high molecular weight DOC, may produce compounds with high O content and high O:H ratio (Bertilsson and Tranvik, 1998). Assimilation of such photoproducts may thus be performed at a relatively high RQ (Berggren et al., 2012). Although photooxidation is known to be an important process in natural freshwater systems, its potential role as RQ regulator has to our knowledge never been addressed.

In Paper III, we therefore tested the impact of photochemical processing of DOC on bacterioplankton RQ. We monitored the bacterial RQ in bioassays of both irradiated and non-irradiated humic lake water. As expected, we found bacterioplankton RQ to be significantly higher in irradiated ( $3.4\text{--}3.5 \pm 0.4$ ) than in non-irradiated samples ( $1.3 \pm 0.1$ ). Both CO<sub>2</sub> production and O<sub>2</sub> consumption were higher in irradiated than in non-irradiated samples, indicating increased bacterial activity and a shift towards more labile substrate. The elevated RQ's found in irradiated samples could however not completely be explained by bacterial use of the photoproducts such that there must have been a broader modification of DOC molecules during irradiation. Further, anabolic metabolism is performed at a higher RQ than catabolic respiration alone (Dilly, 2003, Berggren et al., 2012) and bacterial consumption of a more bioavailable photo-altered substrate has been shown to result in enhanced biomass production compared to the more recalcitrant humic substances (Anesio et al., 2005, Amado et al., 2015). Enhanced biomass production in irradiated samples could thus have contributed to the elevated RQ's we observed.

The consistently much higher RQ values in irradiated samples reflect a mechanism that should be present in all light-exposed DOC-rich waters globally and photochemical processes thus have an important role in regulating bacterioplankton RQ.

When assessing BR in aquatic systems, O<sub>2</sub> measurements are often favoured over direct CO<sub>2</sub> measurements as dissolved O<sub>2</sub> can be measured on the aqueous phase, without gas extraction, and without the need to correct for changing equilibria in the inorganic carbon system. An RQ is thus needed as a conversion factor. Many such studies apply a fixed RQ value of 1 (del Giorgio et al., 1997, Koch et al., 2007), which is the theoretical value of complete oxidation of glucose. However, true bacterioplankton RQ can vary substantially (Cimblaris and Kalff, 1998, Berggren et al., 2012, Romero-Kutzner et al., 2015), for example by consumption of photo-altered DOC, as we showed in Paper III. By using the *a priori* assumption of an RQ value of 1 for inland waters, bacterioplankton CO<sub>2</sub> production may thus be greatly underestimated.

## Lake O<sub>2</sub> production

### *Primary Production*

Although enhanced inputs of allochthonous DOC primarily stimulates heterotrophic metabolism, DOM associated nutrients may also benefit autotrophs (Seekell et al., 2015, Tanentzap et al., 2017). Further, heterotrophic mineralization of DOC and the subsequent increase in heterotrophic CO<sub>2</sub> production may indirectly stimulate autotrophs by increasing CO<sub>2</sub> availability (Jansson et al., 2012). An increase in DOC concentrations in clear unproductive waters can therefore stimulate both primary and secondary production. However, as waters turn browner, increased light attenuation may suppress primary production overriding the positive effects of DOM as a nutrient supply (Thrane et al., 2014, Seekell et al., 2015). The relation between increased DOC concentrations and primary production is thus unimodal with a maximum typically around 5-10 mg l<sup>-1</sup> (Seekell et al., 2015). A decrease in primary production propagates through the food web leading to lower overall production, including fish biomass (Karlsson et al., 2009). Finstad et al., (2014)

reported a unimodal response of fish production to increased DOM concentrations, similar to the response of primary production. In corroboration with this unimodality, we saw in Paper II a unimodal response of total CO<sub>2</sub> production to increased DOC concentration in the lakes, suggesting a shift in substrate from mainly autochthonous to predominantly allochthonous DOC.

In Paper I, II, and IV, we found similar drivers of O<sub>2</sub> saturation and estimated areal primary production rates as for CO<sub>2</sub> saturation and CO<sub>2</sub> production rates in boreal lakes. The major regulator of lake O<sub>2</sub> saturation was unsurprisingly the negative effect of DOC concentrations with browner lakes being undersaturated with O<sub>2</sub> while clearer lakes were at or close to O<sub>2</sub> saturation. Further, the DOC:TP ratio correlated negatively with areal primary production rates (Paper II), indicating increasing autotrophic activity with nutrient availability. This, together with the already discussed correlation between CO<sub>2</sub> and O<sub>2</sub> saturation deficits conform the common picture of DOM inputs regulating lake heterotrophy in boreal lakes.

On the contrary, the majority of the arctic lakes in Paper IV were saturated or supersaturated with both CO<sub>2</sub> and O<sub>2</sub> regardless of DOC concentrations. The CO<sub>2</sub> supersaturation in Arctic lakes would, at first thought, indicate net heterotrophy. However, high levels of O<sub>2</sub> saturation together with weak or non-significant influence of DOC concentration on CO<sub>2</sub> saturation rather indicate low respiratory activity in agreement with generally nutrient-poor conditions. High O<sub>2</sub> saturation levels together with a dominance of allochthonous input of CO<sub>2</sub> suggest autotrophy rather than heterotrophy in the arctic lakes studied in Paper IV.

## Closing Remarks

Allochthonous DOC stimulation of heterotrophic metabolism together with the subsequent reduction in primary production at high DOC concentrations make most lakes of high terrestrial influence net heterotrophic (Paper I, II, and IV) (Hessen et al., 1990, Cole et al., 1994, Sobek et al., 2003, Thrane et al., 2014). Lakes comprise only a small fraction of the global land surface (about 4 % (Verpoorter et al., 2014)) but intensive in-lake carbon processing together with inputs of allochthonous CO<sub>2</sub> make greenhouse gas emissions from lakes equivalent to up to 20 % of the global CO<sub>2</sub> emissions from fossil fuel combustion (DelSontro et al., 2018). Recovery from acidification, increased vegetation cover in catchments, land use change, and a wetter climate all together promote carbon export to lakes (Finstad et al., 2017, de Wit et al., 2018, Škerlep et al., 2020) contributing to enhanced browning (Kritzberg et al., 2020).

Bacterial metabolism depends on substrate quantity and quality, nutrient availability, and temperature (Farrell and Rose, 1967, Hessen et al., 1994, Smith and Prairie, 2004). Productive lakes have a more bioavailable DOC pool with lower C:P ratio, promoting bacterial growth and yielding high bacterial numbers with low per-cell respiration (Paper II) (Søndergaard et al., 1995). On the other hand, in unproductive lakes where the DOC pool is dominated by allochthonous DOC with high C:P ratio, fewer cells may be present but cell-specific respiration is elevated (Vikström and Wikner, 2019). The overall respiratory output of CO<sub>2</sub> is thus primarily regulated by DOC concentrations (Paper II).

While heterotrophic respiration is the dominant DOC mineralization process, photochemistry may have a significant role in processing DOC, both via photomineralization and via partially degrading DOC to smaller compounds (Granéli et al., 1996, Bertilsson and Tranvik, 2000, Cory et al., 2014). However, sunlight attenuates fast in the water column making photo-reactive wavelengths absorbed within the top few meters also in clear lakes

(Paper I). Photochemical processes will thus likely not increase as lakes turn browner but rather move upwards and take place closer to the surface. Decreasing primary production and increasing bacterial respiration due to increasing export of humic-rich DOC will thus enhance net heterotrophy (Paper I, II, and IV). The relative contribution of photomineralization to CO<sub>2</sub> evasion will most probably decline in a changing climate (Paper I).

Besides DOC mineralization and in-lake processing, DIC may also enter lakes via surface- and groundwater flow (Ojala et al., 2011, Vachon and Del Giorgio, 2014). Whether in-lake production or inputs of allochthonous CO<sub>2</sub> dominate as a cause of lake CO<sub>2</sub> saturation is dependent on the catchments characteristics with net heterotrophy tightly coupled to DOC inputs and coniferous forest coverage (Puts et al., 2022). Coniferous forests thus have a fundamental role, governing the organic and inorganic dynamics in lakes (Paper IV).

In the ongoing climate crisis, we can expect a decrease in lake productivity with decreased O<sub>2</sub> levels, having consequences throughout the entire food web. In a forest ecosystem with high lake abundance, including the evasion from lakes in the CO<sub>2</sub> budget of the forest lowers the effect of the vegetation as a net sink, with the potential to shift to a net source on the ecosystem level in a changing climate.

## Selected Methods

### Paper I: Estimating photochemical DIC production

In Paper I, we estimated the role of photochemical DOC mineralization for lake CO<sub>2</sub> emissions in 70 boreal lakes along a gradient in DOC concentrations. Simulation of photochemical mineralization of DOC requires knowledge of the photochemical reactivity of the DOC (apparent quantum yield; AQY, defined as moles photochemically produced DIC per mole photons absorbed by the DOC pool) across the whole spectrum of photochemically active wavelengths. The lakes from the data set we used for Paper I, was samples 2011 and did not include AQY measurements. However, Koehler et al. (2016) found positive linear correlations between AQY and the absorption coefficient at 420 nm ( $a_{420}$ ) and with specific UV absorption coefficients (SUVA<sub>254</sub> and SUVA<sub>400</sub>). These correlations open up for possibilities to improve large-scale model estimates of photomineralization in inland waters based on water colour information.

For the lakes in Paper I, we only had optical data for wavelengths in the PAR band, and even though the correlation between AQY and SUVA 254 was stronger than between AQY and SUVA<sub>400</sub>, we instead used the relation between AQY and SUVA<sub>400</sub> (B. Koehler, unpublished data, 2016). We then used the lme4 package in R (Bates and al., 2014) for a linear mixed effects model with the measured AQY as response variable,  $a_{420}$ , SUVA<sub>400</sub>, and wavelength as fixed effects, and intercept as a random effect.

In order to find whether we lost any using SUVA<sub>400</sub> instead of SUVA<sub>254</sub>, we used the AQY model on data from the lakes in Koehler et al. (2016) using SUVA<sub>254</sub> and  $a_{420}$  and compared it to the model with SUVA<sub>400</sub> and  $a_{420}$ . The models resulted in close to exactly the same AQY spectra (Figure 6). Hence, we did not seem to lose any information by using extrapolated PAR spectra to predict the AQY spectra for the 70 study lakes.

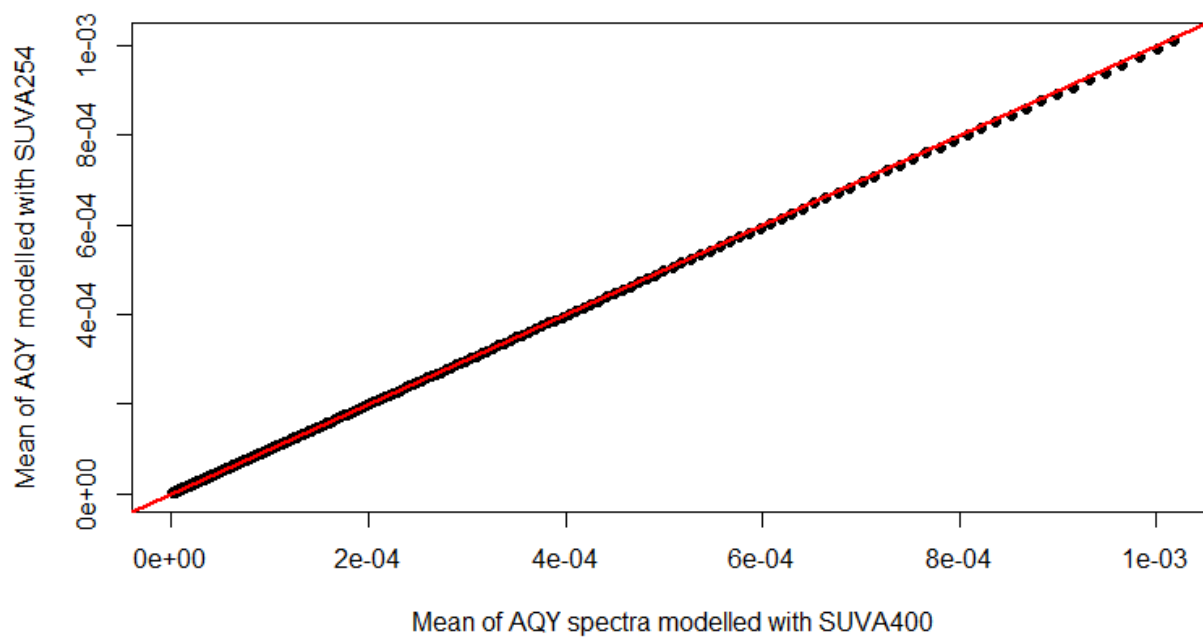


Figure 6. The mean values of each wavelength of the Monte Carlo samples in the AQY model using  $SUVA_{254}$  and  $a_{420}$  versus the AQY model using  $SUVA_{400}$  and  $a_{420}$ . The red line is the 1:1 line.

We then also modelled lake absorption spectra for the different compounds in the lake samples (Twardowski et al., 2004; Shen et al., 2012; Wozniak and Dera, 2007; Kirk, 1994) and calculated the relative contribution of DOC to total light absorption (Figure 7).

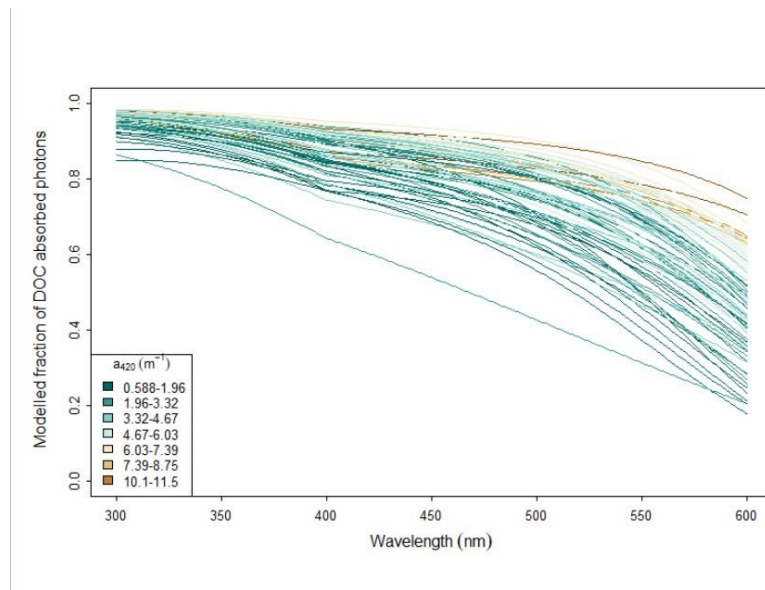


Figure 7. At short wavelengths the fraction of the photons absorbed by DOM is close to 1. The fraction decreases at longer wavelengths, where photochemical reactivity is lower. The color gradient goes from dark blue for clearer lakes with low  $a_{420}$  to brown for darker lakes with high  $a_{420}$ .

Finally, combining the AQY spectra with the absorption spectra and an irradiation model of incoming photon flux, we could calculate the wavelength-specific photoproduction of DIC per depth or per area unit. All absorption spectra were extrapolated from the measured PAR band to 300 nm using linear mixed effect models with prediction uncertainties propagated through Monte Carlo samples generated by the arm package in R (Gelman et al., 2018).

## Paper I and II: Total CO<sub>2</sub> production in the lakes

For Papers I and II, we wanted an estimate of the CO<sub>2</sub> production in lakes. The data set we used included lake CO<sub>2</sub> concentrations. With these concentrations together with wind speed data (the Norwegian Reanalysis Archive (Furevik and Haakenstad, 2012)) we



calculated water-air flux of CO<sub>2</sub> using Henry's law to obtain the CO<sub>2</sub> deficit, and Fick's law of diffusion to obtain the net degassing from the surface. We also obtained necessary coefficients for the calculations according to Jähne et al (1987), Wanninkhof (1992), and Cole and Caraco (1998).

Further, in an earlier study of the same data set, Thrane et al (2014) calculated primary production rates in the lakes. Here, primary production is a measure of CO<sub>2</sub> consumption representing the CO<sub>2</sub> flux from water to primary producers.

The measured parameters included in the dataset did not allow for distinguishing between lateral input of CO<sub>2</sub> via surface- and ground water, and in-lake production of CO<sub>2</sub>. We therefore used the sum of in-lake DOC mineralization and lateral input as an estimate of total CO<sub>2</sub> production. Assuming a steady state of the CO<sub>2</sub> saturation deficit, such that all production and input of CO<sub>2</sub> either evades from the surface or is consumed by autotrophs, we can estimate the mass balance due to production, lateral input, consumption, and evasion as the sum of CO<sub>2</sub> fluxes water-air and water-primary producers.

This is indeed a rather coarse estimate of CO<sub>2</sub> production and does not give any exact values. However, it is useful and fits for our main purpose of finding the relative contribution of photomineralization to CO<sub>2</sub> production and evasion and whether this depends on the level of brownification of the lakes.

## Paper II: Experimental determination of BR

To test for dynamic responses of bacterioplankton respiration to allochthonous DOC concentrations, nutrient availability, and temperature, we additionally performed experimental incubations. During 1-week incubations, we monitored respiration in two experimental set-ups, one addressing CO<sub>2</sub> production and the other O<sub>2</sub> consumption. A gradient of DOC concentrations was achieved by adding natural organic matter (NOM; isolates from a Norwegian humic lake obtained through reverse osmosis(see Gjessing (1999) and Vogt et al. (2001)) to clear lake water.

In the first experimental set-up, we monitored CO<sub>2</sub> concentrations by gas chromatography every 6 hours using the robotized setup described by Molstad et al. (2016) with some modifications. Since the vials contained 50 ml water and 70 ml air, the O<sub>2</sub> uptake was small relative to the large amount of O<sub>2</sub> in the headspace such that we could not measure uptake rates with sufficient precision.

In this set-up, we used 14 levels of DOC additions between 0 and 50 mg L<sup>-1</sup>, crossing it with two levels of PO<sub>4</sub>-P additions (0 and 2 μmol L<sup>-1</sup>). To make sure that N was not limiting, we added 30 μmol L<sup>-1</sup> each of NO<sub>3</sub>-N and NH<sub>4</sub>-N to all samples. The C:N:P ratio ranged between 82:30:1 and 2160:30:1 in samples with P additions.

At the end of the experiment, we aimed to assess biomass measurements using flowcytometry. However, the samples were obscured by background scatter from the added DOC and did not provide reliable counts. Instead, we assessed final microbial biomass from qPCR on filtered samples when terminating the experiment. We approximated bacterial growth yields we measured bacterial 16S rRNA gene copy numbers using a quantitative polymerase chain reaction (qPCR) protocol (Savio et al., 2015).

The main goal of the experiment was to assess BR and although it would have been ideal to verify the dynamic responses in CO<sub>2</sub> with bacterial counts and community response

by genetic screening and transcriptomic, this would have required a different set-up with larger volumes and more frequent sampling. The current set-up allowed us to prioritize the gas analysis while also assessing the final bacterial biomass in each incubation unit.

In the second experimental set-up, we used fewer DOC levels (0 mg L<sup>-1</sup>, 25 mg L<sup>-1</sup>, and 50 mg L<sup>-1</sup>) but added an additional crossing with 4 temperatures (10 °C, 15 °C, 25 °C, and 30 °C). Concentration of dissolved oxygen was recorded automatically every 15 s using a SensorDish Reader (PreSens GmbH, Regensburg, Germany).

### Paper III: Experimental determination of bacterioplankton respiratory quotient

In Paper III, we tested the impact of UV radiation on bacterioplankton RQ. To do so, we conducted incubations of both irradiated and non-irradiated samples of natural lake water from four humic-rich lakes in northern Sweden with catchments dominated by coniferous forest, simultaneously measuring changes in O<sub>2</sub> and CO<sub>2</sub>.

We performed the experiment in three different parts. i) *Irradiated*: where we first irradiated the samples during 48 h and then incubated them for 48 h in the dark. ii) *Biological + irradiated*: where we first incubated the samples in the dark for 48 h followed by 48 h of UV-treatment and then finally another 48 h of biological incubation. iii) *Dark control*: where we incubated the samples in the dark for 144 h. In part *iii* we chose the longer incubation time to mirror the time of the second treatment, controlling that a possible change in RQ was due to bacterial consumption of photo-chemical processed DOC and not incubation time.

In this study, we measured CO<sub>2</sub> production through changes in pH (decreasing as CO<sub>2</sub> is produced) in the water samples. Depending on the alkalinity and pH in the system, a certain share of the respiratory CO<sub>2</sub> dissociates in the water, forming bicarbonate and carbonate ions (Stumm and Morgan, 1996). To cover the total CO<sub>2</sub> (TCO<sub>2</sub> = CO<sub>2</sub> + HCO<sub>3</sub><sup>-</sup> +

CO<sub>3</sub><sup>2-</sup>) production during BR measurements, the impact of the carbonate system on CO<sub>2</sub> hence needs to be corrected for.

Finally, to confirm that the photochemical processing of the samples contributed to increased concentrations of low molecular weight organic acids, we measured concentrations of common low molecular weight organic acids, before and after UV-treatment. To do so, we used a liquid ion chromatography-ionspray tandem mass spectrometry system (IC-MS). The method is described in further detail in Ström et al. (2012).

## Paper I, II, and IV: Coupling lake CO<sub>2</sub> and O<sub>2</sub> to environmental variables

In Papers I, II, and IV, we analyzed environmental drivers of lake CO<sub>2</sub> and O<sub>2</sub> concentrations and saturation deficits. Besides *in situ* measurements of pH, temperature, and electrical conductivity, samples from all lakes were also analyzed total organic carbon, dissolved organic carbon, total nitrogen, total phosphorus. We analyzed dissolved gas concentration using the acidified headspace method (Åberg and Wallin, 2014). We then calculated the level of saturation of each gas relative to equilibrium with ambient air from measured concentrations using Henry's law with temperature-dependent solubility constants.

We used the open-source software R version 3.4.1 (Team, 2017) for all data analysis. For the statistical modelling in Paper II and IV, we used the `mgcv` package (Wood, 2011) fitting gam models for prediction of the dependent variables. To test dependency, we used a gam model with smoothers on each of the explanatory variables. Predictive variable selection was done by applying additional shrinkage on the null space of the penalty with the `select=TRUE` argument in the `mgcv::gam` function, as recommended by Marra and Wood (2011).

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# Paper I



## The role of photomineralization for CO<sub>2</sub> emissions in boreal lakes along a gradient of dissolved organic matter

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### Abstract

Many boreal lakes are experiencing an increase in concentrations of terrestrially derived dissolved organic matter (DOM)—a process commonly labeled “browning.” Browning affects microbial and photochemical mineralization of DOM, and causes increased light attenuation and hence reduced photosynthesis. Consequently, browning regulates lake heterotrophy and net CO<sub>2</sub>-efflux to the atmosphere. Climate and environmental change makes ecological forecasting and global carbon cycle modeling increasingly important. A proper understanding of the magnitude and relative contribution from CO<sub>2</sub>-generating processes for lakes ranging in dissolved organic carbon (DOC) concentrations is therefore crucial for constraining models and forecasts. Here, we aim to study the relative contribution of photomineralization to total CO<sub>2</sub> production in 70 Scandinavian lakes along an ecosystem gradient of DOC concentration. We combined spectral data from the lakes with regression estimates between optical parameters and wavelength specific photochemical reactivity to estimate rates of photochemical DOC mineralization. Further, we estimated total in-lake CO<sub>2</sub>-production and efflux from lake chemical and physical data. Photochemical mineralization corresponded on average to 9% ± 1% of the total CO<sub>2</sub>-evasion, with the highest contribution in clear lakes. The calculated relative contribution of photochemical mineralization to total in-lake CO<sub>2</sub>-production was about 3% ± 0.2% in all lakes. Although lakes differed substantially in color, depth-integrated photomineralization estimates were similar in all lakes, regardless of DOC concentrations. DOC concentrations were positively related to CO<sub>2</sub>-efflux and total in-lake CO<sub>2</sub>-production but negatively related to primary production. We conclude that enhanced rates of photochemical mineralization will be a minor contributor to increased heterotrophy under increased browning.

Most lakes worldwide are supersaturated with carbon dioxide (CO<sub>2</sub>), emitting 0.32–0.53 Pg CO<sub>2</sub>-C yr<sup>-1</sup> to the atmosphere on a global scale (Cole et al. 2007; Raymond et al. 2013). A major part of the CO<sub>2</sub> emitted from lakes is produced through mineralization of dissolved organic matter (DOM) (Vachon et al. 2016). DOM in freshwaters originates both from in situ primary production and from the surrounding terrestrial ecosystems, with a general dominance of the latter (Karlsson et al. 2009). Terrestrially derived DOM consists primarily of high molecular weight humic substances. These substances make the majority of the dissolved organic carbon (DOC) pool in most lakes and thus we will primarily refer to

DOC, hence using C as a common currency, through the following text. DOC poses a multitude of partly contrasting impacts on the physical and chemical properties of water, as well as on the biota (Hessen and Tranvik 1998).

Humic substances are a major source of energy to heterotrophs in aquatic ecosystems with high terrestrial influence and the subsequent increase in heterotrophic CO<sub>2</sub> production may indirectly stimulate autotrophs. Nutrients associated with DOC may stimulate both heterotrophic and autotrophic productivity in nutrient-poor regions. Further, humic substances are often highly aromatic and can protect aquatic organisms from harmful UV-radiation (Dillon and Molot 2005; Kritzberg and Ekström 2011). On the other hand, at a certain threshold concentration (possibly around 5 mg l<sup>-1</sup>; Seekell et al. 2015), terrestrial DOC may shift from acting as a nutrient subsidy to suppressing primary production due to light attenuation (Thrane et al. 2014; Seekell et al. 2015). Lakes with high inputs of terrestrial DOC are often to a larger degree supersaturated with CO<sub>2</sub> than lakes where the major part of the DOC pool originates from in-lake production (Cole et al. 2000; Larsen et al. 2011a).

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Besides being an essential source of energy for bacterioplankton (Hessen 1992), this terrigenous DOC is highly chromophoric and photo-reactive, especially in the UV waveband (Lindell et al. 1995). Photomineralization of DOC to dissolved inorganic carbon (DIC) might therefore be a significant part of the DIC production and carbon cycling in humic lakes, adding to the high respiratory activity of heterotrophic prokaryotes and low autotrophic CO<sub>2</sub>-fixation. The annual photochemical mineralization has been estimated to account for 9–12% of the total lake CO<sub>2</sub> emission in the boreal biome, and amount to 13–35 Tg C yr<sup>-1</sup> from inland waters worldwide (Koehler et al. 2014). However, the relative contribution of photochemical mineralization to in-lake carbon cycling varies significantly both between systems (Granéli et al. 1996; Molot and Dillon 1997; Cory et al. 2014) and temporally within the same system (Groeneveld et al. 2016; Vachon et al. 2016).

In order to simulate photochemical mineralization, knowledge of the reactivity across the whole spectrum of photochemically active wavelengths is needed. This photochemical reactivity or apparent quantum yield (AQY) of DIC photoproduction is defined as moles photochemically produced DIC per mole photons absorbed by the DOC pool (Miller et al. 2002). Besides the quantity of DOC, studies have found photochemical DIC production rates to be dependent on its quality, as well as on water chemistry, such as pH and iron concentration (Lindell et al. 1995; Bertilsson and Tranvik 2000; Panneer Selvam et al. 2019) while other studies have found no such relationships (Cory et al. 2014). A significant share of the AQY variability between lakes can be explained by simple optical parameters (Koehler et al. 2016), allowing for estimates of photochemical DIC production when system-specific AQY spectra are not available.

In this study, we used data of such optical parameters from 70 Scandinavian lakes along a gradient of DOC concentrations, together with correlation estimates between the absorption coefficient at 420 nm ( $a_{420}$ ) and the specific UV absorption coefficient at 400 nm (SUVA<sub>400</sub>) and the AQY (Koehler et al. 2016) to estimate the lakes' wavelength specific AQY spectra. Together with atmospheric radiative modeling, we then simulated the photochemical DIC production in the study lakes. We further estimated the lakes' primary production using lake-specific phytoplankton absorption coefficients and in situ irradiance. Finally, we calculated the air-water CO<sub>2</sub> flux through surface water CO<sub>2</sub> concentrations, temperature and wind speed, using Fick's law of diffusion and Henry's law to find the CO<sub>2</sub> deficit from concentrations at equilibrium with the atmosphere. Assuming that the deviation of CO<sub>2</sub> from saturation is kept at steady state due to production, lateral input, evasion, and consumption we estimated the sum of total lake CO<sub>2</sub> production and the lateral input as the sum of the consumption and evasion. This allowed us to calculate the relative contribution of photochemical DIC production to lake carbon cycling. As short-wave radiation attenuates quickly in the water column of lakes, we expect all incoming photochemically reactive photons to be

absorbed within the top few meters of all lakes, even the clear ones. Therefore, we hypothesized that the total amount of photomineralization of DOC would be similar in all lakes regardless of their CDOM concentrations.

## Methods

### Study sites

During July and August of 2011, 77 lakes along a geographical gradient between western Norway and eastern Sweden were sampled (Fig. 1). The lakes were chosen to represent gradients in DOC and total phosphorus (TP), aiming for an orthogonal gradient between these parameters, and to avoid strong temperature gradients with respect to latitude and altitude. All lakes met the following criteria: latitude 57–64°N, altitude < 600 m, surface area > 1 km<sup>2</sup>, pH > 5, TP < 30 µg l<sup>-1</sup>, and DOC < 30 mg l<sup>-1</sup>.

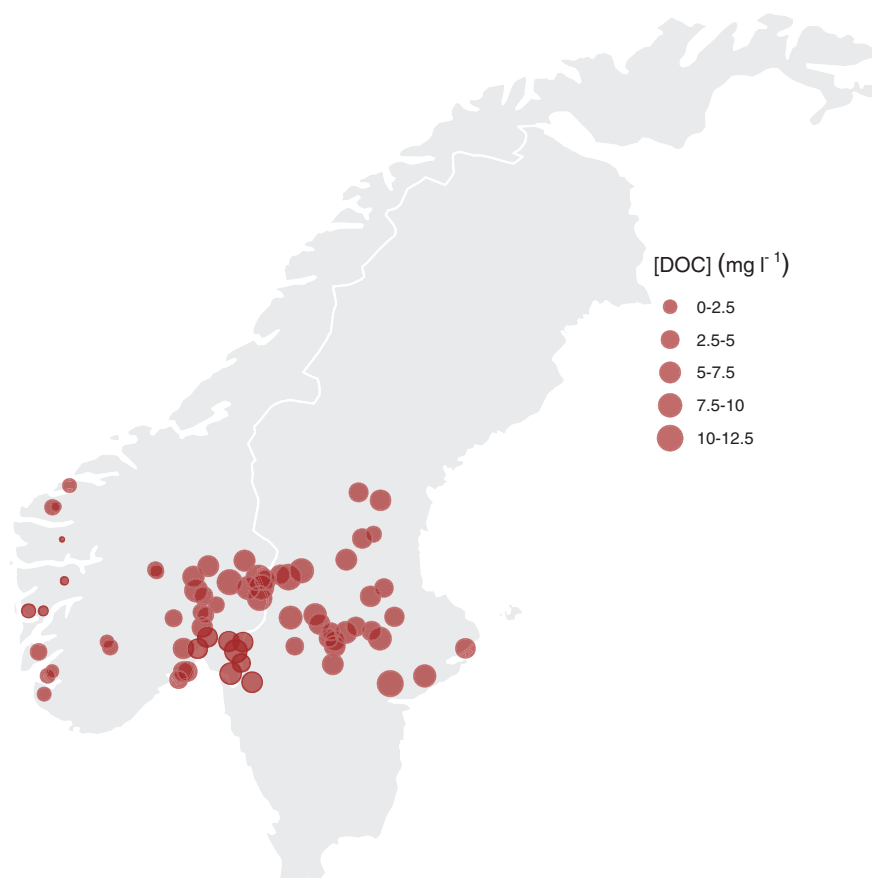
### Field sampling

Composite samples (15 L in total) were taken from 0 to 5 m in the central part of each lake during daytime, using an integrating water sampler (Hydro-BIOS, Germany). Water temperatures were measured using XRX-620 10-channel CTD (RBR Ltd., Canada). Vertical temperature profiles indicated that the thermocline was deeper than 5 m in all lakes (Fig. S1) and the integrated 0–5 m samples could be considered representative of the entire mixed layer of the lakes. Vertical profiles of scalar irradiance in the photosynthetically active radiation (PAR) region (400–700 nm;  $E_d$ ) were measured using a spherical irradiance sensor (BioSpherical instruments) attached to a 10 channel CTD profiler (WRW620, RBR Ltd., Canada). The sensor was lowered at a rate of approximately 20 cm s<sup>-1</sup> with a sampling rate of 6 Hz. The vertical attenuation coefficient for scalar PAR ( $K_d$ PAR) was estimated by taking the median of the distribution of slopes obtained from regressing natural log-transformed  $E_d$  against depth ( $z$ ) for each 10 sampling points (i.e., sliding windows). This was done to correct for temporal changes in irradiance caused by for example wave action and clouds during the haul. pH in the samples was measured within 1 h after sampling using a handheld pH-meter (PHM201, Radiometer Analytical, France).

### Laboratory analyses

Concentrations of total phosphorus (TP), total organic carbon (TOC), and total nitrogen (TN) were measured in two accredited laboratories, at the Norwegian Institute for Water Research (NIVA) and at the University of Oslo (UiO). Differences between laboratories were small for TOC and TN but slightly higher for TP. Regressions of UiO vs. NIVA measurements had the following statistics: TP:  $R^2 = 0.77$ , residual standard error ( $RSE$ ) = 2.27 µg l<sup>-1</sup>; TOC:  $R^2 = 0.99$ ,  $RSE = 0.25$  mg l<sup>-1</sup>; TN:  $R^2 = 0.91$ ,  $RSE = 81$  µg l<sup>-1</sup>. There were no systematic differences between the laboratories and the averages of the results were used in the subsequent analysis. DOC was calculated as the difference between the total organic carbon (TOC) and particulate organic carbon (POC). TOC was





**Fig. 1.** Lakes included in the survey. The sizes of the symbols scale with the concentration of dissolved organic carbon (DOC; mg l<sup>-1</sup>).

measured by infrared CO<sub>2</sub> detection after catalytic high temperature combustion (Shimadzu TOC-VWP analyzer (UiO), or Phoenix 8000 TOC-TC analyzer (NIVA)). On average, >95% of the TOC was in dissolved form (DOC). POC was measured on an elemental analyzer (Flash EA 1112 NC, Thermo Fisher Scientific, Waltham, Massachusetts) through rapid combustion in pure oxygen of a pre-combusted GF/C-filter with particulates. TP was measured on an auto-analyzer as phosphate after wet oxidation with peroxodisulfate in both laboratories. TN was measured on unfiltered samples by detecting nitrogen monoxide by chemiluminescence using a TNM-1 unit attached to the Shimadzu TOC-VWP analyzer (UiO), or detection of nitrate after wet oxidation with peroxodisulfate in a segmented flow auto-analyzer (NIVA). Concentrations of CO<sub>2</sub> and O<sub>2</sub> were determined by automated gas chromatography (GC) analysis with back-flushing H<sub>2</sub>O (see Yang et al. 2015 for details). Total iron (Fe) was measured using an inductively coupled plasma mass spectrometer (ICP-MS, PerkinElmer NexION 300, Norwalk, Connecticut) equipped with three quadrupole mass analyzers, a cyclonic spray chamber, and a concentric nebulizer. Three subsamples from each lake were measured to evaluate the analytical precision.

For measurements of particulate absorbance spectra, water samples (150–170 mL, depending on particle load) were filtered onto 25 nm Whatman GF/C glass filters under low vacuum. The filters were placed in the entrance of an integrating sphere (ISR 2200, Shimadzu scientific instruments, Columbia, Maryland) attached to a double beam Shimadzu UV-2550 spectrophotometer, and optical density was measured for each nm from 400 to 800 nm. After the first measurement, the sample filters were bleached with sodium hypochlorite (Tassan and Ferrari 1995). The bleaching oxidizes all pigments, leaving only organic and inorganic detritus, including de-pigmented algal remains, unbleached. The optical density of this nonalgal particulate (NAP) matter was then measured and the absorption coefficients (m<sup>-1</sup>) of total particulate matter and nonalgal particulate matter were calculated according to Mitchell et al. (2002), using the algorithm of Bricaud and Stramski (1990) to estimate the path-length amplification factor ( $\beta$ ). Finally, the absorption coefficient spectra of phytoplankton pigments were calculated as the difference between the total particulate and the NAP absorption coefficient spectra. DOC absorbance spectra from 400 to 700 nm (1 nm resolution) were measured in 0.2  $\mu$ m filtered water samples

(Acrodisc 0.2  $\mu\text{m}$  polyethersulfone membrane syringe filter, Pall Life Sciences, Port Washington, NY) using a 50 mm quartz cuvette. Absorption coefficient spectra were calculated according to Mitchell et al. (2002). Due to missing values of some of the absorbance measurements, seven lakes had to be omitted, giving a data set of 70 lakes for further analysis.

### Primary production calculations

Area-specific primary production ( $\text{PP}_A$ ;  $\text{mg C m}^{-2} \text{d}^{-2}$ ) was calculated using a bio-optical model based on lake-specific phytoplankton absorption coefficients, in situ irradiance, and the light dependent quantum yield of photosystem II measured by a Pulse Amplitude Modulated (PAM) fluorometer (AquaPen, PSI Czech Republic). In brief, this bio-optical model is based on estimating the in vivo rate of light absorption by phytoplankton, and subsequently electron transport rates (ETRs) through photosystem II (PSII) using information about the light-dependent quantum yield of photochemistry in PSII. ETR can further be converted to a rate of gross carbon fixation by assuming an appropriate value for the quantum yield of  $\text{CO}_2$  fixation (Kromkamp and Forster 2003; Suggett et al. 2010). While the method could be sensitive to phytoplankton community composition related to their pigments and light capturing properties, it has gained increased interest over the last two decades because it offers a fast and inexpensive way of obtaining PP estimates (see Thrane et al. 2014 for details). A comparison of this method and empirical estimates for PP in boreal lakes demonstrates a good accordance (Thrane et al. 2014). The method is thus a feasible tool for assessment of primary production across a large number of sites. It also avoids many of the pitfalls of  $^{14}\text{C}$ -bottle incubation, which in any case could not have been applied in this kind of synoptic, snapshot survey with sampling from a plane spanning many lakes over a large geographical area.

### Wavelength-specific AQY spectrum

Koehler et al. (2016) found the strongest predictors of AQY to be the Napierian absorption coefficient at 420 nm ( $a_{420}$ ;  $\text{m}^{-1}$ ) and specific UV absorption coefficient at 254 nm ( $\text{SUVA}_{254}$ ;  $\text{L mg C}^{-1} \text{m}^{-1}$ ) (Kirk 1994). The data set in this study only contained optical data for wavelengths in the PAR band and therefore the relation between AQY and  $\text{SUVA}_{400}$  (B. Koehler, unpublished data, 2016) (Table S2) was used instead of  $\text{SUVA}_{254}$ .

A linear mixed effects model with the measured AQY as the response variable,  $a_{420}$ ,  $\text{SUVA}_{400}$ , and wavelength as fixed effects, and intercept as a random effect was run for the lakes in Koehler et al. (2016) using the lme4 package in R (Bates et al. 2014).

$$\ln(\Phi) \sim a_{420} + \text{SUVA}_{400} + \lambda + (1|\text{lake}) \quad (1)$$

Where  $\Phi$  is AQY for DIC photoproduction,  $\lambda$  is the wavelengths in the measured wavelength region (400–700 nm in

steps of 1 nm), and the (1|lake) term captures other between-lake variations not related to chromophoric DOM (CDOM) quality. The Napierian absorption coefficient at 420 nm ( $a_{420}$ ) is a proxy for CDOM content, such that the higher the  $a_{420}$ , the browner the lake. We used the AQY model on data from the lakes in Koehler et al. (2016) using  $\text{SUVA}_{254}$  and  $a_{420}$  and compared it to the model with  $\text{SUVA}_{400}$  and  $a_{420}$ . The models resulted in close to exactly the same AQY spectra (Fig. S4) and hence we did not lose information modeling the AQY from  $\text{SUVA}_{400}$  instead of  $\text{SUVA}_{254}$ . The model was then used to predict the AQY spectra for the 70 study lakes.

The arm package in R (Gelman et al. 2018) was used to generate Monte Carlo samples of fixed effect parameters of the linear model, which was used to propagate model uncertainties to the estimated lake specific AQYs over the entire spectrum (300–600 nm; Figs. S2 and S3). AQY spectra were extrapolated to wavelengths  $< 400$  nm using the exponential model (Eq. 1). The irradiation model included wavelengths between 300 and 600 nm and therefore the AQY spectra were also cut at 600 nm.

### Irradiation model

Daily integrated downwelling scalar irradiation spectra (300–600 nm) just below the water surface were obtained using the libRadtran model (version 1.6) for radiative transfer (Mayer and Kylling 2005), parameterized and cloud corrected as described in Koehler et al. (2014). The clear-sky spectra were integrated with calculated solar zenith angles and measurements of ozone column fields in hourly time steps at the coordinates of each lake. The true solar zenith angle was calculated with hourly time step for each lake and day for a month between early July and early August of 2011 (i.e., the time period of field sampling), using approximations in the Astronomical Almanac (Michalsky 1988). The actual ozone column fields for the same time were extracted from the archive operational runs of the Integrated Forecasting System at the European Centre for Medium-Range Weather Forecasts (<http://www.ecmwf.int/research/ifsdocs/CY33r1/index.html>). To correct for attenuation by clouds, total cloud cover data were retrieved for the requested time period at the lakes coordinates from the archive of the operational mesoscale analysis system at the Swedish Meteorological and Hydrological Institute (Häggmark et al. 2000).

### Photochemical DIC production in the lakes

According to the photon budget approach (Kirk 1994), absorption spectra for the lakes were modeled for DOC ( $a_{\text{DOC}}[\lambda]$ ;  $\text{m}^{-1}$ ) (Twardowski et al. 2004), nonalgal particles ( $a_{\text{NAP}}[\lambda]$ ;  $\text{m}^{-1}$ ) (Shen et al. 2012), phytoplankton ( $a_{\text{PP}}[\lambda]$ ;  $\text{m}^{-1}$ ), all from lake samples and for standardized water ( $a_{\text{water}}[\lambda]$ ;  $\text{m}^{-1}$ ) (Wozniak and Dera 2007). All absorption spectra were extrapolated from the measured PAR band to 300 nm using linear mixed effect models with prediction uncertainties

propagated through Monte Carlo samples generated by the arm package in R (Gelman et al. 2018).

The total absorption coefficient spectrum ( $a_{\text{total}}[\lambda]$ ;  $\text{m}^{-1}$ ) was calculated as the sum of  $a_{\text{DOC}}(\lambda)$ ,  $a_{\text{NAP}}(\lambda)$ ,  $a_{\text{water}}(\lambda)$ , and  $a_{\text{PP}}(\lambda)$  (Kirk 1994) and the relative contribution of DOC to the total absorption ( $k_{\text{DOC}}(\lambda)$ ) was calculated as the  $a_{\text{DOC}}(\lambda)$  to  $a_{\text{total}}(\lambda)$  quotient (Fig. S5). Finally, the wavelength-specific photon absorption by DOC per depth unit ( $E_{\text{abs},p}[\lambda, z]$ ;  $\text{mol m}^{-3} \text{d}^{-1} \text{nm}^{-1}$ ) was calculated as the depth derivative of the attenuation profile, weighted by the relative DOC contribution:

$$E_{\text{abs},p}(\lambda, z) = E_p(\lambda) e^{-a_{\text{total}}(\lambda)z} a_{\text{DOC}}(\lambda) \quad (2)$$

where  $E_p$  is the photon flux ( $E_p(\lambda)$ ;  $\text{mol m}^{-2} \text{d}^{-1} \text{nm}^{-1}$ ) at the lake surface from the modeled irradiation spectra and  $z$  is depth (m). Solving Eq. (2) for  $z \rightarrow 0$ , i.e., just below the surface, the DOC absorbed photons per unit volume is given by:

$$E_{\text{abs},p}(\lambda, 0) = E_p(\lambda) a_{\text{DOC}}(\lambda) \quad (3)$$

Boreal lakes generally absorb all incoming irradiation (Kirk 1994; Koehler et al. 2014; Thrane et al. 2014). Assuming that this also is the case for the lakes in this study, integrating Eq. (3) over the entire water column ( $\int_0^\infty E_{\text{abs},p}(\lambda, z) dz$ ), DOC absorbed photons per unit surface area ( $E_{\text{abs},p}(\lambda)$ ;  $\text{mol m}^{-2} \text{d}^{-1} \text{nm}^{-1}$ ) is given by:

$$E_{\text{abs},p}(\lambda) = E_p(\lambda) k_{\text{DOC}}(\lambda) \quad (4)$$

Wavelength-specific photochemical DIC production could then be calculated as either volumetric rates at the surface ( $\psi_{\text{DIC}}[\lambda, 0]$ ;  $\text{mol m}^{-3} \text{d}^{-2} \text{nm}^{-1}$ ) or as production rates per unit area ( $\psi_{\text{DIC}}(\lambda)$ ;  $\text{mol m}^{-2} \text{d}^{-2} \text{nm}^{-1}$ ), multiplying the photon absorption by DOC by the AQY ( $\Phi$ ):

$$\psi_{\text{DIC}}(\lambda, z) = E_{\text{abs},p}(\lambda, z) \Phi_{\text{DIC}}(\lambda) \quad (5)$$

### CO<sub>2</sub> flux

Air-water flux of CO<sub>2</sub> ( $F_{\text{CO}_2}$ ;  $\text{mmol m}^{-2} \text{d}^{-1}$ ) was calculated from the surface CO<sub>2</sub> concentrations in each lake using Fick's law of diffusion:

$$F_{\text{CO}_2} = k_{\text{CO}_2} \Delta_{\text{CO}_2} \quad (6)$$

where  $k_{\text{CO}_2}$  ( $\text{m d}^{-1}$ ) is the CO<sub>2</sub> gas exchange coefficient at a given temperature and  $\Delta_{\text{CO}_2}$  ( $\text{mmol m}^{-3}$ ) is the CO<sub>2</sub> deficit from concentrations at equilibrium with the atmosphere, obtained using Henry's law.  $k_{\text{CO}_2}$  was estimated for each lake using the gas transfer velocity ( $\text{cm h}^{-1}$ ) for a gas-temperature combination with a Schmidt number of 600 ( $k_{600}$ ; CO<sub>2</sub> at 20°C) according to Jähne et al. (1987):

$$k_{\text{CO}_2} = k_{600} \left( \frac{Sc_{\text{CO}_2}}{600} \right)^{-x} \quad (7)$$

where  $x = 2/3$  if wind speed  $\leq 3 \text{ ms}^{-1}$  and  $x = 0.5$  if wind speed  $> 3 \text{ ms}^{-1}$ ,  $Sc$  is the temperature dependent Schmidt number for CO<sub>2</sub> (Wanninkhof 1992).  $k_{600}$  is estimated from the wind speed according to Cole and Caraco (1998):

$$k_{600} = 2.07 + 0.215 U_{10}^{1.7} \quad (8)$$

Hourly wind speed data at 10 m above ground ( $U_{10}$  in Eq. 9) at all 70 lakes were received from the Norwegian Reanalysis Archive (Furevik and Haakenstad 2012) and aggregated into July–August means.

### Lake pelagic CO<sub>2</sub> production

From the dataset, it was not possible to distinguish between lateral input of CO<sub>2</sub> (surface- and ground water flow) and in-lake production of CO<sub>2</sub> (microbial and photochemical mineralization of DOC). Lake pelagic CO<sub>2</sub> production ( $\text{CO}_{2,\text{prod}}$ ;  $\text{mg C m}^{-2} \text{d}^{-1}$ ) will therefore be used as a term for the sum of the in situ DOC mineralization and the lateral input. Assuming that the deviation of CO<sub>2</sub> from saturation is kept at steady state due to production, lateral input, consumption and evasion, the air-water flux of CO<sub>2</sub> ( $F_{\text{CO}_2}$ ;  $\text{mg C m}^{-2} \text{d}^{-1}$ ) can be written as:

$$F_{\text{CO}_2} = \text{CO}_{2,\text{prod}} - \text{PP}_A \quad (9)$$

Positive and negative values of  $F_{\text{CO}_2}$  are evasion and invasion across the air–water interface, respectively. Rearranging Eq. (9), we estimate  $\text{CO}_{2,\text{prod}}$  as the sum of  $F_{\text{CO}_2}$  and  $\text{PP}_A$ .

### Statistical analysis

All data analysis was performed using the open-source software R version 3.4.1 (R Development Core Team, 2017). For linear modeling of the CO<sub>2</sub> production, consumption, and evasion in the lakes the explanatory variables were DOC ( $\text{mg l}^{-1}$ ), TP ( $\mu\text{g l}^{-1}$ ) and TN ( $\text{mg l}^{-1}$ ). The predictors were chosen using AICc in backwards stepwise regression. For estimation of the best predictor, the largest value of the standardized regression coefficients was used. All error estimates are given in standard errors (standard deviation divided by the square root of the number of observations:  $\text{SE} = \text{SD} / \sqrt{n}$ ).

## Results

### Modeling the AQY spectra

The optical parameters  $a_{420}$  and  $\text{SUVA}_{400}$  explained 26–64% of the variation in AQY across lakes. The variation in AQY explained by the parameters decreased with wavelength giving a higher percentage explained at shorter wavelengths where AQY variability between lakes is larger (Table S2; data from Koehler et al. (2016)). The relative magnitude of the

sums of squares (SS) of the fixed terms in the model (Eq. 1) can be used to rank their contribution to the variance of the predicted AQY. While wavelength was by far the largest variance contribution ( $SS = 48.9$ ;  $p \ll 0.001$ ),  $a_{420}$  contributed about five times ( $SS = 0.87$ ;  $p = 0.023$ ) as much to the variance in modeled AQY as  $SUVA_{400}$  ( $SS = 0.18$ ;  $p = 0.027$ ). Monte Carlo simulations of the AQY spectra based on these regression relationships ( $n = 70$ ) resulted in a SE ranging between 0.9% and 1.8% of the wavelength integrated AQY's. The SE was negatively related to  $a_{420}$  ( $r = -0.73$ ; data not shown), indicating that the model fits brown lakes somewhat better than clear ones. The SE of the AQY had an almost one to one fit with the SE of the DIC photoproduction. The uncertainty of the modeled AQY thus propagated through to the DIC photoproduction estimate and the uncertainties of the absorption spectra or the downwelling irradiation did not contribute substantially.

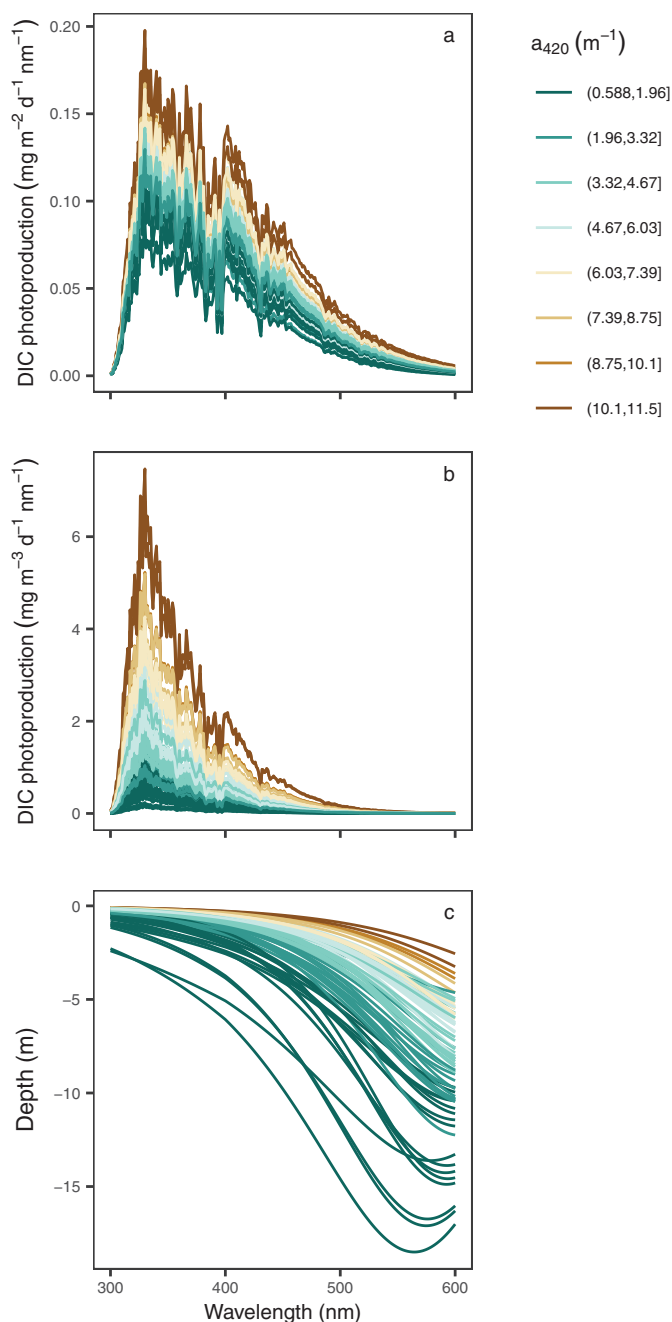
### CO<sub>2</sub> saturation

Out of the 70 lakes in this study, 62 were supersaturated with CO<sub>2</sub> while 6 lakes were close to saturation or slightly undersaturated and 2 were clearly undersaturated with CO<sub>2</sub>. DOC concentrations were strongly related to  $a_{420}$  ( $r = 0.88$ ), and the CO<sub>2</sub> saturation deficit was positively related to both DOC and  $a_{420}$  ( $r = 0.50$  and  $0.61$  for DOC and  $a_{420}$ , respectively). The CO<sub>2</sub> and O<sub>2</sub> saturation deficits were negatively correlated ( $r = -0.70$ ), and the O<sub>2</sub> saturation deficit was negatively related to DOC concentrations and  $a_{420}$  ( $r = -0.74$  and  $-0.69$ , respectively; Fig. S6).

### Photochemical DIC production

$a_{420}$  and  $SUVA_{400}$  in the sampled lakes varied between 0.60 and 11.47 m<sup>-1</sup>, and 0.16 and 1.33 L mg C<sup>-1</sup> m<sup>-1</sup>, respectively (Table S1). Integrating the estimated areal photochemical production of DIC (Fig. 2a), over wavelengths (300–600 nm) gave a range in photoproduced DIC between 8.4 mg C m<sup>-2</sup> d<sup>-1</sup> ± 1.5% and 21.4 mg C m<sup>-2</sup> d<sup>-1</sup> ± 1.0%; (Table S1) in the lakes. Both  $SUVA_{400}$  and  $a_{420}$  were negatively related to pH ( $r = -0.51$  and  $r = -0.28$ , respectively; Fig. S7) and positively related to iron concentrations (Fe;  $r = 0.35$  and  $r = 0.74$  for  $SUVA_{400}$  and  $a_{420}$  respectively; Fig. S7). A multiple linear regression model showed equal sized but opposite effects of pH and Fe concentrations on the estimated DIC photoproduction rates ( $R^2 = 0.31$ , Table S3). The interaction term between the predictor variables was nonsignificant ( $p > 0.05$ , Table S3).

In lakes with high  $a_{420}$ , the shorter wavelengths are absorbed at the surface, resulting in high DIC photoproduction in the top layer compared to lakes with lower  $a_{420}$  (Fig. 2b). While in the brownest lakes irradiance of all photochemically active wavelengths was absorbed within the first meter, this irradiance penetrated further in clearer lakes, allowing for DIC photoproduction to take place at greater depth. Most DIC photoproduction is induced by absorption of photons with wavelengths in the UV and violet part of the



**Fig. 2.** Estimated photoproduction spectra of dissolved inorganic carbon (DIC) from all 70 study lakes. In (a), the estimated areal DIC photoproduction (mg C m<sup>-2</sup> d<sup>-1</sup> nm<sup>-1</sup>) spectra are shown; and (b) shows the estimated volumetric DIC photoproduction (mg C m<sup>-3</sup> d<sup>-1</sup> nm<sup>-1</sup>) spectra just below the surface. In (c) the depth at which the volumetric DIC photoproduction (mg C m<sup>-3</sup> d<sup>-1</sup> nm<sup>-1</sup>) is 1% of that just below the surface is shown, indicating also the depth that receives 1% of incoming radiation. The color gradient goes from dark blue for lakes with low  $a_{420}$  to brown for lakes with high  $a_{420}$ .

spectrum (Vähätalo et al. 2000). Of the estimated areal photochemical DIC production in all lakes 85% ± 0.1% and 93% ±

**Table 1.** Regression coefficients for regressions predicting lake pelagic CO<sub>2</sub> production, consumption, and evasion.

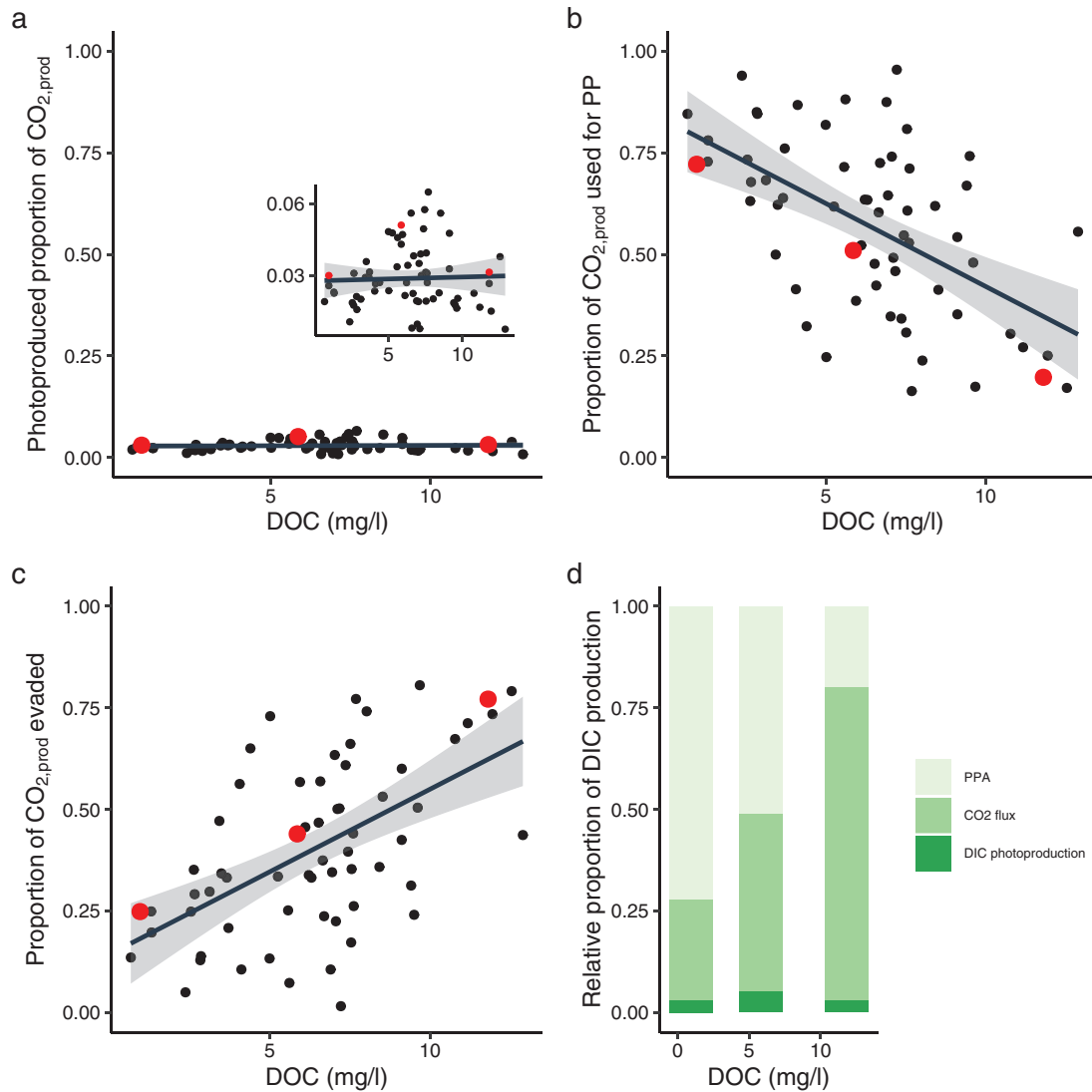
Response	Predictors	Coefficient estimates (SE, significance levels)	R <sup>2</sup>
Lake pelagic CO <sub>2</sub> production	TP + TN	29.9 (±6.6***), 338.0 (±116.9**)	0.47
Areal primary production (PP <sub>A</sub> )	DOC + TP + TN	-29.2 (±7.4***), 21.9 (±4.6***), 174.8 (±75.7*)	0.47
CO <sub>2</sub> flux	DOC + TP	35.4 (±8.9***), 11.2 (±4.9*)	0.33

Significance codes:

\*\*\*  $p < 0.001$ ,

\*\*  $p < 0.01$ ,

\*  $p < 0.05$ .



**Fig. 3.** The relative proportions of **(a)** DIC photoproduction (the inset figure is zoomed in on the y-axis); **(b)** CO<sub>2</sub> flux and; **(c)** areal primary production (PP<sub>A</sub>) to total lake pelagic CO<sub>2</sub> production ( $CO_{2,prod} = F_{CO_2} + PP_A$ ; Eq. 9). In **(d)** is an example of the relative proportion of DIC production for three lakes with low (0.95 mg C l<sup>-1</sup>), medium (5.85 mg C l<sup>-1</sup>), and high (11.84 mg C l<sup>-1</sup>) DOC concentrations. Red dots in the regression plots indicate the three example lakes.



0.1% was induced by wavelengths shorter than 465 and 500 nm, respectively (Fig. S5). Therefore, almost all DIC photoproduction took place in the top 5 m of all lakes, regardless of their color (Fig. 2c). In some of the clearest lakes, light of wavelengths > 500 nm penetrated as deep as 15–20 m (Fig. 2c). The contribution to total DOC photomineralization by wavelengths > 500 nm was minor (Figs. 2a and S3), and the majority of DIC photoproduction thus occurred in the top 5 m of the water column, even in the clearest lakes. The estimated percentage of the DOC standing stock that was photomineralized each day averaged about 2% (0.2–3%) at the surface and 0.3% (0.1–1%) at 1-m depth (Fig. S8). At the surface, the photomineralized share of the standing stock of DOC increased somewhat with increased DOC concentration, while at one meter the relationship was the opposite.

#### Lake pelagic CO<sub>2</sub> production and consumption

Estimations of summer lake pelagic CO<sub>2</sub> production (photochemical and biological mineralization + lateral input of CO<sub>2</sub>) in the studied lakes ranged between 120 and 1770 mg C m<sup>-2</sup> d<sup>-1</sup>. The best predictors for lake pelagic CO<sub>2</sub> production were TP (μg P l<sup>-1</sup>) and TN (mg N l<sup>-1</sup>; Table 1). The relative contribution of DIC photoproduction to lake pelagic CO<sub>2</sub> production averaged 3.0% ± 0.2% regardless of DOC concentration (Fig. 3a,d). Primary production in the lakes was negatively related to DOC concentration and positively related to nutrient content, mainly TP (Table 1). The share of lake pelagic CO<sub>2</sub> production used for primary production was thus smaller in lakes with a high DOC concentration than in lakes with a low DOC concentration (Fig. 3b,d).

#### CO<sub>2</sub> flux

The majority of the lakes were net sources of CO<sub>2</sub> to the atmosphere. The CO<sub>2</sub> flux ranged from -0.12 to 1.0 g C m<sup>-2</sup> d<sup>-1</sup>. CO<sub>2</sub> evasion from lakes was best explained by DOC concentration, followed by TP (Table 1). Assuming that all photochemically produced DIC was emitted as CO<sub>2</sub> from supersaturated lakes, the relative contribution of estimated DIC photoproduction to total CO<sub>2</sub> efflux ranged between 1.4% and 36%, averaging 9% ± 1%, and with higher contribution in lakes with low than in lakes with high DOC concentrations (Fig. S7). The source of the remaining CO<sub>2</sub> efflux must be attributed to respiration and lateral CO<sub>2</sub> input. Of the total DIC production in the lakes, a larger share was emitted as CO<sub>2</sub> in lakes with high than in lakes with low DOC concentrations (Fig. 3c,d).

#### Discussion

We estimated DIC photoproduction in boreal lakes using modeled spectra of irradiance and AQY, and spectra of attenuation coefficients and absorption extrapolated from the measured PAR to the UV region from 70 lakes in Norway and Sweden. We found that DIC photoproduction contributed on average 9% ± 1% to the CO<sub>2</sub> emission from the lakes.

Regarding that this percentage decreases with increased DOC concentrations and that water temperatures as well as DOC and nutrient concentrations in boreal lakes are increasing (Larsen et al. 2011b; O'Reilly et al. 2015), we expect that the relative contribution of sunlight for CO<sub>2</sub> production in boreal lakes may decline in the future.

The AQY spectra were modeled using regressions between AQY at discrete wavelengths and the optical parameters SUVA<sub>400</sub> and *a*<sub>420</sub>, which were set up based on AQY spectral measurements of 25 lakes worldwide (Koehler et al. 2016). While *a*<sub>420</sub> is a proxy for CDOM content, SUVA<sub>400</sub> is well correlated with DOC aromaticity, and both parameters describe absorbing properties of the DOC. (Koehler et al. 2016). Even though SUVA<sub>400</sub> is well correlated with DOC aromaticity, SUVA<sub>254</sub> is usually a better indication of DOC aromaticity. Likewise, in the study by Koehler et al. (2016), SUVA<sub>254</sub> was somewhat better correlated with AQY than SUVA<sub>400</sub> was. However, the difference in *R*<sup>2</sup> between SUVA<sub>400</sub> and SUVA<sub>254</sub> as linear predictors of AQY was minor (Table S2). Running the AQY model (Eq. 1) on the data from Koehler et al. (2016) with SUVA<sub>254</sub> produced similar spectra as with SUVA<sub>400</sub>, mean values of the Monte Carlo simulations had a one to one fit (Fig. S4) and there were no significant differences in SEs between the two. We therefore used the measured SUVA<sub>400</sub> instead of an extrapolated value of SUVA<sub>254</sub>. The uncertainty in the modeled AQY spectra propagated through to the DIC photoproduction estimates. The SEs in the modeled AQY's were however small (1.2% ± 0.02%) and the errors in the estimated DIC photoproduction were therefore also small. Additionally, the AQY spectra estimated in this study match spectra from other studies on boreal lakes well (Koehler et al. 2014; Groeneveld et al. 2016; Vachon et al. 2016).

Estimated DIC photoproduction contributed about 3% (1–5%) of the total production and lateral inflow of CO<sub>2</sub> in the 62 lakes supersaturated with CO<sub>2</sub>. Further, assuming that all photochemically produced DIC is outgassed from the lakes, the relative share of DIC photoproduction to total CO<sub>2</sub> emission averaged about 9% across the 70 study lakes. These results conform to earlier studies on photomineralization of DOC and CO<sub>2</sub> flux from boreal lakes. For example, the contribution of DIC photoproduction to total DOC mineralization in two Swedish humic lakes amounted to about 7% (Jonsson et al. 2001) and 6% (Chmiel et al. 2016). In a third lake, the mean contribution of DIC photoproduction to CO<sub>2</sub> out flux was 1–8%, depending on the time of the year (Groeneveld et al. 2016). Or, in a large-scale modeling study for 1086 Swedish lakes, the mean contribution of DIC photoproduction to out flux of CO<sub>2</sub> was about 12% and upscaling to the entire boreal region about 9–12% (Koehler et al. 2014). However, in other aquatic systems than boreal lakes, photochemical degradation has been found to have an important role in aquatic carbon cycling. In arctic surface waters photochemical reactions accounted for 75% of the total DOC processed (Cory et al. 2014) and in a number of boreal streams photochemical

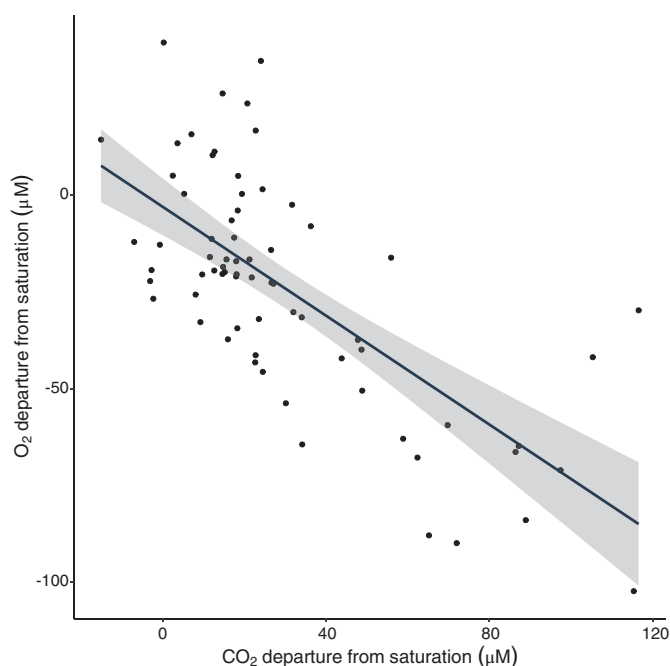
degradation accounted for more than 60% of DOC losses (Molot and Dillon 1997).

Rates of DIC photoproduction in lakes are controlled by three wavelength-dependent processes: the amount of sunlight reaching the lake surface; the fraction of this that is absorbed by CDOM across wavelengths; and the amount of DIC produced per unit absorbed light (AQY) (Cory and Kling 2018). The latter two processes had the largest variations between our 70 study lakes while the solar irradiation spectra were similar, owing to the fact that we sampled in a similar geographic region and time, and that cloud cover variability was low. Both AQY and the CDOM fraction of absorbed irradiance are dependent on the quantity and quality of CDOM in the water. Volumetric DIC photoproduction rates at specific depths are therefore closely related to CDOM content.

While the variability in absorption coefficients between lakes was substantial, the total estimated areal photochemical production of DIC did not differ as much, as similarly shown in earlier studies (Granéli et al. 1996; Koehler et al. 2014). Lower  $a_{420}$  allows light to penetrate deeper down in the water column and DIC photoproduction to take place at greater depths compared to waters with higher  $a_{420}$ . In the latter, all short wavelength photons are strongly absorbed by the DOC and therefore all photoproduction occurs close to the water surface. The absolute areal DIC photoproduction rates were similar whether they were integrated over the entire lake depth or over five meters, indicating that even in the clearest lakes all photochemical production of DIC takes place in the top five meters, where the sampling took place. Both  $SUVA_{400}$  and  $a_{420}$  were negatively related to pH, and positively related to Fe concentrations (Fig. S9). This implies that the effect of extrinsic variables may affect the intrinsic properties of the DOC and therefore the DIC photoproduction rates. A positive correlation between Fe concentrations and CDOM absorption (e.g.,  $a_{420}$ ) has been shown before (Kritzberg and Ekström 2011).  $SUVA_{400}$  was principally related to pH. As  $SUVA_{400}$  is a measure of the aromatic character of the DOC, this implies that aromaticity is increasing at decreasing pH. In acidic waters, DIC photoproduction rates have frequently been reported to increase with decreasing pH (Panneer Selvam et al. 2019). In alkaline waters, the relationship between photochemical degradation of DOC and pH is less certain. While some studies find photomineralization rates to keep decreasing as pH increases (Bertilsson and Tranvik 2000; Molot et al. 2005), others report that they start increasing as pH increases above 7 (Pace et al. 2012; Panneer Selvam et al. 2019). Iron concentrations are also known to interact with pH, having a stronger positive effect on CDOM absorption and hence DIC photoproduction rate under acidic conditions (Gu et al. 2017). However, the pH in the study lakes ranged between 6.3 and 8.0 with two outliers at 5.4 and 8.9 and was thus close to neutral, possibly explaining why the interaction term between Fe and pH in our model was not significant.

Photons entering the water column are likely to be absorbed, if not by DOC, by phytoplankton, nonalgal particles, or by the water itself. Lake absorption spectra show that close to all the photons in the UV region and the largest fraction of the photons in the PAR region were absorbed by DOM, and only a small number were absorbed by other chromophoric compounds (Fig. S3; see also Thrane et al. (2014)). In this study, absorption spectra were only measured in the PAR region. Since the major part of absorption by DOC and thereby the major part of photochemical mineralization of DOC takes place in the UV region, we extrapolated the absorption spectra to wavelengths  $< 400$  nm. We acknowledge that the extrapolation may have led to increased uncertainties of the absorption estimates and through that to increased uncertainties of the DIC photoproduction estimates. However, DOC absorption is rather well studied and the spectra are known to be approximately exponential (Bricaud et al. 1981). Therefore, the mean value of the Monte Carlo simulated spectra and their SE can be assumed to capture most of the uncertainty of the absorption and its propagation through to the DIC photoproduction estimates. For wavelengths between 280 and 400 nm, the DOC absorption fraction in boreal lakes is generally close to 1, and the DOC concentrations are often sufficient for absorption of all incoming photons in this waveband in the top meters of the water column (Williamson et al. 1996). In lakes with high  $a_{420}$ , primary production is constrained to the surface layer due to high light attenuation, resulting in lower rates of primary production on the whole-lake scale (Thrane et al. 2014). However, in regard to the areal photochemical DIC production, the critical limitation is the total amount of DOM-absorbed photons regardless of where in the water column they are absorbed. The major part of the estimated photoproduction of DIC took place above 5 m (Fig. 2c) and was therefore within the mixed zone of the lakes (Fig. S1). The photic zone is deeper in clear than in brown lakes and we can expect that some DIC photoproduction might take place below the mixed zone. However, the photons reaching depths deeper than 5 m are of longer, less photoreactive wavelengths and the contribution of DIC photoproduction at such depth to total lake DIC photoproduction is minor.

While some studies have reported that the vast majority of the  $CO_2$  evasion from boreal lake surfaces is explained by pelagic respiration (Jonsson et al. 2001), others have shown that input of DIC has a larger role than previously thought (Weyhenmeyer et al. 2015). In this study, it was not possible to distinguish between lateral flow and respiration; lake pelagic  $CO_2$  production is therefore used as a common term for the sum of the two. There was, however, a strong relationship between  $O_2$  and  $CO_2$  saturation deficits ( $r = -0.70$ ; Fig. 4). The intercept was not significantly different from 0, meaning that lakes that were saturated with  $O_2$  were also saturated with  $CO_2$ . This relationship indicates that microbial respiration was the predominant source of  $CO_2$  in the lakes. Furthermore, both  $O_2$  and  $CO_2$  concentrations correlated well with DOC,



**Fig. 4.** Lake O<sub>2</sub> departure from saturation with the atmosphere vs. CO<sub>2</sub> departure from saturation with the atmosphere ( $y = -0.70x - 3.02$ ,  $R^2 = 0.49$ ,  $p < 0.001$ ).

but not with chlorophyll *a* (Fig. S10). This suggests that the major DOC source for microbial degradation was of terrestrial origin. The strong relationship between DOC concentrations and  $a_{420}$  confirms that the dominant part of the DOC pool in the lakes originated from the terrestrial surroundings.

Sampling of the lakes used in this study was performed during mid-summer in July and August. Our results cannot be extrapolated to estimate annual rates, but rather present a picture of summer conditions. Photochemical reactivity of DOC depends on the degree of aromaticity (Bertilsson and Tranvik 2000). As DOC leaves the soil and enters the aquatic systems, it will be altered through both biological and photochemical reactions and lose aromaticity (Brinkmann et al. 2003), becoming less photoreactive. Hence, the DOC photochemical reactivity is linked to light exposure time. AQY spectra of photochemical DOC mineralization show pronounced seasonal variability. Photomineralization rates were found to be higher during seasons with high inputs of DOC to lakes, after snowmelt during spring flood (Vachon et al. 2016), and in connection to rain events in autumn, and lower in summer when DOC inputs are low (Groeneveld et al. 2016; Vachon et al. 2016). Photochemical DIC production is dependent on both irradiation and on DOC composition. The AQY might thus be higher in autumn than in summer but due to less sunlight in autumn than in summer, the amount of DIC photoproduction does not necessarily differ substantially between the two seasons. However, increased

light absorption due to brownification may lead to enhanced lake stratification (Williamson et al. 2015), and especially it may give rise to microlayers of stratification at the surface where most irradiance is absorbed. The CO<sub>2</sub> concentrations in these microlayers could thus be much higher than in the underlying water, causing increased rates of photochemically induced CO<sub>2</sub> emissions from brown lakes, especially during the summer months when daily irradiation rates are high. We did not see any indication of increased thermal stratification with increased CDOM content in the study lakes. The CO<sub>2</sub> concentrations of the composite samples can therefore be assumed to represent the concentrations in the entire mixed layers.

On the other hand, pelagic respiration is strongly related to temperature and, therefore, also has a seasonal pattern with higher rates during summer than the rest of the year (Vachon et al. 2016). The relative contribution of photomineralization to total pelagic CO<sub>2</sub> production can thus be assumed to be lower during summer. This was confirmed by Vachon et al. (2016) where the relative contribution of photochemical DIC production to total pelagic CO<sub>2</sub> production in three lakes averaged 14% over the year with larger contribution in spring (26%) than in summer (7.6%) and autumn (12%). The mean value of lake pelagic CO<sub>2</sub> production in the 70 lakes in our study was 616.4 mg C m<sup>-2</sup> d<sup>-1</sup>, ranging from 118.7 to 1769.1 mg C m<sup>-2</sup> d<sup>-1</sup>. These numbers accord with measurements of DOC mineralization in other boreal lakes at summer conditions (Jonsson et al. 2001; Vachon et al. 2016). The DIC photoproduction rates and their relative contribution to lake CO<sub>2</sub> production and evasion also correspond to measures and estimates in previous studies (Jonsson et al. 2001, Vachon et al. 2016). The typical seasonal variations in both microbial and photochemical mineralization rates reported from other lakes make it likely that the role of DIC photoproduction also in the 70 boreal lakes of this study is larger during spring and autumn conditions than found here at summer conditions.

In this study, we estimated photochemical mineralization of DOC. Other photochemical processes in the water column may also have a large impact on the aquatic carbon cycle. Such processes are photomineralization of organic nutrients and partial photooxidation of DOC (Bertilsson and Tranvik 2000). In the latter processes, recalcitrant DOC is transformed to more biologically available organic compounds (Bertilsson and Tranvik 1998). Microbial consumption of such photodegraded compounds is thus often preferred over the nonphotodegraded compounds (Alleson et al. 2016). Although most photochemical processes take place near the surface, photochemically produced carboxylic acids may mix downwards and be a source of labile DOC in the entire mixed layer (Bertilsson and Tranvik 1998). Enhanced microbial degradation of photodegraded DOC may have an impact on aquatic carbon cycling as large as photomineralization.

Post-acidification recovery, increased vegetation cover in catchments, and a wetter climate promote carbon export to



lakes (Finstad et al. 2017; de Wit et al. 2018). Concentrations of allochthonous DOC are thus predicted to increase in most boreal lakes (Larsen et al. 2011b). Input of DOC in boreal lakes is correlated with export of TP and TN (Dillon and Molot 2005), and the predicted increased nutrient levels will most likely promote microbial activity and thus pelagic CO<sub>2</sub> production. Although DOC and TP have contrasting effects on primary production, the net effect of enhanced levels will probably be a reduced primary production due to light attenuation in most lakes with initial moderate to high DOC concentrations (Thrane et al. 2014). In lakes with low initial levels of DOC, an increase in DOC and nutrient levels could lead to enhanced primary production, and thus an enhanced level of autochthonous DOC which in turn could result in enhanced microbial respiration (Lapierre and del Giorgio 2014). Increased DOC input will thus most likely lead to enhanced levels of heterotrophy in boreal lakes (Larsen et al. 2011a). Moreover, higher levels and more frequent input of fresh, photolabile, DOC are to be expected and therefore the AQY and by that DIC photoproduction can be expected to increase as well. However, the rather small difference in estimated areal DIC photoproduction between lakes compared to the wide ranges in DOC and  $a_{420}$  indicates that enhanced rates of photochemical mineralization will not be a major contributor to shifting levels of boreal lake net heterotrophy. In all lakes, all photons active to DOC photochemistry were absorbed within the top five meters, regardless of DOC concentration. This suggests that the contribution of enhanced rates of DIC photoproduction to lake net heterotrophy will probably be largest when clear, shallow lakes undergo browning. While the observed strong increase in surface water temperatures (O'Reilly et al. 2015) will promote microbial respiratory activity, photomineralization is only weakly temperature dependent (Chatwal and Arora 2007), hence the relative contribution of DIC photoproduction to total CO<sub>2</sub> production will most likely decrease in boreal lakes under a changing climate.

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#### Conflict of interest

None declared.

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**Table S1.** Measured and modelled lake parameters for the 70 lakes used in this study. The first 11 parameters from the left are measured while the last 4 are modelled or calculated as described under methods.

Abbreviations.  $a_{420}$ : absorption coefficient at 420 nm; SUVA<sub>400</sub>: Specific ultraviolet absorbance at 400 nm; PP<sub>A</sub>: Areal primary production

Lake	Area (km <sup>2</sup> )	Depth (m)	pH	O <sub>2</sub> (μmol/l)	CO <sub>2</sub> (μmol/l)	TOC (mg/l)	TN (mg/l)	TP (μg/l)	Fe (μg/l)	$a_{420}$ (m <sup>-1</sup> )	SUVA <sub>400</sub> (L mgC <sup>-1</sup> m <sup>-1</sup> )	PP <sub>A</sub> (mg C m <sup>-2</sup> d <sup>-1</sup> )	Lake pelagic		
													CO <sub>2</sub> flux (mg C m <sup>-2</sup> d <sup>-1</sup> )	CO <sub>2</sub> production (mg C m <sup>-2</sup> d <sup>-1</sup> )	Areal DIC photoproduction (mg C m <sup>-2</sup> d <sup>-1</sup> ± se)
Gjersjøen	2.64	22.0	7.69	265.7	24.8	6.90	1.28	9.8	162.4	3.41	0.71	581.3	85.6	666.9	12.3 (± 1.2 %)
Øgderen	12.7	9.5	7.23	266.6	63.8	6.94	0.33	19.2	270.0	3.78	0.76	776.0	428.4	1204.4	13.3 (± 1.3 %)
Krøderen	43.9	14.0	6.70	305.5	73.4	4.40	0.25	3.55	97.0	3.09	1.00	181.2	380.2	561.5	12.9 (± 1.1 %)
Rødbyvatnet	1.2	10.0	7.54	258.7	40.5	6.65	0.97	9.35	287.3	3.27	0.70	322.4	214.3	536.7	11.6 (± 1.1 %)
Gjesåsjøen	4.0	3.5	7.07	258.7	64.2	9.61	0.39	15.1	1537.5	5.62	0.83	382.1	415.9	798.0	15.2 (± 1.3 %)
Osensjøen	43.6	70.0	6.56	297.8	52.2	7.69	0.28	3.05	172.7	6.77	1.24	48.2	245.1	293.3	12.0 (± 1.1 %)
Rokossjøen	3.8	9.0	6.71	220.4	81.6	11.81	0.33	8.25	830.7	7.92	0.96	122.6	499.6	622.1	13.0 (± 1.0 %)
Sør Mesna	6.9	17.0	6.71	270.2	66.3	7.51	0.25	6.7	174.8	6.03	1.13	1.3	378.3	546.8	17.7 (± 1.0 %)
Vermundsjøen	3.3	12.0	6.29	219.9	132.9	11.95	0.36	9.25	1333.8	9.07	1.08	283.8	848.7	1132.5	12.5 (± 1.2 %)
Gjørvatnet	2.9	46.0	6.44	352.9	38.9	1.30	0.24	2	7.2	0.60	0.64	330.6	126.3	456.9	15.6 (± 1.1 %)
Kalandsvatnet	3.4	43.0	6.90	337.9	31.3	2.85	0.35	3.8	70.4	2.03	1.00	711.0	129.5	840.5	19.2 (± 1.3 %)
Myrkdalsvatnet	1.6	76.0	6.36	358.2	42.0	0.95	0.09	1	21.0	0.74	1.06	348.4	133.8	482.2	16.6 (± 0.9 %)
Engsetdalsvatnet	4.4	13.0	6.87	310.9	34.6	2.66	0.20	1.15	25.0	1.80	0.95	305.3	145.3	450.6	10.7 (± 1.3 %)
Rotevatnet	1.4	23.0	6.74	315.1	48.8	3.43	0.17	2.55	50.2	2.49	1.05	255.3	255.5	510.8	17.2 (± 1.2 %)
Vatnevatnet	2.0	25.0	6.66	362.2	42.1	1.31	0.17	4.7	24.8	1.34	1.33	539.6	148.9	688.5	17.1 (± 0.9 %)
Einavatnet	13.7	24.0	7.43	292.9	31.4	5.57	1.09	3	73.4	3.13	0.79	292.2	118.3	410.4	10.5 (± 1.4 %)
Ringsjøen	1.2	19.0	7.19	260.5	105.0	8.02	0.56	5.55	383.0	5.62	0.99	188.5	602.3	790.7	13.3 (± 1.4 %)
Sæbufjorden	1.5	18.4	7.08	298.8	39.2	3.66	0.36	3.1	82.0	2.21	0.84	278.3	158.7	437.0	12.4 (± 1.6 %)
Strondafjorden	13.4	34.0	7.22	329.9	29.9	2.83	0.35	2	41.3	1.89	0.91	525.8	93.3	619.0	15.6 (± 1.1 %)
Trevatna	4.8	17.0	6.47	239.8	106.6	9.68	0.35	4.3	422.8	6.54	0.96	136.8	649.8	786.6	13.9 (± 1.4 %)
Bogstadvatnet	1.1	8.0	6.94	266.3	87.6	7.03	0.39	6.6	348.1	4.74	0.96	274.4	516.6	790.9	14.9 (± 1.4 %)
Rødenessjøen	16.0	20.0	7.04	279.3	43.0	9.11	0.98	12.2	279.1	6.72	1.04	285.2	240.2	525.4	10.3 (± 1.3 %)

Rømsjøen	13.7	30.0	6.83	291.7	27.9	6.69	0.40	1.2	65.0	3.96	0.85	277.9	106.8	384.7	14.5 (± 1.2 %)
Edlandsvatnet	2.1	8.0	6.94	316.5	41.4	2.63	0.79	3.85	52.7	1.47	0.79	418.0	245.8	663.8	13.5 (± 1.1 %)
Hetlandsvatn	2.1	33.0	7.19	327.0	23.7	2.37	1.01	2.35	25.0	1.20	0.68	993.0	66.0	1059.0	17.7 (± 1.2 %)
Vatsvatnet	2.2	18.0	7.53	355.6	17.3	4.09	0.48	18.9	168.8	2.39	0.83	673.0	2.4	675.3	15.7 (± 1.8 %)
Vostervatnet	2.7	60.0	6.97	304.0	34.7	3.12	0.61	3.6	31.7	1.66	0.72	427.4	201.2	628.7	13.9 (± 1.2 %)
Jølstravatnet	39.3	46.0	6.33	356.2	40.0	0.66	0.29	0.5	13.0	0.60	1.11	565.5	102.2	667.7	19.8 (± 1.0 %)
Grungevatnet	1.6	27.0	6.75	293.9	41.4	2.54	0.15	1.55	42.2	1.52	0.83	486.8	178.4	665.2	16.4 (± 1.1 %)
Vinjevatt	3.2	13.0	6.68	316.8	37.0	3.49	0.17	1.05	35.0	1.29	0.53	190.2	119.8	310.0	15.6 (± 1.1 %)
Årumvatnet	1.1	20.0	7.07	268.3	121.8	7.11	1.12	20.2	359.4	4.61	0.91	869.1	900.0	1769.1	14.8 (± 1.1 %)
Bergsvannet	3.0	12.1	8.88	317.1	0.8	7.07	0.37	17.85	288.8	4.05	0.82	450.2	-120.7	329.5	15.4 (± 1.1 %)
Goksjø	3.4	16.0	7.23	278.3	132.9	6.58	1.53	27.45	342.6	4.10	0.87	739.4	1009.0	1748.3	15.7 (± 1.1 %)
Hallevatnet	3.7	45.0	6.92	297.9	52.4	5.01	0.74	2	141.3	2.30	0.67	133.3	409.8	543.1	12.6 (± 1.2 %)
Dagarn	1.7	5.5	7.10	276.9	30.8	6.22	0.25	3	50.9	2.12	0.49	260.3	154.1	414.4	13.5 (± 1.2 %)
Langen	1.4	14.0	8.00	277.4	7.8	6.39	0.35	4.9	63.6	0.74	0.16	303.4	-77.3	226.1	11.7 (± 1.1 %)
Rotnessjøen	1.1	26.0	6.64	244.2	114.4	11.18	0.28	4.55	778.3	8.93	1.13	286.1	768.1	1054.2	15.9 (± 1.1 %)
Mylla	1.7	33.0	7.59	287.6	24.9	4.11	0.21	2.45	102.5	2.35	0.78	407.7	63.9	471.7	14.6 (± 1.2 %)
Femsjøen	10.7	18.5	7.02	274.1	42.0	8.42	0.82	7.45	228.6	5.25	0.90	423.7	261.7	685.4	12.7 (± 1.1 %)
Klæmningen	10.0	35.0	7.82	270.9	12.1	8.83	0.43	11.45	118.1	2.44	0.40	582.2	-30.3	551.9	21.4 (± 1.0 %)
Tortrsjøen	1.8	25.0	7.21	290.1	34.3	7.62	0.37	2.35	57.2	4.93	0.91	409.9	167.3	577.2	17.4 (± 1.0 %)
Visten	32.0	10.0	7.20	278.2	14.2	4.86	0.25	6	27.1	1.43	0.42	269.0	-6.7	262.3	17.7 (± 1.1 %)
Stora Le	84.9	52.0	6.97	287.1	28.9	5.25	0.51	1.9	44.6	3.04	0.82	0.7	117.5	304.6	14.7 (± 1.2 %)
Nåsråmnen	2.7	9.0	7.02	255.7	38.2	7.60	0.22	5.85	203.2	3.82	0.73	229.9	207.7	437.6	13.8 (± 1.2 %)
Rangsjøn	2.7	9.0	7.01	258.8	46.9	7.37	0.22	3.2	169.9	5.99	1.14	122.1	234.0	356.1	11.4 (± 1.3 %)
Tisjøen	27.1	17.0	6.88	287.8	31.5	6.53	0.21	4.55	168.1	3.82	0.83	118.3	131.3	249.6	16.0 (± 1.1 %)
Halsjøen	5.2	14.0	5.41	244.3	103.0	12.28	0.27	5.8	1249.5	11.05	1.25	NA	694.6	NA	10.8 (± 1.3 %)
Jangen	4.5	8.0	7.04	237.4	50.0	9.41	0.24	5.25	245.3	6.40	0.96	590.9	292.5	883.4	13.0 (± 1.1 %)
Sør-ålgren	15.5	32.0	7.11	301.8	17.9	7.22	0.29	5.85	164.0	4.70	0.92	496.2	24.3	520.4	12.9 (± 1.3 %)
Möckeln	18.0	22.0	7.27	275.5	30.0	7.54	0.47	12.6	204.3	4.97	0.93	618.3	146.9	765.2	13.2 (± 1.3 %)
Ljusnaren	9.6	20.0	6.94	287.7	32.1	8.52	0.30	6.75	441.8	5.43	0.91	113.2	161.9	275.1	16.1 (± 1.0 %)
Halvarsnoren	16.9	26.0	7.23	305.3	27.5	7.44	0.29	5.4	252.1	4.84	0.92	148.8	124.2	273.0	15.1 (± 1.1 %)
Nåtsjön	2.9	20.0	7.15	297.2	20.7	4.99	0.19	3.45	44.8	2.35	0.65	212.3	50.1	262.4	15.1 (± 1.2 %)

Saxen	7.0	14.0	7.33	309.9	19.0	5.61	0.25	4.85	63.2	2.99	0.76	259.3	37.1	296.4	14.6 (± 1.2 %)
Långbjörken	1.7	4.0	7.16	281.7	32.6	9.50	0.31	9.65	285.9	3.82	0.58	490.8	174.2	665.0	17.3 (± 1.1 %)
Skattungen	19.5	17.5	7.18	277.9	33.6	7.55	0.23	4.25	329.1	5.34	1.00	244.8	158.0	402.8	14.7 (± 1.1 %)
Bäsingen	12.7	10.5	7.25	264.2	47.2	6.11	0.26	8.5	370.7	4.28	0.99	351.1	321.2	672.4	11.7 (± 1.3 %)
Tisken	63.4	8.0	7.30	273.6	24.9	7.06	0.48	8	206.5	3.36	0.68	268.9	97.0	365.9	11.3 (± 1.3 %)
Stora Almsjön	2.0	17.0	6.98	251.9	75.8	12.55	0.27	5.55	537.5	11.47	1.27	2.3	465.2	561.7	10.0 (± 1.3 %)
Dragsjön	1.4	3.5	6.71	241.2	78.8	10.78	0.23	7.25	755.1	8.57	1.12	234.7	535.7	770.5	12.6 (± 1.3 %)
Milsjön	3.3	18.0	6.86	286.4	42.3	9.12	0.25	3.75	220.1	6.86	1.06	130.3	239.0	369.3	12.8 (± 1.6 %)
Stora Korslängen	3.5	15.0	6.70	289.2	37.4	5.94	0.24	2.05	78.3	3.64	0.87	1.1	192.9	313.1	12.4 (± 1.3 %)
Hinsen	11.9	9.0	7.18	275.4	12.4	5.49	0.22	3.55	41.0	1.84	0.49	195.7	-28.9	166.8	11.0 (± 1.3 %)
Storsjön	1.9	9.0	7.19	276.3	13.6	3.77	0.17	3.15	32.3	1.43	0.50	138.1	-19.4	118.7	13.8 (± 1.1 %)
Grycken	3.2	9.0	6.94	264.9	31.8	6.30	0.20	3.3	221.7	3.36	0.75	240.3	140.8	381.1	12.8 (± 1.1 %)
Holmsjön	50.7	8.0	7.33	268.0	34.1	5.86	0.20	3.1	156.2	4.01	0.96	178.0	171.8	349.8	14.0 (± 1.1 %)
Stormaggen	3.0	7.0	7.08	263.4	38.8	7.17	0.21	4.7	138.2	4.28	0.85	171.2	203.3	374.5	12.8 (± 1.2 %)
Forsjösjön	1.9	14.0	7.52	204.1	87.1	12.90	0.91	16.3	149.3	4.74	0.54	895.0	719.5	1614.5	11.1 (± 1.2 %)
Hurdalsjøen	32.8	20.0	6.87	296.6	28.3	3.71	0.41	1.6	51.9	1.84	0.71	296.9	96.0	392.9	12.2 (± 1.1 %)
Harestuvatnet	2.0	13.0	7.37	277.1	61.1	4.06	0.37	4.35	154.8	2.53	0.86	233.7	332.0	565.6	8.4 (± 1.5 %)

Supplementary information

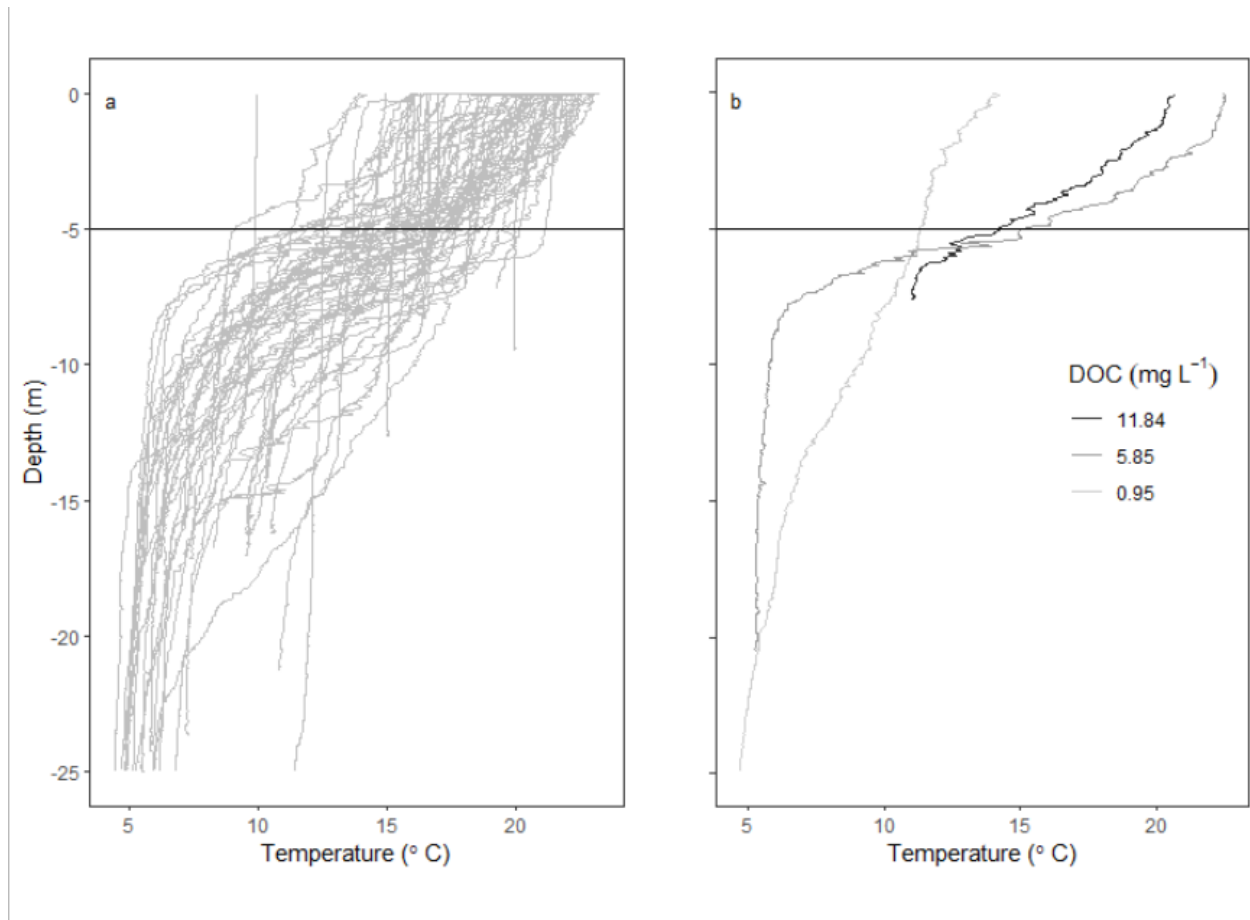


Figure S1. a) Temperature profiles in all the surveyed lakes. b) Temperature profiles of three example lakes with low ( $0.95 \text{ mg C l}^{-1}$ ), medium ( $5.85 \text{ mg C l}^{-1}$ ), and high ( $11.84 \text{ mg C l}^{-1}$ ) DOC concentrations. Samples were taken over the period of July-August during summer stratification, as a single snap-shot record during daytime.



## Modelling apparent quantum yield

**Table S2.** Parameter estimates and model statistics for the linear regression equations between the apparent quantum yield (AQY) and specific UV absorption coefficient at 400 nm ( $SUVA_{400}$ ) as well as the Napierian absorption coefficient at 420 nm ( $a_{420}$ ) for AQY integrated across 300–500 nm and at eight wavelengths between 300 and 440 nm. Data from Koehler et al. (2016).

Predictor variable	Wavelength (nm)	Intercept $\cdot 10^{-6}$	Slope $\cdot 10^{-6}$	R <sup>2</sup>
$SUVA_{400}$	300	282.0	933.4	0.55
	320	203.1	638.6	0.54
	340	149.8	437.7	0.51
	360	113.2	300.0	0.47
	380	87.4	205.3	0.42
	400	68.9	140.0	0.36
$a_{420}$	300	528.1	55.5	0.64
	320	387.8	36.7	0.59
	340	288.7	24.1	0.52
	360	217.4	15.8	0.43
	380	165.4	10.3	0.35
	400	126.8	6.6	0.26
$SUVA_{254}$	300	-206.7	163.3	0.56
	320	-139.5	112.7	0.56
	340	-88.8	77.7	0.53
	360	-52.0	53.4	0.49
	380	-26.1	36.6	0.44
	400	-8.5	25.0	0.38



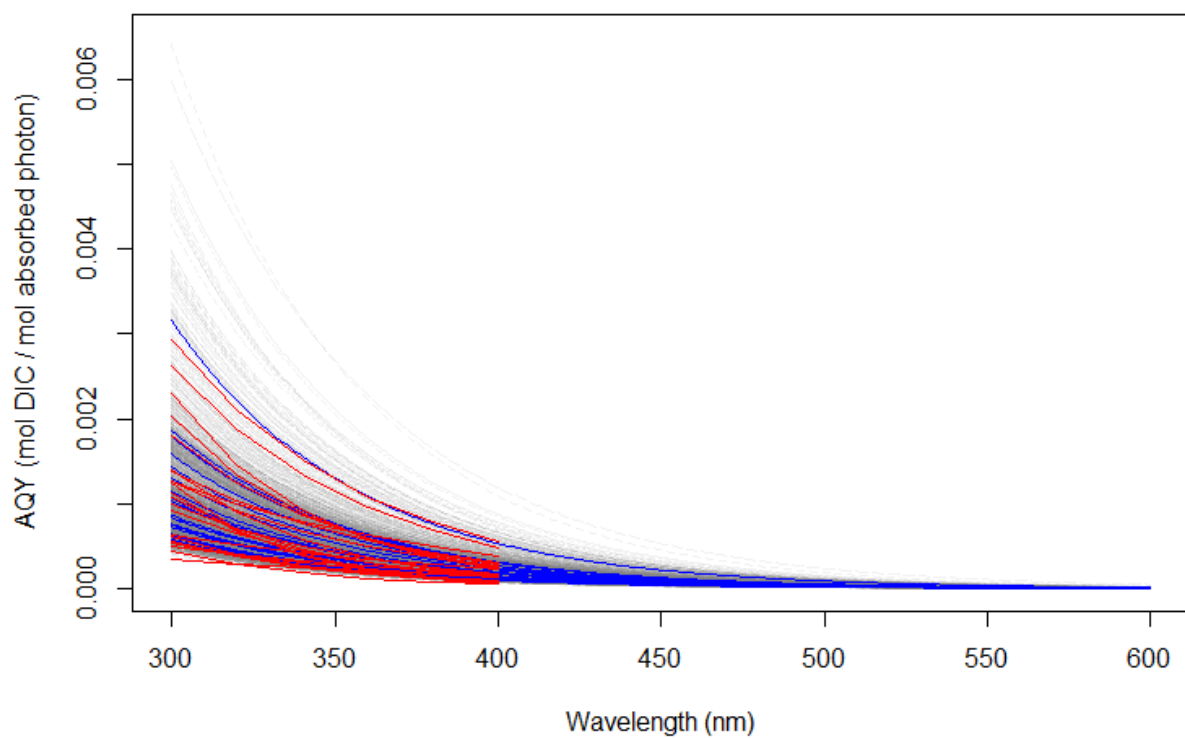


Figure S2. Monte Carlo samples of AQY spectra for the lakes used to derive the predictive regression relationships (Table S2; data from Koehler et al. (2016)). The blue lines are the mean spectra and the red lines are the measured AQY's at six discrete wavelengths between 300 and 400 nm.

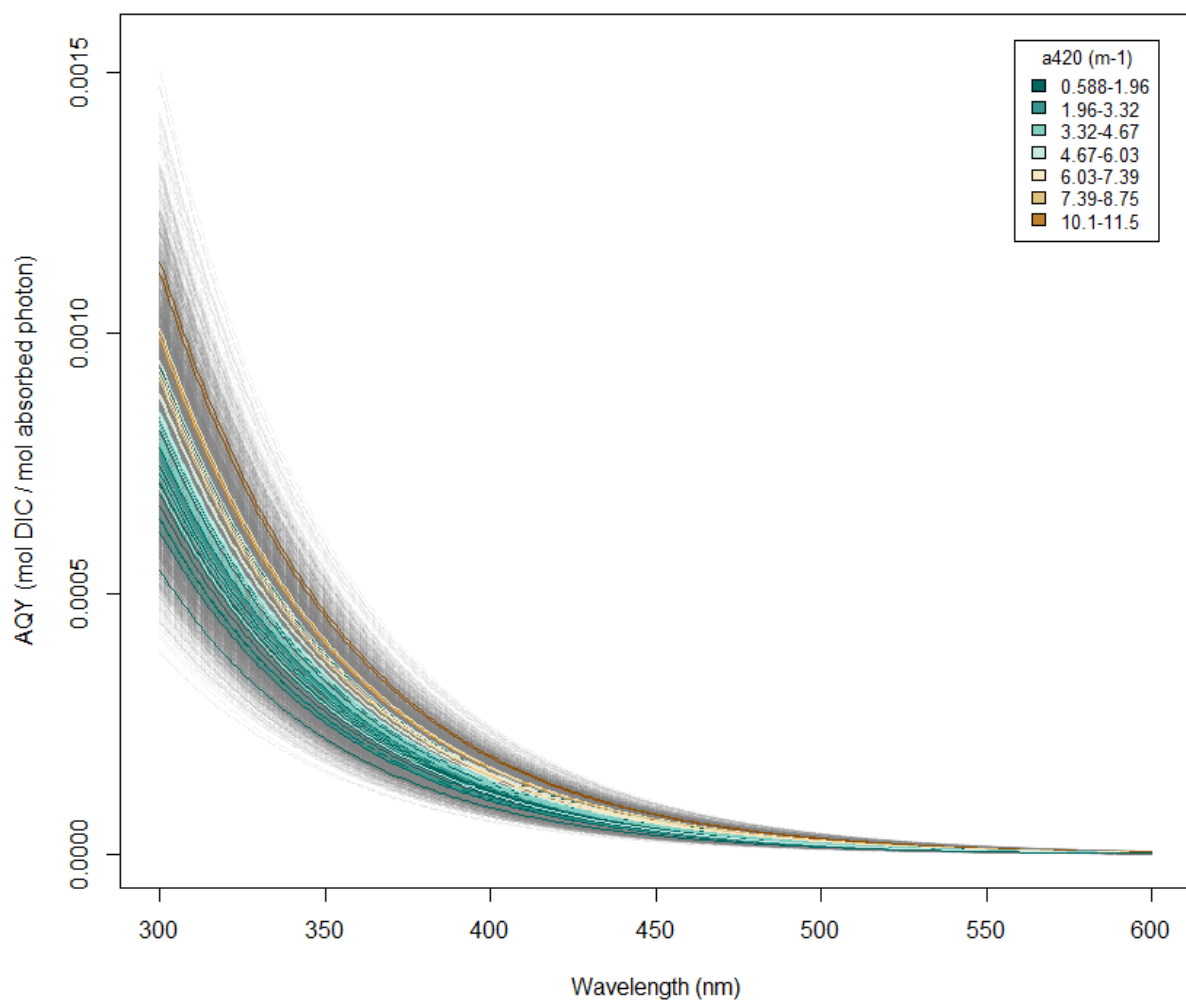


Figure S3. Monte Carlo samples of AQY spectra for all sampled lakes, the colored lines represent the mean spectra of each lake.

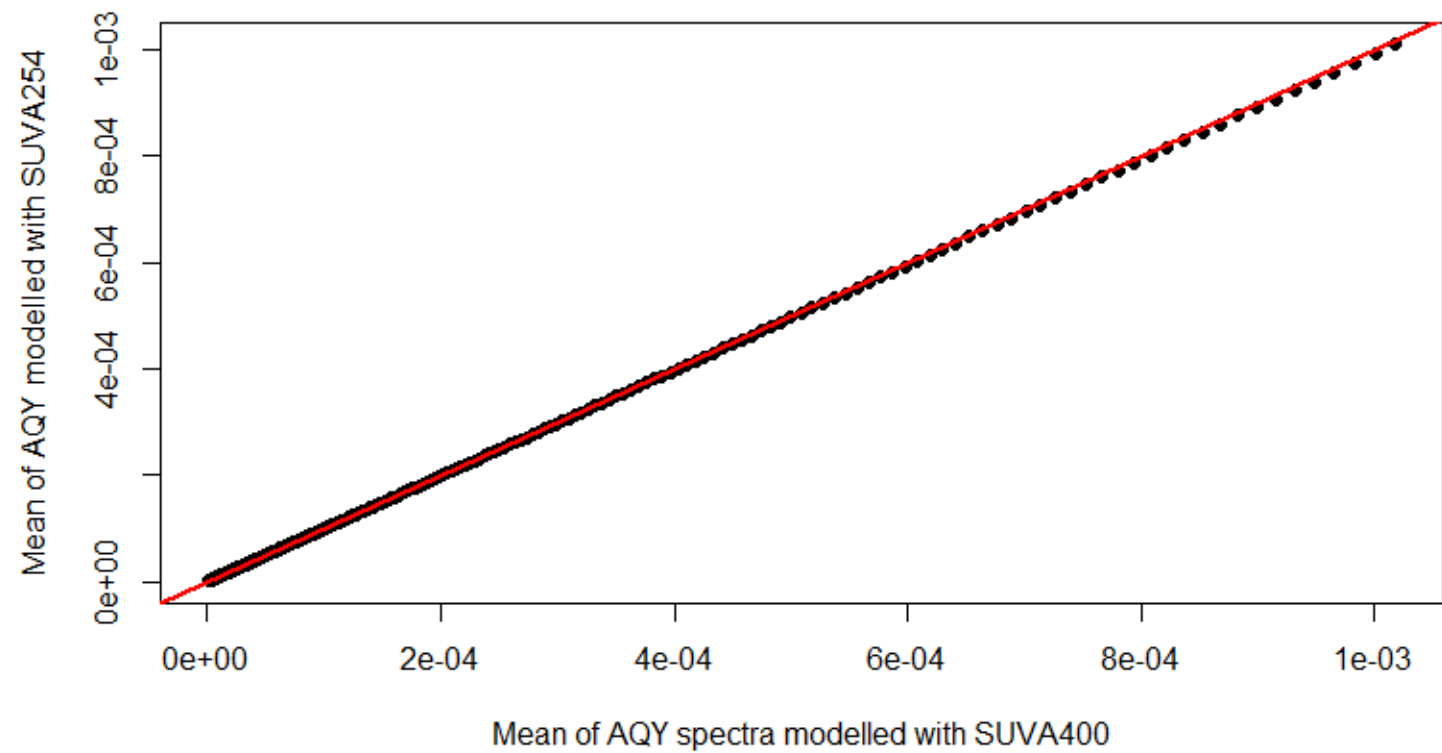


Figure S4. The mean values of each wavelength of the Monte Carlo samples in the AQY model using SUVA254 and a420 versus the AQY model using SUVA400 and a420 (equation 1 in main text). The red line is the 1:1 line.

## Irradiance

The irradiance in the model ( $\text{mWh m}^{-2} \text{d}^{-1} \text{nm}^{-1}$ ) was converted to photon flux ( $\text{mole photons m}^{-2} \text{d}^{-1} \text{nm}^{-2}$ ). The irradiance energy was first converted from mWh to J by multiplication with 3.6 and was then divided by the amount of energy one mole photons contain, using Planck's law and Avogadro's constant:

$$\text{photon flux} = \frac{\text{irradiance} \cdot 3.6}{\frac{hc}{\lambda} \cdot N_A}$$

where  $h$  is Planck's constant (Js),  $c$  is speed of light (m/s),  $\lambda$  is wave length (nm) and  $N_A$  is Avogadro's number ( $\text{mol}^{-1}$ ).

## Error propagation

The uncertainty in DIC photoproduction was calculated through the uncertainties of the absorption and AQY spectra. First the uncertainties ( $\delta$ ) were calculated as the standard error of the mean of the Monte Carlo samples from the standard deviation ( $\sigma$ ) and the sample size ( $n$ ).

$$\delta x = \frac{\sigma x}{\sqrt{n}}$$

The uncertainty of the total absorption coefficient spectrum is the combination of the uncertainties of all absorption coefficient spectra.

$$\delta a_{total} = \sqrt{(\delta a_{NAP})^2 + (\delta a_{PP})^2 + (\delta a_{DOM})^2 + (\delta a_{water})^2}$$

The fraction of the DOM absorbance to total absorption ( $k_{DOM}$ ) had thus the following uncertainty.

$$\frac{\delta k_{DOM}}{\langle k_{DOM} \rangle} = \sqrt{\left(\frac{\delta a_{DOM}}{\langle a_{DOM} \rangle}\right)^2 + \left(\frac{\delta a_{total}}{\langle a_{total} \rangle}\right)^2}$$

And finally the uncertainty of the DIC photoproduction becomes the combination of the uncertainties of  $k_{DOM}$  and AQY for DIC production ( $\Phi_{DIC}$ ).

$$\frac{\delta \psi_{DIC}}{\langle \psi_{DIC} \rangle} = \sqrt{\left(\frac{\delta k_{DOM}}{\langle k_{DOM} \rangle}\right)^2 + \left(\frac{\delta \Phi_{DIC}}{\langle \Phi_{DIC} \rangle}\right)^2}$$

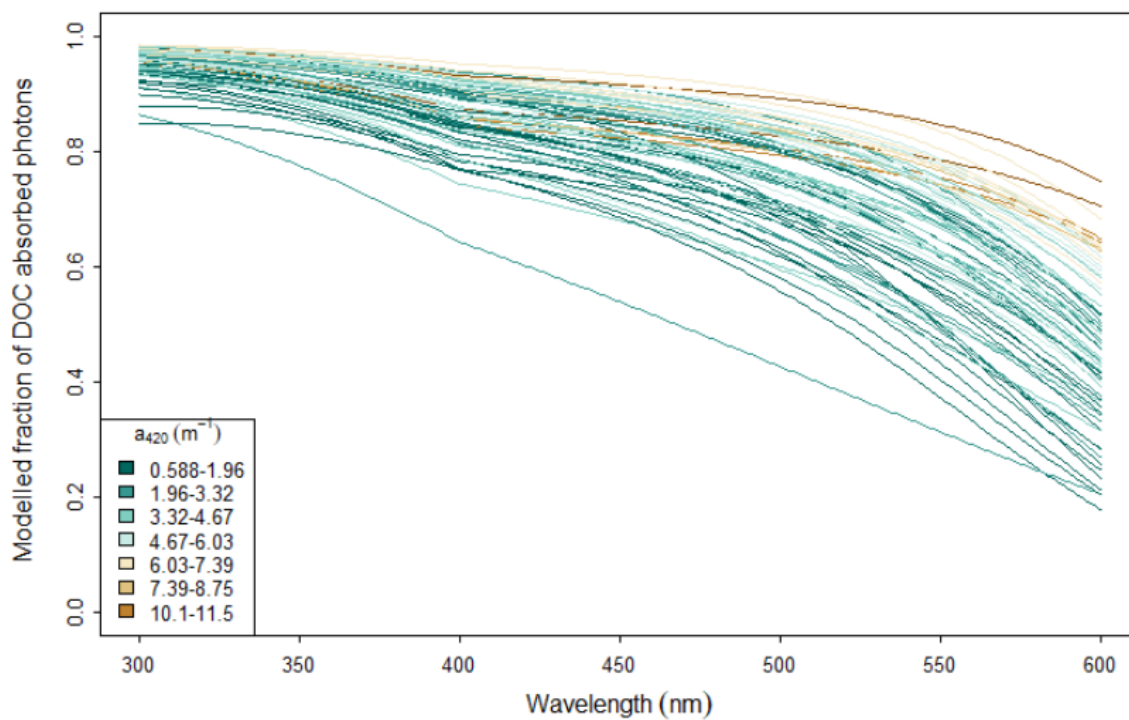


Figure S5. At short wavelengths the fraction of the photons absorbed by DOM is close to 1. The fraction decreases at longer wavelengths, where photochemical reactivity is lower.

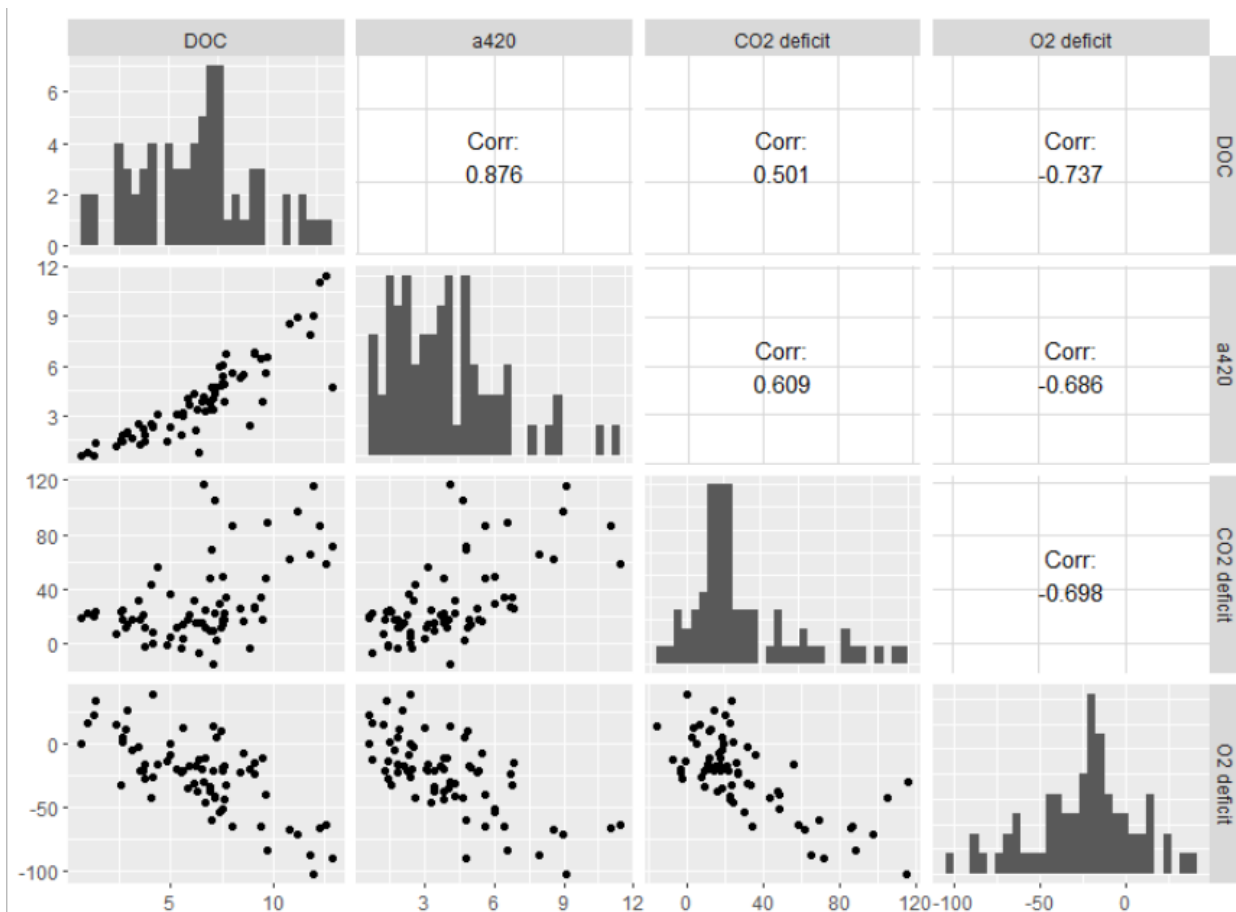


Figure S6. Correlation matrix plot with DOC ( $\text{mg L}^{-1}$ ),  $a_{420}$  ( $\text{m}^{-1}$ ), CO<sub>2</sub> saturation deficit ( $\mu\text{mol L}^{-1}$ ) and O<sub>2</sub> saturation deficit ( $\mu\text{mol L}^{-1}$ ) concentrations. There was a significant negative correlation between CO<sub>2</sub> and O<sub>2</sub> saturation deficits ( $r = -0.70$ ) and saturation deficits of both CO<sub>2</sub> and O<sub>2</sub> were significantly correlated with TOC concentration ( $r = 0.50$  and  $r = -0.74$  respectively) and with  $a_{420}$  ( $r = 0.61$  and  $r = -0.69$  respectively). DOC concentrations were strongly related to  $a_{420}$  ( $r = 0.88$ ).

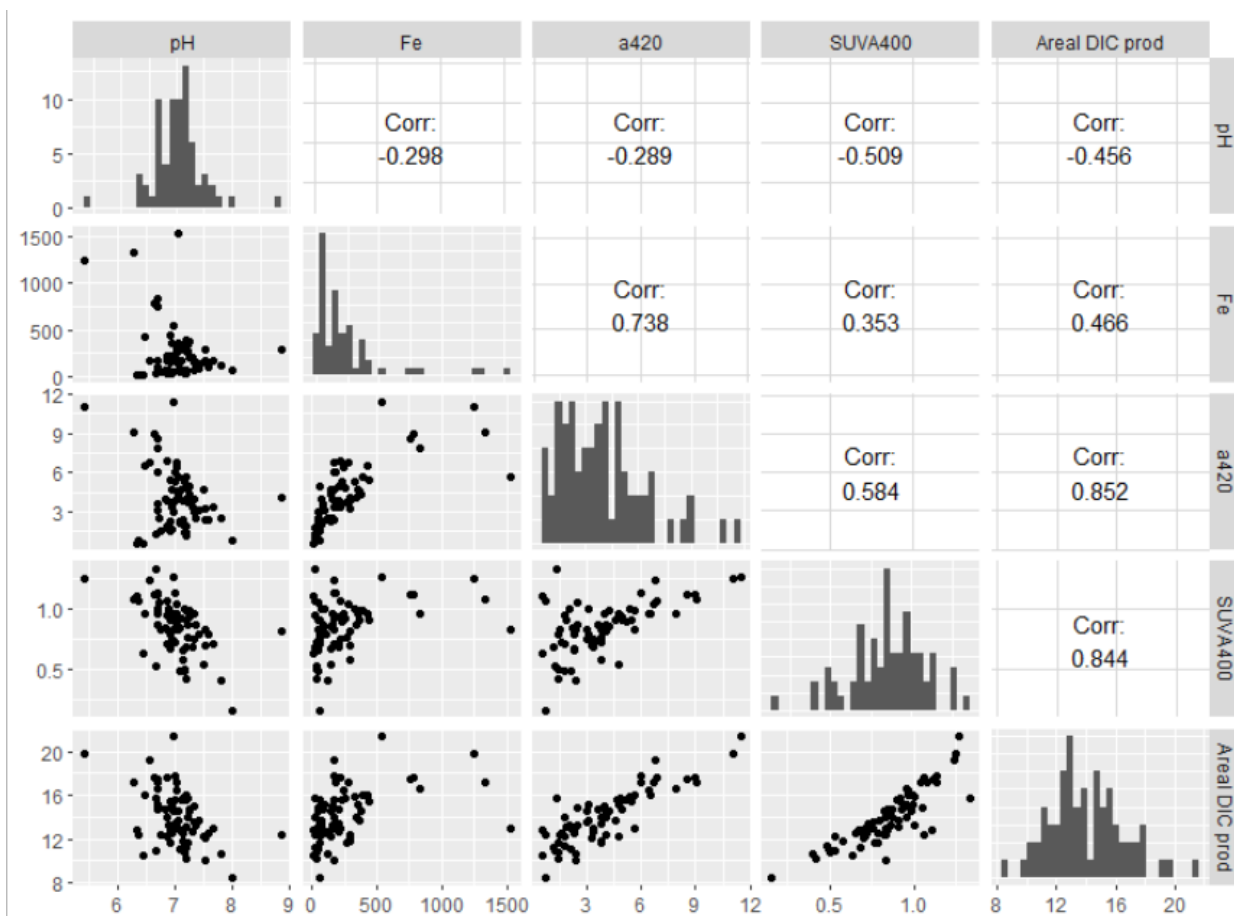


Figure S7. Correlation matrix plot with pH, Fe ( $\mu\text{g/L}$ ),  $a_{420}$  ( $\text{m}^{-1}$ ), SUVA<sub>400</sub> ( $\text{L mg C}^{-1} \text{m}^{-1}$ ) and estimated areal DIC photoproduction ( $\text{mg C m}^{-2} \text{d}^{-1}$ ). SUVA<sub>400</sub> and  $a_{420}$  were negatively related to pH and positively related to Fe. These relations propagated through to the areal DIC photoproduction rates.



**Table S3.** Regression coefficients for multiple linear regressions predicting areal DIC photoproduction rates from Fe concentrations ( $\mu\text{g L}^{-1}$ ) and pH. Significance codes: \*\*\*  $p < 0.001$ , \*\*  $p < 0.01$ , \*  $p < 0.05$ , non-sig  $p > 0.05$ .

<b>Response</b>	<b>Predictors</b>	<b>Estimates (se, significance levels)</b>	<b>R<sup>2</sup><sub>adj</sub></b>
Areal DIC photoproduction	pH + Fe	-1.9(0.6, **), 0.003(0.0009, ***)	0.31
	pH + Fe + pH:Fe	-2.1(0.8, ***), -0.0004(0.01, non-sig), 0.0005 (0.001, non-sig)	0.30

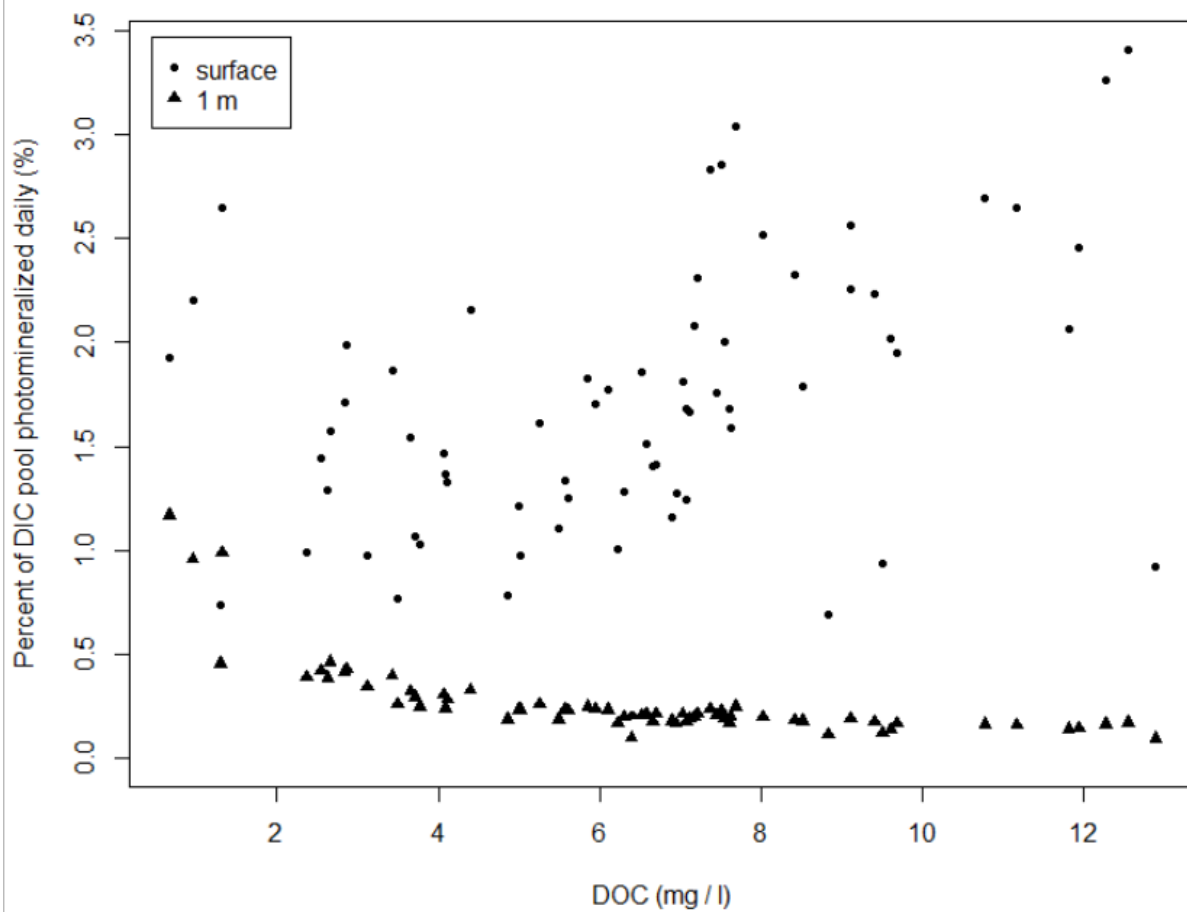


Figure S8. The percentage of the DOC pool that was photomineralized each day as a function of DOC concentration. At the surface, the photomineralized share of the DOC pool increased with DOC concentration, while the opposite was true further down in the water column.

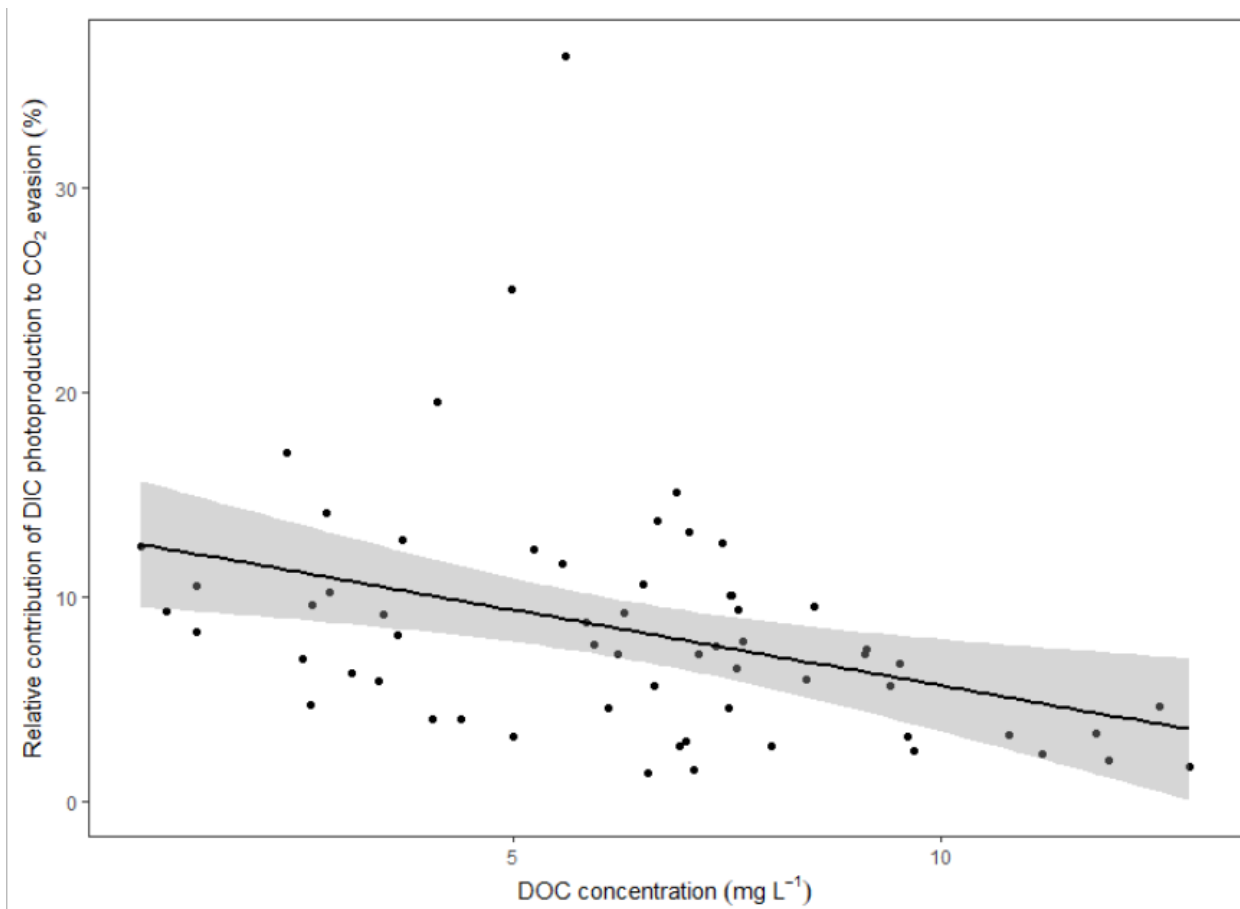


Figure S9. Relative contribution of photoproduced DIC to CO<sub>2</sub> evasion from the lakes as a function of DOC concentration. Assuming that all photochemically produced DIC was emitted as CO<sub>2</sub> from the supersaturated lakes, the relative contribution of estimated DIC photoproduction to total CO<sub>2</sub> efflux decreased with DOC concentration.

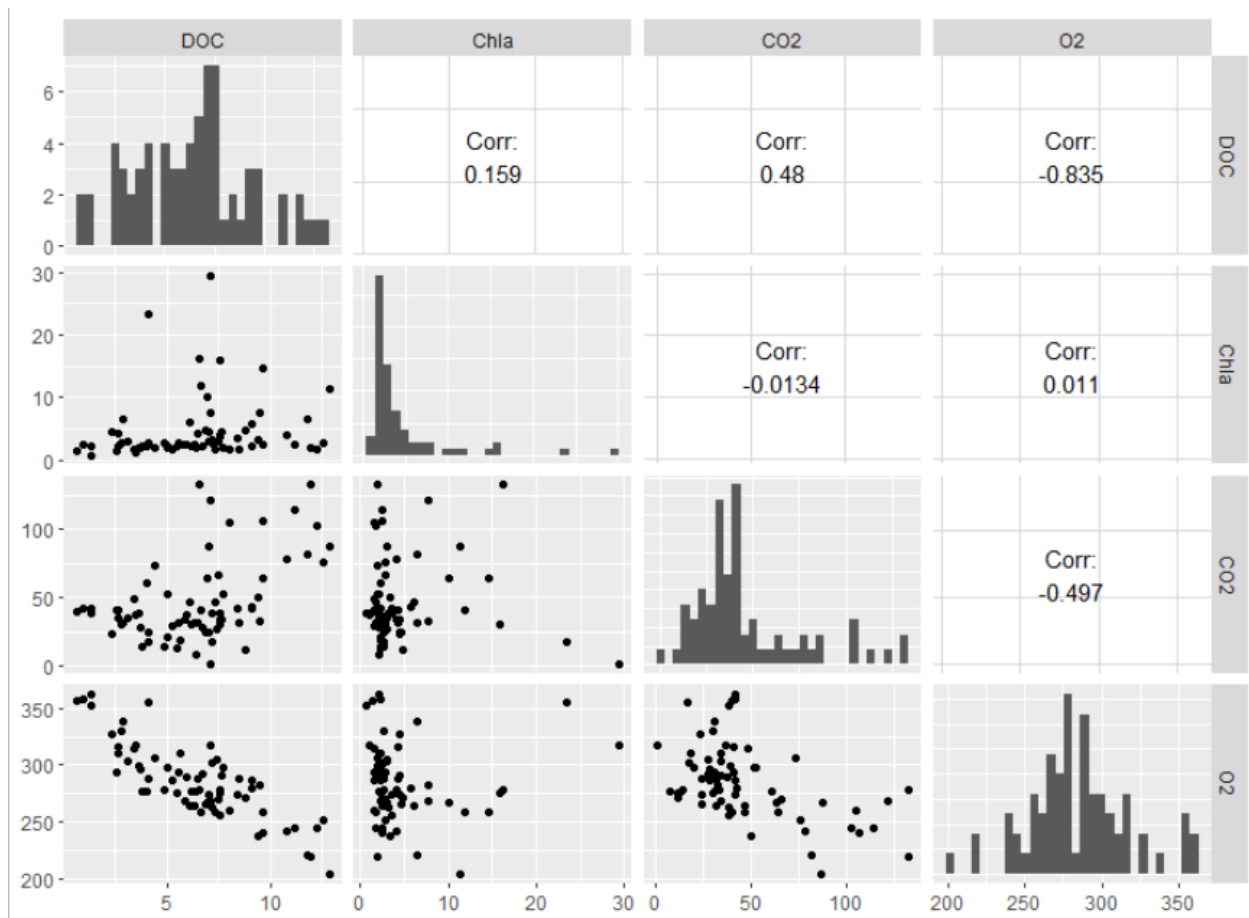


Figure S10. Correlation matrix plot with DOC (mg/L), chlorophyll *a* ( $\mu\text{g L}^{-1}$ ),  $\text{CO}_2$  ( $\mu\text{mol L}^{-1}$ ) and  $\text{O}_2$  ( $\mu\text{mol L}^{-1}$ ) concentrations. There was a negative relation between  $\text{CO}_2$  and  $\text{O}_2$  concentrations ( $r = 0.50$ ) and both  $\text{CO}_2$  and  $\text{O}_2$  were significantly but opposite correlated with TOC concentration ( $r = 0.48$  and  $r = -0.83$  respectively). There was no correlation between  $\text{CO}_2$  or  $\text{O}_2$  concentrations and chlorophyll *a*.

KOEHLER, B., BROMAN, E. & TRANVIK, L. J. 2016. Apparent quantum yield of photochemical dissolved organic carbon mineralization in lakes. *Limnology and Oceanography*, 61, 2207-2221.



# Paper II







# Phosphorus Availability Promotes Bacterial DOC-Mineralization, but Not Cumulative CO<sub>2</sub>-Production

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The current trend of increasing input of terrestrially derived dissolved organic carbon (DOC) to boreal freshwater systems is causing increased levels of carbon dioxide (CO<sub>2</sub>) supersaturation and degassing. Phosphorus (P) is often the most limiting nutrient for bacterial growth and would thus be expected to increase overall mineralization rates and CO<sub>2</sub> production. However, high carbon (C) to P ratios of terrestrially derived DOC could also cause elevated cell-specific respiration of the excess C in heterotrophic bacteria. Using data from a survey of 75 Scandinavian lakes along an ecosystem gradient of DOC, we estimated *in situ* CO<sub>2</sub> production rates. These rates showed a unimodal response with DOC-specific CO<sub>2</sub> production negatively related to DOC:total phosphorus (TP) ratio, and a turning point at 5 mg C L<sup>-1</sup>, indicating higher DOC turnover rates in productive than in unproductive lakes. To further assess the dependency of bacterial respiration (BR) on DOC and P, we monitored CO<sub>2</sub> production in incubations of water with a gradient of DOC crossed with two levels of inorganic P. Finally, we crossed DOC and P with a temperature gradient to test the temperature dependency of respiration rates [as oxygen (O<sub>2</sub>) consumption]. While total CO<sub>2</sub> production seemed to be unaffected by P additions, respiration rates, and growth yields, as estimated by ribosomal gene copy numbers, suggest increased bacterial growth and decreased cell-specific respiration under non-limited P conditions. Respiration rates showed a sigmoid response to increasing DOC availability reaching a plateau at about 20 mg C L<sup>-1</sup> of initial DOC concentrations. In addition to these P and DOC level effects, respiration rates responded in a non-monotonic fashion to temperature with an increase in respiration rates by a factor of 2.6 (±0.2) from 15 to 25°C and a decrease above 30°C. The combined results from the survey and experiments highlight DOC as the major determinant of CO<sub>2</sub> production in boreal lakes, with P and temperature as significant modulators of respiration kinetics.

**Keywords:** dissolved organic carbon-mineralization, lake metabolism, response curves, phosphorus addition, stoichiometry

## INTRODUCTION

Heterotrophic bacteria play a key role in aquatic ecosystems, consuming dissolved organic carbon (DOC) and converting it to carbon dioxide (CO<sub>2</sub>) through bacterial respiration (BR; Del Giorgio et al., 1997; Duarte and Prairie, 2005) and biomass through bacterial production (BP; Del Giorgio and Cole, 1998; Jansson et al., 2006; Berggren et al., 2010).

Bacterial respiration is probably the largest biotic sink of organic carbon on Earth and DOC constitutes a major part of the bulk organic carbon globally (Del Giorgio and Williams, 2005; Drake et al., 2018). Together this makes aquatic bacteria an essential part of the global carbon (C) budget.

The DOC sustaining heterotrophic bacterial metabolism in aquatic ecosystems originates either from primary production within the system (autochthonous DOC) or from terrestrial primary production in the catchment (allochthonous DOC). There is a current trend of increasing transport of terrestrial DOC, to some extent also of total phosphorus (TP) and total nitrogen (TN), to inland waters, caused by factors such as recovery from acidification, climate change, and land use change (Monteith et al., 2007; Kellerman et al., 2015; Finstad et al., 2016; Kritzberg, 2017; Škerlep et al., 2020).

Allochthonous DOC contains a mixture of substances with a variety of molecular size, age, and bioavailability (Neff and Asner, 2001). A large portion of the allochthonous DOC is composed of humic substances, containing aromatic hydrocarbons of high C to nutrient ratios (Mcknight and Aiken, 1998). In humic-rich, low-productive lakes, typical for the boreal zone terrestrially derived substrates often make up the main source of energy and nutrients for bacterial maintenance and growth (Hessen et al., 1990; Karlsson et al., 2007). This decouples the microbial metabolism from the conventional “microbial loop” fueled by autochthonous DOC from algal exudates. In such DOC-rich systems, BR and BP are positively correlated to concentrations of allochthonous DOC rather than to primary production (Jones, 1992; Jansson et al., 2000; Karlsson et al., 2007).

The aromaticity of humic substances causes efficient light absorption and renders DOC prone to photochemical degradation. Allochthonous DOC thus attenuates light and reduces CO<sub>2</sub> uptake through primary production, while promoting CO<sub>2</sub> production through both biological and photochemical mineralization. Allochthonous DOC is a key driver of the in-lake partial pressure of CO<sub>2</sub> (Sobek et al., 2003; Humborg et al., 2010; Larsen et al., 2011). High DOC input renders most boreal lakes net heterotrophic, serving as major conduits of CO<sub>2</sub> to the atmosphere (Hessen et al., 1990; Cole et al., 1994; Sobek et al., 2003).

The share of the total assimilated organic carbon used for BP is given by the bacterial growth efficiency [BGE = BP/(BP + BR)], determining to what degree bacterial metabolism results in bacterial biomass production or in mineralization of organic carbon (Del Giorgio and Cole, 1998). In planktonic communities, BGE varies substantially and has been shown to depend on the quality rather than the quantity of the DOC (Vallino et al., 1996).

Bacterial carbon utilization efficiency is governed by the nutrient to C ratio of the substrate and availability of inorganic nutrients. Bacteria have a high nutrient demand, such that heterotrophic bacteria may dispose of “excess C” under high C-to-nutrient regimes (Hessen, 1992; Hessen and Anderson, 2008). While bacterial metabolism is often limited by C, N, and P, bacterial biomass accumulation is primarily limited by P and N as these are essential building blocks for RNAs and proteins (Sterner and Elser, 2002). However, there is a trade-off in microbial response to substrate C:P ratios. High C:P promotes increased cell-specific respiration, while elevated P support increased growth and biomass accumulation, thus increasing community respiration while the cell-specific respiration still may be reduced and BGE high (Hessen, 1992). Bacterial degradation of DOC at nutrient sufficiency will most likely result in C allocation to bacterial growth, while nutrient limitation may result in higher respiratory rates as the bacteria dispose of excess C (Smith and Prairie, 2004; Hessen and Anderson, 2008; Berggren et al., 2010).

The C to nutrient ratio thus has great implications for the BGE and the cycling and fate of C in a planktonic habitat. Inorganic P is the most frequently reported limiting nutrient for BP (Vadstein, 2000) and it is mainly the P availability that regulates the use of DOC for growth. Experiments of adding DOC and inorganic P to oligotrophic lake waters have shown that low BGE's accompanying increased C:P ratios do not necessarily mean that BP is decreasing but rather that BR is increasing (Jansson et al., 2006).

Together with increased run-off and biomass production, air and water temperatures are increasing with the ongoing climate warming (Schneider and Hook, 2010; O'Reilly et al., 2015). Temperature has a fundamental role in regulating the activity and growth of microorganisms (Farrell and Rose, 1967; Madigan et al., 1997). While it can be broadly stated that metabolism increases with temperature up to a certain level, the rate of the exponential increase differs between organisms, reactions, and temperature ranges. Although the rates of both BP and BR increase with temperature, several studies have reported that the temperature dependency of BR is stronger than that of BP and as a consequence, BGE has often been shown to decrease at increasing temperatures (Rivkin and Legendre, 2001; Apple et al., 2006; Berggren et al., 2010; Kritzberg et al., 2010).

Furthermore, the temperature dependency of bacterioplankton metabolic rates interacts with the substrate quantity and quality. Metabolic rates have been shown to be less temperature dependent for heterotrophic bacteria growing on labile autochthonous DOC than when growing on complex and recalcitrant allochthonous DOC (Ylla et al., 2012; Jane and Rose, 2018). In aquatic systems with a DOC pool heavily influenced by terrestrial inputs, increased temperatures are expected to further enhance BR, expanding the role of heterotrophic bacteria as CO<sub>2</sub> conduits to the atmosphere.

Microbial mineralization of DOC thus depends on several interacting factors. Although we can expect that increased loadings of terrestrially derived DOC and nutrients and enhanced temperatures increase bacterial growth and

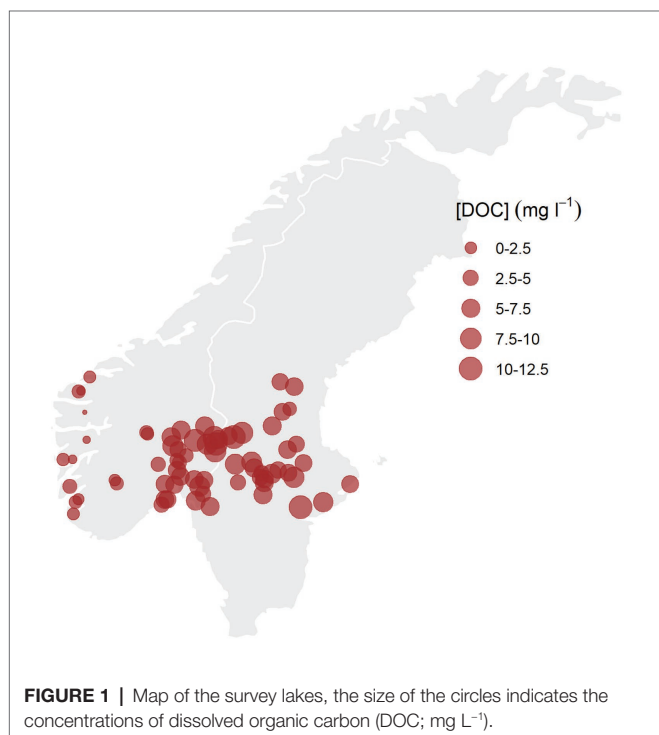
metabolism, more studies are needed to elucidate how the different environmental factors interact.

In this study, we used chemical and physical data from 75 Scandinavian lakes to estimate in-lake CO<sub>2</sub> production. The lakes spanned close to orthogonal ecosystem gradients in DOC and TP, allowing us to assess the interactive effects of these two parameters on CO<sub>2</sub> production. To test for dynamic responses of bacterioplankton respiration to allochthonous DOC concentrations, nutrient availability, and temperature, we additionally performed experimental incubations. During 1-week incubations, we monitored respiration in two experimental set-ups, one addressing CO<sub>2</sub> production and the other O<sub>2</sub> consumption. A gradient of DOC concentrations was achieved by adding natural organic matter (NOM; isolates from a Norwegian humic lake obtained through reverse osmosis) to clear lake water.

## MATERIALS AND METHODS

### Field Sites

During July and August of 2011, a set of 75 large lakes spread out over a geographical gradient from western Norway to eastern Sweden was sampled (Figure 1). The lakes were chosen to represent wide and close to orthogonal gradients in dissolved organic matter (DOM) and TP. To avoid strong temperature gradients the lakes were chosen within a narrow latitudinal and altitudinal range. All lakes met the following criteria: latitude 57–64°N, altitude <600 m, surface area > 1 km<sup>2</sup>, pH > 5, TP < 30 μg L<sup>-1</sup>, and DOC <30 mg L<sup>-1</sup>. The lakes were sampled by plane in a synoptic survey,



and composite samples of a total of 15 L were taken from 0 to 5 m in the central part of each lake during daytime, using an integrating water sampler (Hydro-BIOS, Germany). Vertical temperature profiles and vertical profiles of scalar irradiance (see SI for more detail) were measured using XRX-620 10-channel CTD (RBR Ltd., Canada). Vertical temperature profiles indicated that the thermocline was deeper than 5 m in all lakes. The integrated 0–5 m samples could thus be considered representative of the mixed layers of the lakes.

### Laboratory Analyses of Lake Samples

Concentrations of TP, total organic carbon (TOC), and TN were measured in two accredited laboratories, at the Norwegian Institute for Water Research (NIVA) and at the University of Oslo (UiO). Total inorganic carbon (TIC) was measured at UiO (see SI and Thrane et al., 2014 for details).

Dissolved CO<sub>2</sub> and O<sub>2</sub> were measured as headspace concentrations in acid (0.2% HgCl) fixed samples by gas chromatography (GC) analysis (see SI and Yang et al., 2015 for details). Chlorophyll *a* (chl *a*) concentration was measured in two different ways, both by high performance liquid chromatography (HPLC; Schagerl and Donabum, 2003) and by fluorescence spectrometry after extraction in 96% ethanol. The averages of the two methods (which generally matched well) were used in further analyses. Chromophoric dissolved organic matter (CDOM) optical density [ODCDOM(λ)] of 20 ml filtered lake water (Acrodisc 0.2 μm polyethersulfone membrane syringe filter, Pall Life Sciences) was measured from 400 to 750 nm in steps of 1 nm. According to Mitchell et al. (2002), from which we calculated the absorption coefficient spectra of CDOM [aCDOM(λ); m<sup>-1</sup>].

Area-specific primary production (PP<sub>A</sub>; mg C m<sup>-2</sup> d<sup>-1</sup>) was calculated using a bio-optical model based on lake-specific phytoplankton absorption coefficients, daily *in situ* irradiance, and the light dependent quantum yield of photosystem II measured by a Pulse Amplitude Modulated (PAM) fluorometer (AquaPen-C 100, PSI Czech Republic; for details see SI and Thrane et al., 2014). As PP<sub>A</sub> here is a measure of CO<sub>2</sub> consumption, we can note it as the CO<sub>2</sub> flux from water to primary producers F<sub>PP</sub>.

Water-air flux of CO<sub>2</sub> (F<sub>net</sub>) represents the net degassing of CO<sub>2</sub> from the surface and was calculated from surface CO<sub>2</sub> concentrations of each lake using Fick's law of diffusion.

$$F_{net} = k_{CO_2} \Delta CO_2 \quad (1)$$

where the CO<sub>2</sub> gas exchange (k<sub>CO<sub>2</sub></sub>) coefficient was obtained according to Jähne et al. (1987) and Cole and Caraco (1998; for details see SI and Yang et al., 2015).

### Total CO<sub>2</sub> Production

From the dataset, it was not possible to distinguish between lateral input of CO<sub>2</sub> (F<sub>lat</sub>; surface and ground water inflow) and in-lake production of CO<sub>2</sub> (F<sub>min</sub>; microbial and photochemical mineralization of DOC). The sum of in-lake DOC mineralization and lateral input was therefore used as an estimate of total CO<sub>2</sub> production (F<sub>tot</sub> = F<sub>lat</sub> + F<sub>min</sub>). Assuming a steady state

of the CO<sub>2</sub> saturation deficit, the mass balance due to production, lateral input, consumption, and evasion can be written as:

$$F_{net} = F_{tot} - F_{PP} \quad (2)$$

And therefore

$$F_{tot} = F_{min} + F_{lat} = F_{net} + F_{PP} \quad (3)$$

## Experimental Design

To investigate the effects of DOC and P additions on bacterioplankton respiration in more detail, we incubated water samples with a gradient in dissolved NOM concentrations crossing it with two levels of inorganic P. Samples were incubated in the dark using two different experimental set-ups and methods. During the incubations, we monitored bacterioplankton respiration through measurements of either CO<sub>2</sub> production or O<sub>2</sub> consumption, depending on the set-up. Samples for microbial biomass were taken upon termination of the experiments with the aim of assessing biomass from a flow-cytometer equipped with a plate reader set-up. However, despite trying with different stains and protocols, the samples were obscured by background scatter from the added DOC, and hence did not provide reliable counts. Ideally, the dynamic responses in CO<sub>2</sub> and O<sub>2</sub> should have been verified not only with bacterial counts but also assessment of community response by genetic screening and transcriptomics. This does, however, require a different set-up with larger volumes and a more frequent sampling regime that does not compromise the semi-continuous gas analysis. With the current set-up, we prioritized the gas analysis as the ultimate response output but assessed final microbial biomass from quantitative PCR (qPCR) on filtered samples when terminating the experiment.

## SOURCE OF DOM

The DOM gradient was obtained using a NOM isolates from the DOC-rich boreal bog/lake Hellerudmyra close to Oslo, produced within the NOM-typing and the NOMiNiC projects (see Gjessing et al., 1999 and Vogt et al., 2001 for isolation protocol and characterization). This is a powder material up-concentrated from freshwater through reverse osmosis and isolated by freeze-drying of the concentrate. While also containing non-humic material, the major fraction of the NOM isolate consists of humic substances. The carbon fraction of the isolate is 33.7%. The NOM powder was mixed in deionized water to a stock solution of DOM with a DOC concentration of 1,000 mg C L<sup>-1</sup> and filter sterilized through a 0.2 μm pore size Supor membrane filter (Gelman, CO, USA). While the isolation and up-concentration of the material enriches various elements, the DOM retains its “natural” properties (Gjessing et al., 1999; Vogt et al., 2001), and should be superior to artificial sources of DOC.

## Preparation of the Media

The DOM stock solution was diluted to the desired DOC concentrations in sterile filtered (0.2 μm pore size Supor

membrane filter; Gelman, CO, United States) drinking water from the tap. This water comes from the oligotrophic and pristine lake Maridalsvannet in the municipality of Oslo, Norway. The water is treated following protocols for drinking water. The processing includes alkalization/carbonization by marble and CO<sub>2</sub>, coagulation and particle separation in Actiflo followed by two media filter, and UV irradiation for disinfection. Besides disinfecting, the UV treatment also lowers the CDOM content in the water. The treatments do not eliminate essential trace metals and macronutrients in the water, giving background concentrations of DOC and total N of 2 mg L<sup>-1</sup> and 0.01 mg L<sup>-1</sup>, respectively, while the background total P was below detection limits.

The samples were inoculated with 1% of the total sample volume of fresh water from a stream draining Hellerudmyra from where the DOM was isolated. The inoculum was filtered (2 μm) to remove large particles and protists.

## Experimental Set-Up 1: CO<sub>2</sub> Production

We used 14 levels of DOC additions between 0 and 50 mg L<sup>-1</sup> (0 mg L<sup>-1</sup>, 2.5 mg L<sup>-1</sup>, 5 mg L<sup>-1</sup>, 7.5 mg L<sup>-1</sup>, 10 mg L<sup>-1</sup>, 12.5 mg L<sup>-1</sup>, 15 mg L<sup>-1</sup>, 17.5 mg L<sup>-1</sup>, 20 mg L<sup>-1</sup>, 22.5 mg L<sup>-1</sup>, 25 mg L<sup>-1</sup>, 34 mg L<sup>-1</sup>, 41 mg L<sup>-1</sup>, and 50 mg L<sup>-1</sup>). The DOC gradient was crossed with two levels of PO<sub>4</sub>-P additions (0 and 2 μmol L<sup>-1</sup>). To make sure that N was not limiting, 30 μmol L<sup>-1</sup> each of NO<sub>3</sub>-N and NH<sub>4</sub>-N, resulting in a total of 60 μmol L<sup>-1</sup> N, were added to all samples. In samples with P additions, the C:N:P ratio thus ranged between 82:30:1 and 2160:30:1.

The incubations were carried out in the dark in a temperature-controlled water bath at 20°C. Samples of 50 ml were transferred into 122 ml, acid washed glass vials equipped with acid washed magnetic stirrers. The vials were crimp sealed with butyl rubber septa. Before incubation, the vials were washed with HeO<sub>2</sub> (80/20%) by 6 cycles of evacuation and filling using a manifold, while stirring the samples at 400 rpm to remove CO<sub>2</sub> and nitrogen gas (N<sub>2</sub>). The samples were incubated constantly stirred (400 rpm) for 10 days at 20°C, while CO<sub>2</sub> production and O<sub>2</sub> uptake were measured automatically by GC every 6 h using the robotized set-up described by Molstad et al. (2016) with some modifications. Since these vials contained 50 ml water and 70 ml air, the O<sub>2</sub> uptake was small relative to the large amount of O<sub>2</sub> in the headspace and we could not measure uptake rates with sufficient precision.

## Experimental Set-Up 1: Ribosomal RNA Gene Copy Numbers

To approximate bacterial growth yields we measured bacterial 16S ribosomal RNA (rRNA) gene copy numbers using a qPCR protocol (Savio et al., 2015). In short, 40 ml of water from each incubation were filtered through 0.2 μm Supor PES membrane filters (Pall Corporation, CA, United States) at the end of the experiment. Filters were stored at -80°C until DNA extraction was performed using the Dneasy PowerSoil kit as recommended by the manufacturer (Qiagen, Germany). Total DNA concentration was assessed



using a Qubit ds DNA Broad-Range Assay (London, United Kingdom), and 16S rRNA genes were quantified using a bacteria-specific qPCR. qPCR reactions contained 2.5  $\mu$ l DNA extract as the template and 0.2  $\mu$ M each of the primers 8F and 338 (Frank et al., 2007; Fierer et al., 2008) targeting the V1-V2 region of most bacterial 16S rRNA genes and iQ SYBR Green Supermix (Bio-Rad Laboratories, Hercules, USA). Samples were run in triplicates together with a dilution series of the ZymoBIOMICS Gut Microbiome Standard (Zymo Research, Irvine, USA) to obtain 16S rRNA gene copy numbers in each incubation.

## Experimental Set-Up 2: Dissolved Oxygen Consumption

The second experimental set-up was designed with fewer DOC levels (0 mg L<sup>-1</sup>, 25 mg L<sup>-1</sup>, and 50 mg L<sup>-1</sup>) plus an additional crossing with four temperatures (10, 15, 25, and 30°C). All treatments were run in quadruplicates.

The incubations were carried out in the dark, placed in a water bath in a climate chamber to assure stable temperature. Dissolved oxygen concentrations were measured over 7 days of incubation with a SensorDish Reader (SDR; resolution:  $\pm 0.4\%$  O<sub>2</sub> at 20.9% O<sub>2</sub>, PreSens GmbH, Regensburg, Germany) using non-invasive fluorescence sensor spots placed in the bottoms of 5 ml vials, which were measured by optodes. The vials were washed with 70% ethanol and baked at 80°C for 10 h before filling them with sample and leaving them in the incubator for temperature equilibration for 2 h. After the equilibration, we inoculated the samples and filled the vials to the top, leaving no headspace. The vials were then closed and additionally sealed with parafilm to avoid gas diffusion. Vials with deionized water were used as controls to check for O<sub>2</sub> leakage. Dissolved oxygen concentrations were recorded automatically every 15 s. O<sub>2</sub> consumption rates were may represent CO<sub>2</sub> production rates, assuming a respiratory quotient (RQ: mole CO<sub>2</sub> produced/mole O<sub>2</sub> consumed) equal to 1. Although humic-rich substances are completely oxidized at an RQ of 0.9 (Dilly, 2001), anabolic processes contributes to higher RQs than catabolic respiration alone (Dilly, 2003; Berggren et al., 2012). We therefore believe that an RQ of 1 is an appropriate assumption. However, in the analysis, we use the rates of O<sub>2</sub> consumption that indirectly represent CO<sub>2</sub> production rates (see below), and thus the RQ value does not affect the results. In some treatments, the O<sub>2</sub> was depleted toward anoxia, but this should not affect our analysis since the initial slopes of the uptake curves, when the incubations still were oxic, to estimate bacterial growth.

Temperature sensitivity of the respiration rates were analyzed using the  $Q_{10}$  coefficient as the relative change in rate when increasing the temperature by 10°C.

$$Q_{10} = \left( \frac{R_2}{R_1} \right)^{\left( \frac{10}{T_2 - T_1} \right)} \quad (4)$$

where R is the rate, here the respiration rate ( $\mu$ mol CO<sub>2</sub> L<sup>-1</sup> h<sup>-1</sup>) and T is the temperature in centigrade.

## Modeling the Respiration Curves

Respiration curves and growth rates were modeled using the packages *mgcv* (Wood, 2011) and *gratia* (Simpson, 2018) in R (R Core Team, 2020). A generalized additive model (*gam*) with simple factor smoothers on time and grouped by experimental unit was fitted to all the measured time-series data. The fitted curves were then differentiated, using the derivatives function from the *gratia* package (Simpson, 2018) to estimate the time course of the net rate of change in each experimental unit. From the fitted derivatives we calculated maximum O<sub>2</sub> consumption or CO<sub>2</sub> production rates for each experimental unit. The time until the maximum rate was reached was used as a measure of the lag phase.

## Statistics

All data analysis was performed using the open-source software R version 4.0.2 (R Core Team, 2020). Lake variables were checked for normality and log-transformed where needed. Correlations are reported using Pearson's correlation coefficients and all error estimates are given in standard errors. For the statistical modeling, we used the *mgcv* package (Wood, 2011) fitting *gam* models for prediction of the dependent variables. To test the dependency of total CO<sub>2</sub> production ( $F_{\text{tot}}$ ; mg C m<sup>-2</sup> d<sup>-1</sup>), we used a *gam* model with smoothers on each of the explanatory variables DOC (mg L<sup>-1</sup>), TP ( $\mu$ g L<sup>-1</sup>), TN (mg L<sup>-1</sup>), TIC (mg L<sup>-1</sup>), SUVA<sub>400</sub> (L mg-C<sup>-1</sup> m<sup>-1</sup>), and temperature (°C). Predictive variable selection was done by applying additional shrinkage on the null space of the penalty with the *select = TRUE* argument in the *mgcv::gam* function, as recommended by Marra and Wood (2011). The resulting model has all smoothers that are not necessary for the fit as close to zero as possible.

For the analysis of the experimental results, individual *gam* models were fitted to samples with and without P additions (and the three levels of DOC concentrations in the second experiment), respectively. In the case of experimental set-up 1, *gams* were fitted with DOC concentration as the explanatory variable, while in experimental set-up 2 temperature was used. We also performed an analysis of covariance to test for treatment effects of P (experimental set-up 1) or P and temperature (experimental set-up 2).

## RESULTS

### Lake Survey

Concentrations of both DOC and TP varied largely among lakes, with DOC concentrations ranging between 0.25 and 12.9 mg L<sup>-1</sup> and TP between 0.5 and 27.5  $\mu$ g L<sup>-1</sup>. Although lakes had been selected for orthogonality between DOC and TP, there was a correlation between the two variables ( $p < 0.001$ ,  $r = 0.61$ , log-log). Still a considerable scatter indicates reasonable orthogonality. Other important cross-correlations that need to be considered in data interpretation are positive relationships of TN with both TP and DOC concentrations (Supplementary Figure S1).

Dissolved CO<sub>2</sub> concentrations spanned two orders of magnitude (0.82–133  $\mu$ mol L<sup>-1</sup>) with the majority of the lakes

being strongly supersaturated with  $\text{CO}_2$  as indicated by an average  $\text{CO}_2$  of more than twice that at atmospheric equilibrium. This supersaturation conveys evasion of  $\text{CO}_2$  to the atmosphere from most lakes and net-heterotrophy of the lake systems during the sampling campaign. As expected, the level of  $\text{CO}_2$  saturation was positively correlated with DOC concentration ( $p < 0.001$ ;  $r = 0.52$ ) and negatively correlated with  $\text{O}_2$  saturation ( $p < 0.001$ ;  $r = -0.69$ ; **Supplementary Figure S2**) and thus reflects the role of DOC as a driver of heterotrophy. There was no correlation between TIC and  $\text{CO}_2$  saturation deficit, suggesting that in-lake processes were the main cause of  $\text{CO}_2$  supersaturation.

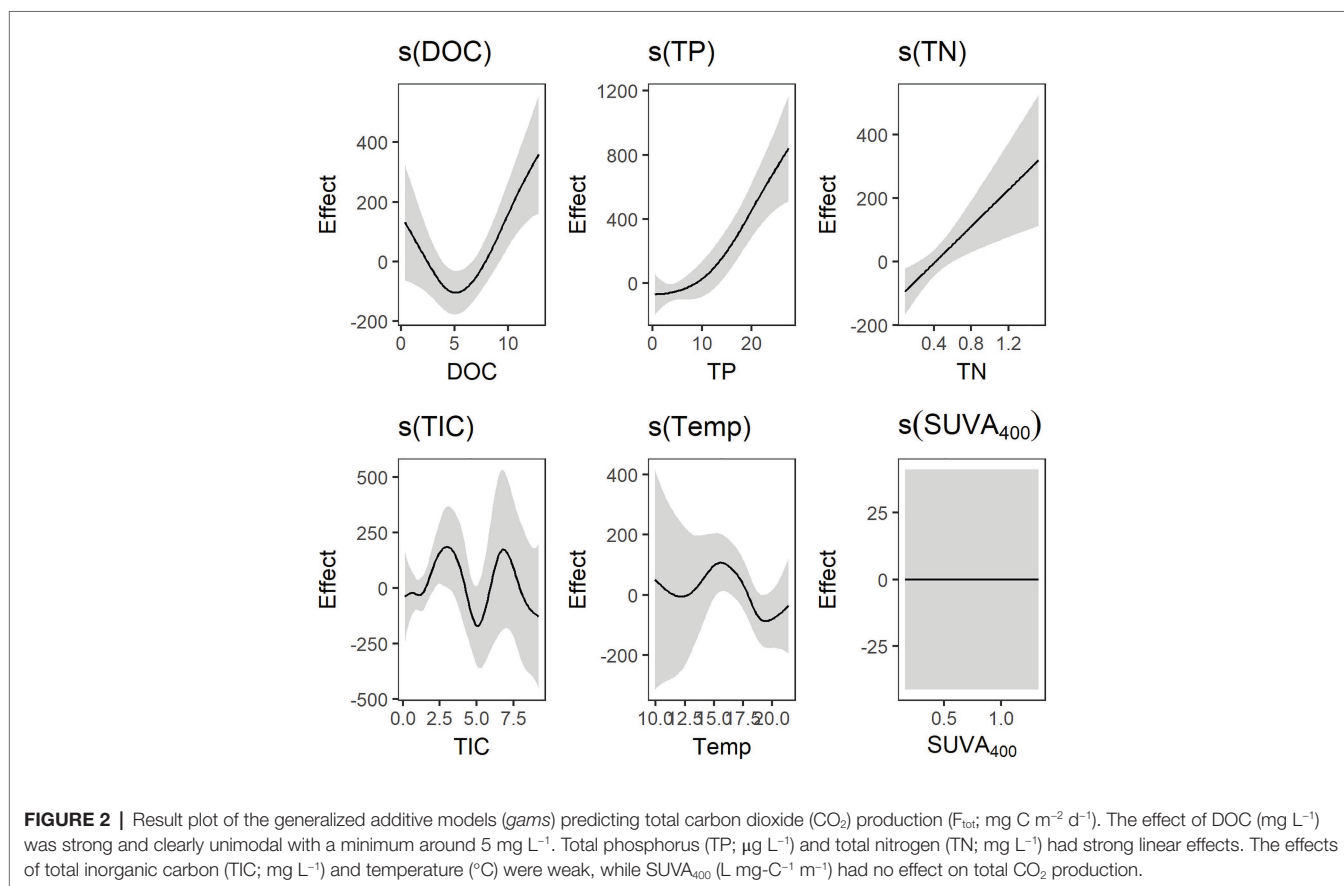
Areal primary production ( $\text{PP}_A$ ) was not significantly correlated to  $\text{CO}_2$  concentrations, providing no support for primary production being boosted by  $\text{CO}_2$  in  $\text{CO}_2$  rich lakes. Still,  $\text{PP}_A$  was weakly but negatively related to DOC concentrations ( $p = 0.05$ ;  $r = -0.23$ , log-log; Thrane et al., 2014), and somewhat (but non-significant) lower in high  $\text{CO}_2$  lakes than in lakes with low  $\text{CO}_2$  concentrations. In addition,  $\text{PP}_A$  was negatively related to the DOC: TP ratio ( $p < 0.001$ ;  $r = -0.60$ ; **Supplementary Figure S3A**). Total  $\text{CO}_2$  ( $F_{\text{tot}}$ ) production here represents both  $\text{CO}_2$  produced in lakes and  $\text{CO}_2$  coming into lakes from the surroundings *via* ground water and run-off and was estimated as the sum of net  $\text{CO}_2$  evasion and a real primary production ( $F_{\text{tot}} = F_{\text{min}} + F_{\text{lat}} = F_{\text{net}} + F_{\text{pp}}$ ). The gam model explained 77% of the total deviance in total  $\text{CO}_2$  production with strong

effects of DOC, TP, and TN. TIC and temperature had weak effects and SUVA had no effect on total  $\text{CO}_2$  production (**Figure 2**). Relating total  $\text{CO}_2$  production to DOC concentration revealed a unimodal response to increased DOC concentrations with a minimum value at around  $5 \text{ mg L}^{-1}$  (**Figure 2**). Carbon concentration-specific  $\text{CO}_2$  production [ $F_{\text{tot}}$ : ( $\text{mg C m}^{-2} \text{ d}^{-1}$ )/ $\text{DOC}(\text{mg C m}^{-2})$ ] was positively related to  $\text{PP}_A$  ( $p < 0.001$ ,  $r = 0.60$ , log-log) and was thus, similar to  $\text{PP}_A$ , decreasing with an increasing DOC:TP ratio (**Supplementary Figure S3B**).

## Incubation Experiments

To test for dynamic responses underlying the patterns observed across lakes, two experimental incubations were performed, one addressing the production of  $\text{CO}_2$ , and the other the consumption of  $\text{O}_2$ . Both incubations were performed in the dark, hence they reflect heterotrophic microbial mineralization.

In the  $\text{CO}_2$  production experiment, DOC concentration had a positive effect on  $\text{CO}_2$  production in all samples, regardless of P level. However, P had a strong effect on the kinetics of  $\text{CO}_2$  production and hence the shape of the  $\text{CO}_2$  accumulation curves (**Supplementary Figure S4**). While all treatments showed non-linear  $\text{CO}_2$  accumulation similar to logistic growth, the exponential phase was more distinct and steeper in the treatments receiving P additions.  $\text{CO}_2$  production was faster and leveled off earlier and more distinctively in P-spiked than in P-limited samples. This effect was most pronounced for initial DOC



concentrations up to 25 mg L<sup>-1</sup>. The lag phase was more pronounced in P-spiked samples but decreased in length with increasing DOC concentrations. At DOC concentrations >25 mg L<sup>-1</sup>, CO<sub>2</sub> concentrations kept increasing at a slower pace after an initial exponential phase without reaching a plateau (**Supplementary Figure S4**).

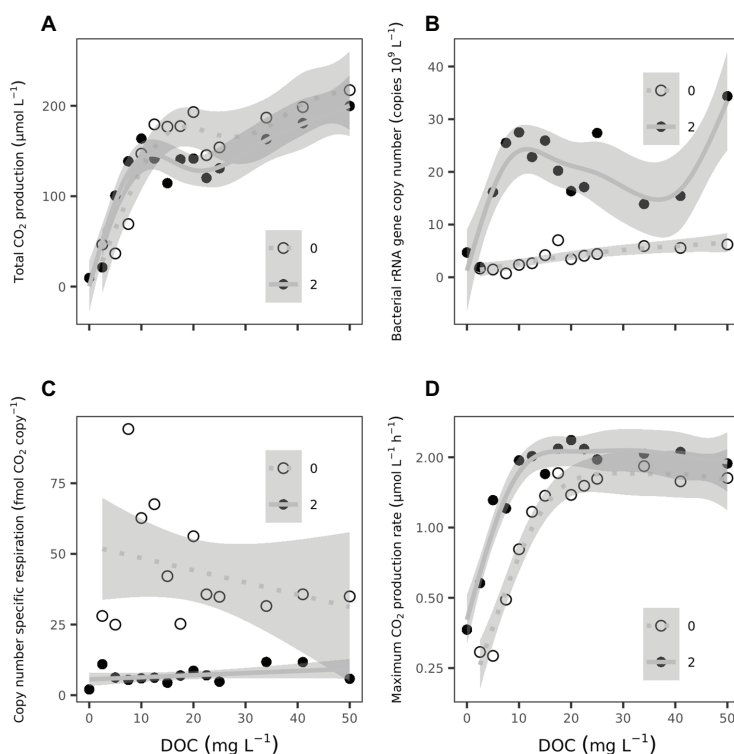
The total amount of CO<sub>2</sub> produced during the laboratory incubation increased monotonously with initial DOC concentration until reaching a threshold value (~10 mg L<sup>-1</sup>), above which cumulative CO<sub>2</sub> production appeared to be independent of the amount of DOC supplied, before increasing again with DOC concentration above 30 mg L<sup>-1</sup>, albeit at lower rate (**Figure 3A**). Treatments with P additions had a somewhat lower threshold value than treatments without P additions. Still, the total amount of CO<sub>2</sub> produced during the incubations were similar in samples with or without P additions (**Supplementary Table S1**), while higher rRNA gene copy numbers were observed in P-spiked samples (**Figure 3B**; **Supplementary Table S1**). Consequently, estimates of rRNA gene copy number-specific respiration, used here as a proxy for bacterial growth yield, were lower in P-spiked than non-spiked samples (**Figure 3C**; **Supplementary Table S1**), indicating less respiration per unit biomass produced and hence larger growth yields due to P addition. There was also an increase in rRNA gene copy numbers with increasing DOC concentrations in

P-spiked and non-spiked samples (**Figure 3B**), while estimates of rRNA gene copy number-specific respiration showed no clear trends with regards to DOC concentrations (**Figure 3C**).

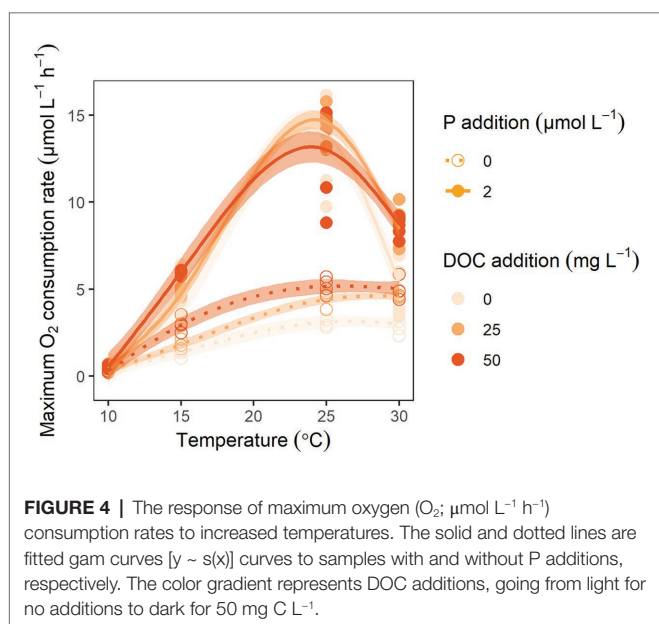
Similar to total CO<sub>2</sub> production, the maximum observed respiration rates at any time during the incubation, increased steadily with increasing DOC up to 20 mg L<sup>-1</sup> (**Figure 3D**). Below this threshold value, maximum inducible respiration rates were clearly higher in P spiked than in P limited samples. Maximum inducible respiration rates increased by about 9% for each mg DOC L<sup>-1</sup> up to 20 mg L<sup>-1</sup>, regardless of P level. At DOC concentrations >20 mg L<sup>-1</sup>, the increase in respiration rates with increasing DOC halted abruptly and stayed constant with similar rates at the two P levels.

The fraction of the DOC pool that was respired to CO<sub>2</sub> during the course of the experiments was similar across the P levels, but decreased with DOC concentration. At concentrations above the threshold of 20 mg L<sup>-1</sup>, the respired fraction of the DOC pool was about 5% (**Supplementary Figure S6**).

In the O<sub>2</sub> consumption experiment, the dynamic response in O<sub>2</sub> uptake to DOC and P as well as temperature was tested. Similar to the CO<sub>2</sub> production curves, the shapes of the O<sub>2</sub> consumption curves differed substantially depending on P level with a more pronounced exponential phase in samples receiving P additions than in samples without P additions (**Supplementary Figure S5**). Respiration rate was also highly dependent on



**FIGURE 3** | The response of **(A)** total CO<sub>2</sub> production (μmol L<sup>-1</sup>) during the entire incubation (240 h), **(B)** yield in bacterial 16S ribosomal RNA (rRNA) gene copy number (copies L<sup>-1</sup>), **(C)** bacterial 16S rRNA gene copy number-specific respiration (fmol CO<sub>2</sub> copy<sup>-1</sup>), and **(D)** maximum CO<sub>2</sub> production rates (μmol L<sup>-1</sup> h<sup>-1</sup>) to increased DOC concentrations (mg L<sup>-1</sup>). The solid and dotted lines are fitted *gams* to samples with and without phosphorus (P) additions, respectively, with shaded areas representing the confidence interval of the *gams*. Statistics of the *gams* can be found in **Supplementary Table S1**.



temperature (Figure 4). Maximum  $O_2$  consumption rates were mainly related to P level and temperature (Supplementary Table S1), while the regression estimate of the DOC concentration effect was non-significant (Supplementary Table S1).

Respiration rates in the  $10^\circ\text{C}$  incubation were close to zero, possibly reflecting that respiration had not started to increase before the incubation was stopped and that the incubation time thus was too short with the lowest temperature. Respiration rates increased from 15 to  $25^\circ\text{C}$  with a  $Q_{10}$  of  $2.6 (\pm 0.2)$  and no significant difference in  $Q_{10}$  between samples of different P levels. From 25 to  $30^\circ\text{C}$ , the respiration rates decreased with a  $Q_{10} < 1$ . Here the  $Q_{10}$  also differed between P spiked ( $0.44 \pm 0.09$ ) and P limited ( $0.94 \pm 0.05$ ) samples.

## DISCUSSION

### DOC and P Availability Regulating Total $\text{CO}_2$ Production in Lakes

In this study, we analyzed data from a lake survey comprising 75 Scandinavian lakes chosen to represent gradients in DOM and P concentrations. Chemical and physical data from the lakes were used to estimate total  $\text{CO}_2$  production. As it was not possible to distinguish between in-lake production and lateral input of  $\text{CO}_2$  (from inflowing rivers or groundwater input), total  $\text{CO}_2$  production was used to lump both sources. Notwithstanding, we found a strong relationship between  $O_2$  and  $\text{CO}_2$  saturation deficits ( $r = -0.70$ ; Supplementary Figure S2). The intercept was not significantly different from zero and lakes that were saturated with  $O_2$  were thus also saturated with  $\text{CO}_2$ , indicating that microbial respiration was the predominant source of  $\text{CO}_2$  in the lakes. Furthermore, we found no correlation between TIC and  $\text{CO}_2$  deficit, suggesting that processes within the lakes rather than lateral input regulate the  $\text{CO}_2$  supersaturation. Although some studies have shown that DIC input from the catchment plays a

larger role for explaining  $\text{CO}_2$  evasion from lakes than previously thought (Maberly et al., 2013; Leith et al., 2015; Weyhenmeyer et al., 2015), the largest contributor to  $\text{CO}_2$  supersaturation in the studied lakes was most probably microbial mineralization.

The observed unimodal response of total  $\text{CO}_2$  production to increased DOC concentrations (Figure 2), however, suggests a shift in substrate from mainly autochthonous to predominantly allochthonous DOC. Autochthonous DOC is generally more bioavailable and of higher nutritious value with a lower C:P ratio (Søndergaard et al., 1995). When DOC of both phytoplankton and terrestrial origin is available, heterotrophic bacteria prefer the former as substrate for catabolic processes (Kritzberg et al., 2004). Estimated  $\text{CO}_2$  production rates decreased with increasing DOC concentration until a minimum was reached at around  $5 \text{ mg L}^{-1}$ . Correspondingly, primary production rates are commonly reported to increase with DOC concentrations until around  $5 \text{ mg C L}^{-1}$ , after which the rates are declining (Karlsson et al., 2007; Seekell et al., 2015; Tanentzap et al., 2017). This unimodal response is likely reflecting a trade-off between nutrients associated with DOC and the increasing light attenuation caused by CDOM. Modest increases in DOC may also be beneficial by blocking out short-wave UV-radiation (Palen et al., 2002). Above  $5 \text{ mg L}^{-1}$ , an increasing portion of the DOC pool is of terrestrial origin and  $\text{CO}_2$  production rates thus increase linearly with increasing DOC concentrations.

The DOC concentration-specific  $\text{CO}_2$  production, i.e., the rates of  $\text{CO}_2$  production per unit of DOC concentration, was positively related to  $PP_A$ , indicating that a larger share of the DOC pool was respired in more productive lakes. The DOC:TP ratio had a negative effect on  $PP_A$ , and consequently, the DOC:TP ratio also had a negative effect on the DOC concentration-specific  $\text{CO}_2$  production (Supplementary Figure S3). This may seem to contradict the notion that BR increases with increased C:P ratios (Jansson et al., 2006). However, on a community level low BGE at high C:P ratios has been coupled to decreasing BP rates rather than increasing BR rates (Smith and Prairie, 2004). Higher DOC concentration-specific  $\text{CO}_2$  production indicates faster DOC turnover in the low than in the high C:P lakes. A larger share of the DOC pool is degraded, probably accompanied with higher bacterial density in productive than in unproductive lakes. This implies a more bioavailable DOC pool in productive than in unproductive lakes, and could also suggest that this is explained by a lower C:P ratio of the substrate.

### Experimental Validation of Drivers

While lake gradients may provide general patterns, the mechanistic drivers can only be revealed experimentally. To disentangle the role of DOC relative to P, we conducted two experiments. First, we measured BR along a gradient of DOC concentrations crossed with two levels of inorganic P concentrations. Since this DOC was an isolate from a humic lake (see section "Materials and Methods"), it represented primarily allochthonous C. We found clear differences in the kinetics of  $\text{CO}_2$  production between P spiked and P-limited samples. The  $\text{CO}_2$  accumulation curves of P-spiked samples



showed a pronounced exponential phase until reaching a plateau, similar to a bacterial growth curve reaching substrate limitation (**Supplementary Figure S4**). The kinetic patterns suggested that P-spiking boosted respiratory rates leading to substrate limitation earlier during incubation. The longer lag phase in P-spiked samples could be explained by a major shift in community composition. P-limited samples would represent a situation with minor shifts in community composition. The rRNA gene copy number-specific respiration suggests that cell-specific respiration increased under P-limited conditions. Accordingly, Smith and Prairie (2004) found cell-specific respiration to be negatively related to P supply. They further report that on a per cell basis, BR explained the greatest amount of variation in BGE. This would suggest a higher BGE in P-spiked than in P-limited samples in our experiment, provided a higher BR cell<sup>-1</sup> in P-limited compared to P-spiked samples, as reflected by the lower rRNA gene copy number-specific respiration in P-spiked samples. More P, however, also releases the bacteria from P-limitation, hence causing higher metabolic activity. The balance between respiration due to excess C (i.e., under high substrate C:P) or respiration powered by increased metabolic activity under elevated P (i.e., low C:P) is not straight forward, since increased P also would stimulate bacterial growth and thus community metabolism. This unpredictability is corroborated by our results as rRNA gene copy number-specific respiration in non-spiked samples showed a negative trend with increasing DOC concentrations, while under P addition a significant increase in rRNA gene copy number-specific respiration could be observed. Previous studies with C and P manipulations, showed good correspondence between bacterial biomass and CO<sub>2</sub> production (Hessen et al., 1994), but in the absence of reliable day-to-day microbial counts we cannot fully resolve this stoichiometric response at the cellular versus the community level.

The cumulative amount of CO<sub>2</sub> produced during the incubation was higher in P-spiked than in P-limited samples at low initial DOC concentrations (<10 mg L<sup>-1</sup>; **Figure 3A**). In the intermediate DOC range (12.5–25 mg L<sup>-1</sup>), however, this trend shifted and more of the available DOC was respired in P-limited than in P-spiked samples (**Supplementary Figure S4**). The larger cumulative CO<sub>2</sub> production in P-limited samples above ~20 mg L<sup>-1</sup> initial DOC could reflect that the slow-growing population assimilated the substrate at a slower pace and that the major share of the assimilated DOC was used for maintenance and hence was respired. However, this is not corroborated by rRNA gene copy number-specific respiration. The bioavailable fraction of DOC was consumed more rapidly when P was available than when P was limiting, reflecting faster DOC turnover similar to the findings in the lake survey.

We added a fixed amount of naturally isolated DOC at the beginning of the experiment, and microbial assimilation and respiration depleted a large share of the bioavailable fraction of this DOC during the course of the experiment. In a natural environment, however, fresh DOC would enter the aquatic system, e.g., by rainfall and influx from the catchment *via* rivers, brooks, or surface run-off, eventually by vertical mixing, constantly refreshing the DOC pool. Further, recalcitrant DOC

would undergo photochemical processing that breaks down high molecular weight humic acids into more bioavailable substrates (Bertilsson and Tranvik, 1998). Under such conditions, mineralization would continue to increase with increasing DOC concentrations, and continuous supply of P would support higher rates of mineralization. This is in accordance with the findings from the lake survey where total CO<sub>2</sub> production rates increased with decreasing DOC:TP ratios (**Supplementary Figure S3B**) and increasing TP.

In the laboratory experiments, respiration rates increased with increasing DOC concentration up to 20 mg L<sup>-1</sup>. This increase in respiration with DOC was in accordance with the increase in total CO<sub>2</sub> production rates at increased DOC concentration >5 mg L<sup>-1</sup> found in the lake survey (**Figure 2**). Many boreal lakes have DOC concentrations below 20 mg L<sup>-1</sup> (in our lake survey, for instance, the maximum value was 12.9 mg C L<sup>-1</sup>) and a continued increase in BR with increased terrestrially derived DOC up to about 20 mg L<sup>-1</sup> could be expected.

The respired fraction of DOC in the laboratory experiment was small (max 18%) and decreased with increased DOC concentration beyond a distinct peak at 12.5 mg DOC L<sup>-1</sup> (**Supplementary Figure S6**). Since the source of DOC and thus the bioavailable share was the same in all samples, this suggests a lower mineralization efficiency with increased concentrations of DOC. In high DOC (>25 mg L<sup>-1</sup>) treatments, the CO<sub>2</sub> accumulation curves of neither P-spiked nor P-limited samples reached a plateau (**Supplementary Figure S4**), suggesting that mineralization would continue beyond the 200 h of incubation, although at lower pace.

In the experiment using O<sub>2</sub>-consumption as proxy for microbial activity (**Supplementary Figure S5**), temperature and P were strong predictors of O<sub>2</sub> while DOC was a poor predictor. A temperature increase from 15 to 25°C yielded an increase in respiration rates by a factor of 2.6 (±0.2) with no significant difference between P levels. This Q<sub>10</sub> value is in accordance with reported Q<sub>10</sub> values for physiological processes (Pomeroy and Wiebe, 2001). While respiration rates did not increase further at temperatures >25°C in P-limited samples, they decreased in P-spiked samples. Between 25 and 30°C the Q<sub>10</sub> thus differed significantly between P spiked (0.44 ± 0.09) and P limited (0.94 ± 0.05) samples. This difference in Q<sub>10</sub> may reflect a difference in bacterial taxonomic composition between treatments of different P levels with a faster growing and less robust community in P spiked than in P limited samples. Although there is a trend of increasing water temperatures (O'Reilly et al., 2015), an increase in temperature >25°C is unlikely to occur within the nearest future. Around 15–25°C may therefore be the most relevant temperature range for natural systems.

The observed temperature response together with the temperature sensitivity of secondary production being higher than that of primary production (Brown et al., 2004; Apple et al., 2006), further reinforce the idea that net heterotrophy in lakes will increase with increasing temperatures, which also would lead to increased emissions of CO<sub>2</sub> (Sobek et al., 2003). While we found no temperature effect on neither lake CO<sub>2</sub>

production, nor CO<sub>2</sub> flux in the lake survey, likely reflecting that temperature measurements were snapshots, we further speculate that the narrow temperature gradient sampled in our study is overridden by a dynamic natural environment with several potentially confounding and fluctuating factors. Moreover, the role of primary production as a regulator of CO<sub>2</sub> is minor in these lakes. Given the positive temperature effect on microbial metabolism (Farrell and Rose, 1967), the ongoing rise in temperature, along with current browning (O'Reilly et al., 2015; Solomon et al., 2015), means that an increased CO<sub>2</sub> output from boreal lakes and rivers can be expected. The availability of P will serve as an additional regulator of carbon emissions, with increased respiration rates and DOC turnover rates in DOC-rich lakes where the role of primary production is small, and vice versa in cases where P is declining (Thrane et al., 2014).

Bacterioplankton respiration is a key process for converting organic carbon to CO<sub>2</sub> in aquatic ecosystems (Williams and Del Giorgio, 2005). This mineralization of DOC driven by microbial respiration is accompanied by O<sub>2</sub> consumption, often with an assumed RQ of 1, yet this quotient depends on substrate properties and metabolic states (Dilly, 2003; Berggren et al., 2012; Alleson et al., 2016). Similar to the CO<sub>2</sub> production rates in experimental set-up 1 and the higher O<sub>2</sub> consumption rates in P-spiked samples in experimental set-up 2 could represent a situation with higher BGE than in P-limited samples. As anabolic processes often are accompanied with elevated RQ's, we can expect some differences in RQ between samples of different P levels and possibly some underestimation of respiration rates in P-spiked samples by the use of an RQ value of 1. However, such differences in RQ values between P levels would make the differences in respiration rates between treatments more pronounced and the conclusions thus are still valid. High rates of heterotrophic respiration together with low rates of primary production promote oxygen depletion with major consequences for aquatic life as well as redox processes and biogeochemical cycling of C, N, P, and other elements. While increased respiration rates following increased DOC concentrations may favor primary production due to increased access to CO<sub>2</sub> as well as nutrients associated with DOC, increased browning and thereby increased light attenuation would most probably result in a net decline in primary production and thus increased net heterotrophy (Thrane et al., 2014; Seekell et al., 2015), with a tentative turning point around 5 mg C L<sup>-1</sup>.

In conclusion, the DOC concentration regulates the overall respiratory output of CO<sub>2</sub> (and consumption of O<sub>2</sub>), while

additions of P changes the dynamics by boosting respiration, as did elevated temperatures. The overall respiratory outcome depends on substrate stoichiometry and the potentially different cell-specific responses versus community responses; i.e., larger biomass will generate a larger total CO<sub>2</sub> output despite lower cell-specific respiration. The dynamic responses revealed in the small-scale batch experiments do not necessarily capture inter-lake responses to changing DOC:TP ratios, partly because "fresh" DOC becomes available for microbial respiration due to inflow and mixing *in situ*, and partly because phytoplankton responses will impact the net CO<sub>2</sub> balance. Also the full nature of biotic uptake and recycling can clearly not be captured, but the combination of a gradient lake surveys together with the laboratory experiments revealed DOC as the major determinant of CO<sub>2</sub> production in boreal lakes, with P as a significant modulator.

## DATA AVAILABILITY STATEMENT

The raw data supporting the conclusions of this article will be made available by the authors, without undue reservation.

## AUTHOR CONTRIBUTIONS

LA and DH conceived the idea. LA, DH, PD, and AE conducted the experiments. All authors were involved in the analysis of data and final writing. All authors contributed to the article and approved the submitted version.

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## SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: <https://www.frontiersin.org/articles/10.3389/fmicb.2020.569879/full#supplementary-material>

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**Conflict of Interest:** The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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## Supporting information materials and methods

Vertical profiles of scalar irradiance in the photosynthetically active radiation (PAR) region (400-700 nm;  $E_0$ ) were measured using a spherical irradiance sensor (BioSpherical instruments) attached to a 10 channel CTD profiler (WRW620, RBR Ltd., Canada). The sensor was lowered at a rate of approximately  $20 \text{ cm s}^{-1}$  with a sampling rate of 6 Hz. In order to correct for temporal changes in irradiance caused by for example wave action and clouds during the CTD cast, we regressed the log-transformed  $E_0$  against depth ( $z$ ) for each ten sampling point (i.e. sliding windows). The vertical attenuation coefficient for scalar PAR ( $K_0\text{PAR}$ ) was then estimated by taking the median of the distribution of these slopes.

Measurements of concentrations of total phosphorus (TP), total organic carbon (TOC) and total nitrogen (TN) were carried out both at the Norwegian Institute for Water Research (NIVA) and at the University of Oslo (UiO). Regressions between the measurements at the two laboratories showed no systematic differences (TP:  $R^2 = 0.77$ , residual standard error ( $RSE$ ) =  $2.27 \mu\text{g l}^{-1}$ ; TOC:  $R^2 = 0.99$ ,  $RSE = 0.25 \text{ mg l}^{-1}$ ; TN:  $R^2 = 0.91$ ,  $RSE = 81 \mu\text{g l}^{-1}$ ) and the averages of the results were used in the subsequent analysis. DOC was calculated as the difference between the total organic carbon (TOC) and particulate organic carbon (POC). TOC was measured by infrared  $\text{CO}_2$  detection after catalytic high temperature combustion (Shimadzu TOC-VWP analyser (UiO), or Phoenix 8000 TOC-TC analyser (NIVA)).

POC was measured on an elemental analyser (Flash EA 1112 NC, Thermo Fisher Scientific, Waltham, Massachusetts, USA) through rapid combustion in pure oxygen of particulates captured on a pre-combusted GF/C-filter. The major part ( $> 95 \%$ ) of the TOC was in dissolved



form (DOC). At UiO, TOC was measured together with total inorganic carbon (TIC). TP was measured on an auto-analyser as phosphate after wet oxidation with peroxodisulfate. The two labs measured TN in different ways. UiO measured TN on unfiltered samples by detecting nitrogen monoxide by chemiluminescence using a TNM-1 unit attached to the Shimadzu TOC-VWP analyser, and NIVA measured TN through detection of nitrate after wet oxidation with peroxodisulfate in a segmented flow auto-analyser.

For gas analyses, water from the composite water sample (integrated from 0 to 5 m depth) was gently let into 120 ml glass serum vials without bubbling. The samples were fixed with 0.2% HgCl and sealed with gas-tight butyl rubber stoppers (see Yang et al. (2015) for details). Prior to analysis, the vials were stored dark and cold (4 °C). In order to prepare a 20-30 ml headspace, a gentle helium pressure (needle valve) was applied to the top of the bottle volume replacing ca. 40 ml of sample with pure He, before venting the bottles to 1 atmosphere. Equilibration between liquid and headspace was achieved by shaking the bottles horizontally at 150 rpm for 2 h at room temperature. Headspace concentrations of CO<sub>2</sub> and O<sub>2</sub> were determined by automated gas chromatography (GC; Model 7890A, Agilent, CA, USA).

In brief, the bio-optical model calculating area specific primary production (PP<sub>A</sub>) is based on estimating the *in vivo* rate of light absorption by phytoplankton, and subsequently electron transport rates (ETRs) through photosystem II (PSII) using information about the light-dependent quantum yield of the PSII photochemistry. ETR can further be converted to a rate of gross carbon fixation by assuming an appropriate value for the quantum yield of CO<sub>2</sub> fixation (Kromkamp and Forster, 2003, Suggett et al., 2010). While the method could be sensitive to phytoplankton community composition, it has gained increased interest over the last two decades because it offers a fast and inexpensive way of obtaining PP<sub>A</sub> estimates (see Thrane et al (2014)

for details). A comparison of this method and empirical estimates for  $PP_A$  in boreal lakes demonstrated good accordance (Thrane et al., 2014). The method is thus a feasible tool for assessment of primary production across a large number of sites. It also avoids many of the pitfalls of  $^{14}C$ -bottle incubation, which in any case could not have been applied in this kind of synoptic survey.

### CO<sub>2</sub> flux

We used Fick's law of diffusion to calculate the water-air flux of CO<sub>2</sub> ( $F_{net}$ ; mmol m<sup>-2</sup> d<sup>-1</sup>) from lake surface CO<sub>2</sub> concentrations.:

$$F_{net} = k_{CO_2} \Delta_{CO_2} \quad (1)$$

where  $k_{CO_2}$  (m d<sup>-1</sup>) is the CO<sub>2</sub> gas exchange coefficient at a given temperature and  $\Delta_{CO_2}$  (mmol m<sup>-3</sup>) is the CO<sub>2</sub> deficit from concentrations at equilibrium with the atmosphere, obtained using Henry's law.  $k_{CO_2}$  was estimated for each lake using the gas transfer velocity (m d<sup>-1</sup>) for a gas-temperature combination with a Schmidt number of 600 ( $k_{600}$ ; CO<sub>2</sub> at 20 °C) according to Jähne et al. (1987):

$$k_{CO_2} = k_{600} \left( \frac{Sc_{CO_2}}{600} \right)^{-x} \quad (2)$$

where  $x = 2/3$  if wind speed  $\leq 3\text{ms}^{-1}$  and  $x = 0.5$  if wind speed  $> 3\text{m s}^{-1}$ ,  $Sc$  is the temperature dependent Schmidt number for CO<sub>2</sub>.  $k_{600}$  is estimated from the wind speed according to Cole and Caraco (1998):

$$k_{600} = 2.07 + 0.215 U_{10}^{1.7} \quad (3)$$

Hourly wind speed data at 10 m above ground ( $U_{10}$  in equation 9) at all 75 lakes were extracted from the Norwegian Reanalysis Archive (NORA10) and aggregated into July-August means.

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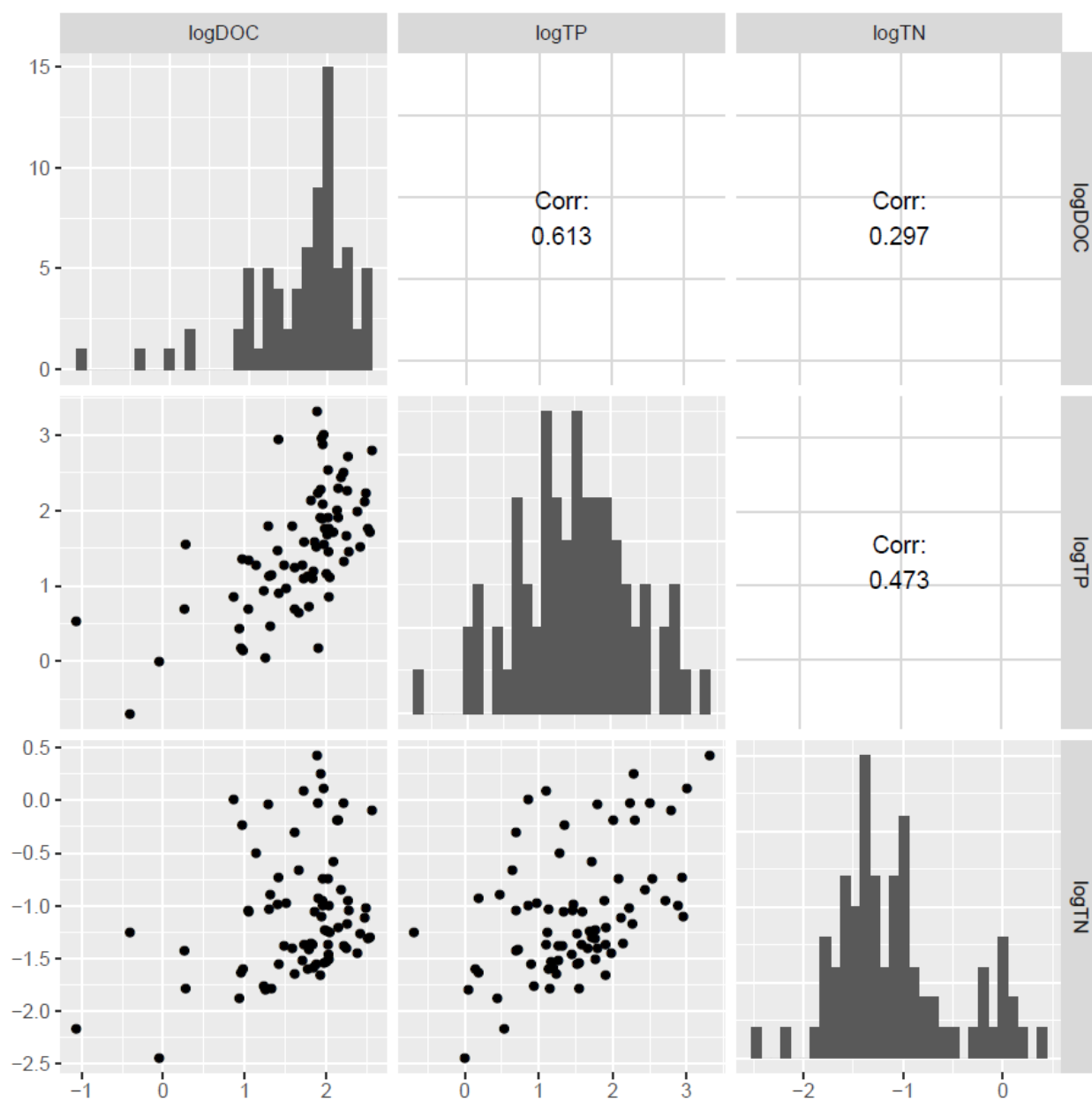


Figure S1. Correlation matrix between dissolved organic carbon (DOC;  $\text{mg L}^{-1}$ ), total phosphorus (TP;  $\mu\text{g L}^{-1}$ ), and total nitrogen (TN;  $\text{mg L}^{-1}$ ). All variables are log-transformed.

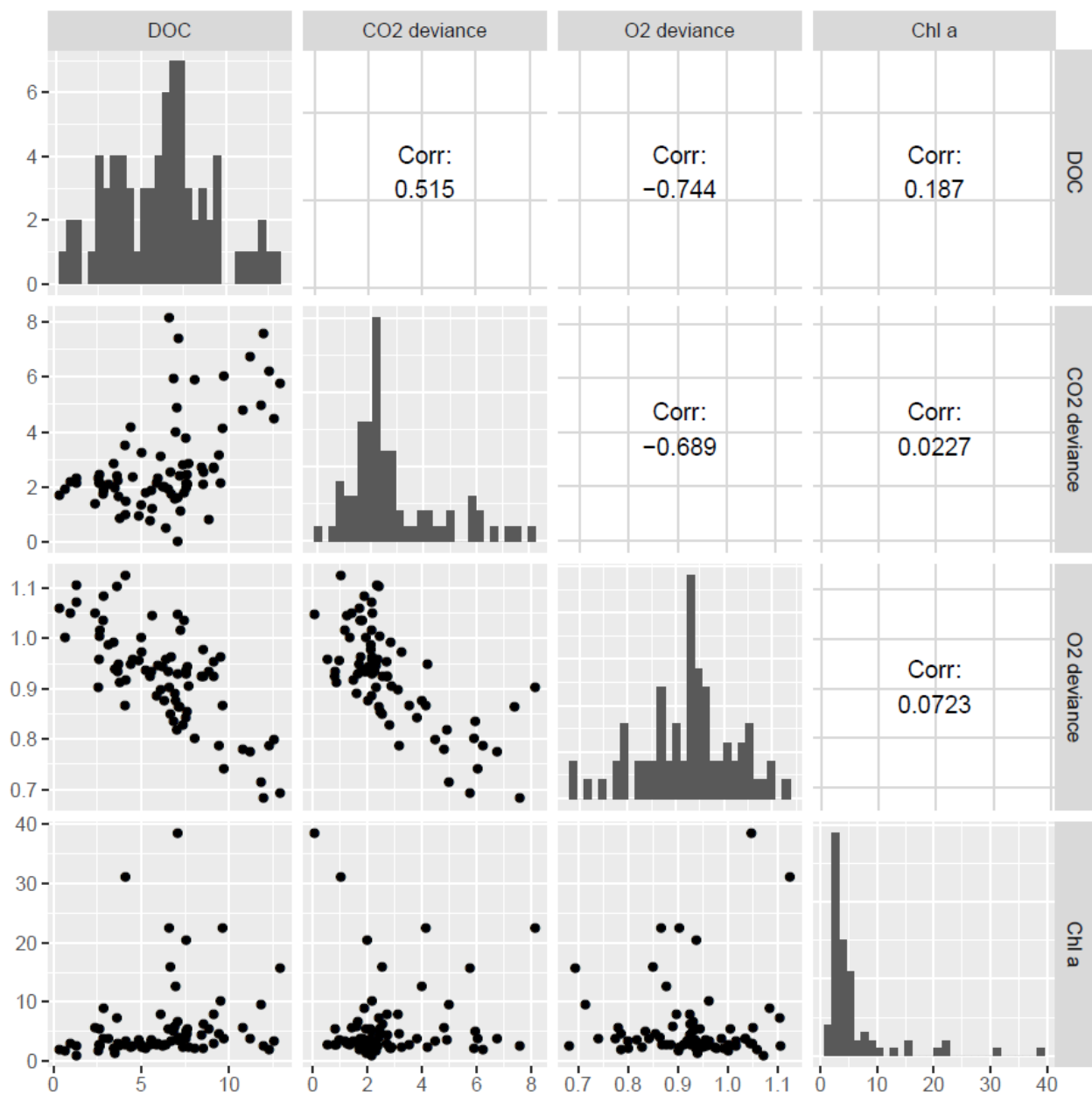


Figure S2. Correlation matrix between DOC ( $\text{mg L}^{-1}$ ), CO<sub>2</sub> deviance from saturation (%), O<sub>2</sub> deviance from saturation (%), and concentration of chlorophyll *a* (chl *a*;  $\mu\text{g L}^{-1}$ ).

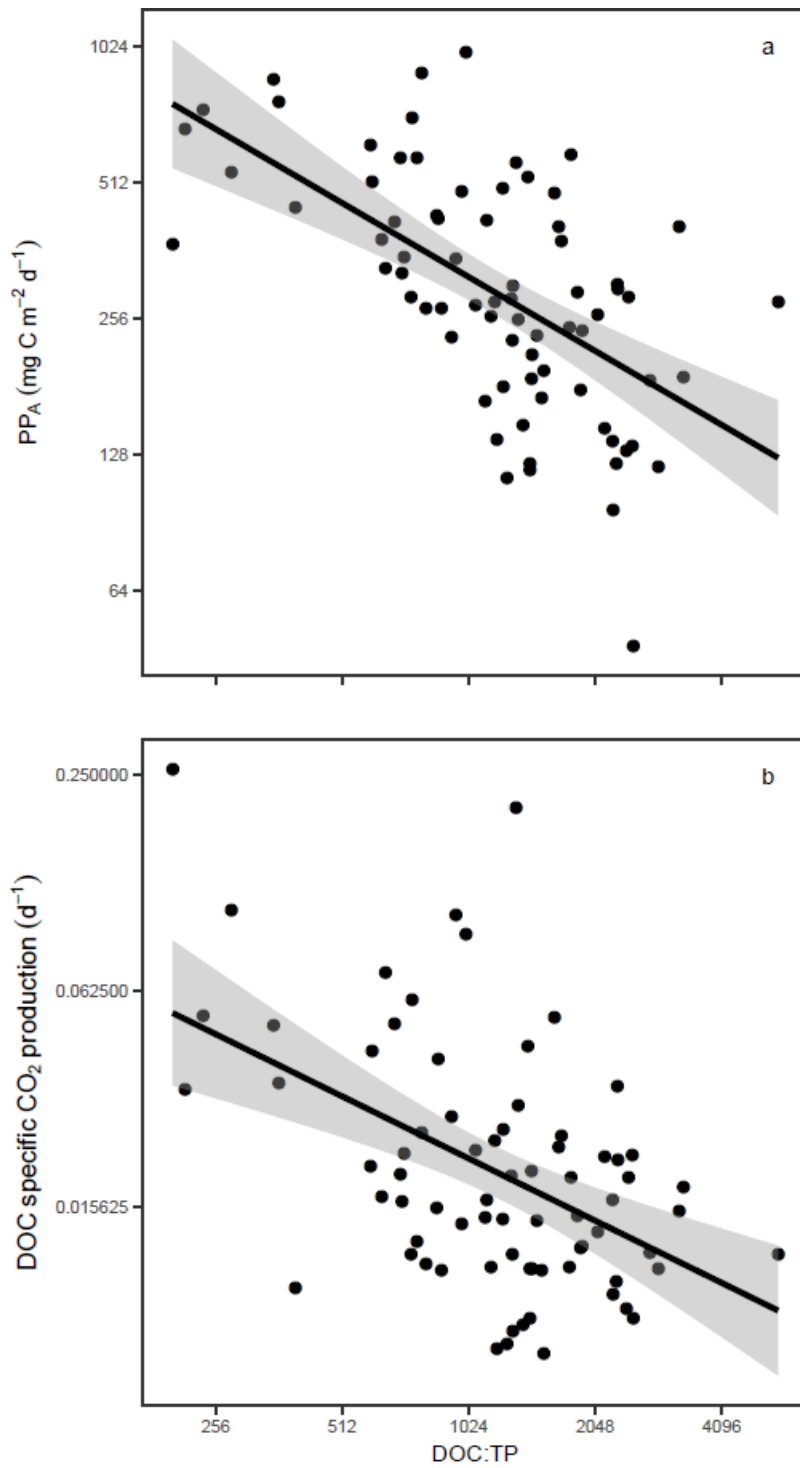


Figure S3. The response of a) area specific primary production ( $PP_A$ ;  $\text{mg C m}^{-2} \text{d}^{-1}$ ), and b) DOC specific  $\text{CO}_2$  production rates ( $F_{\text{tot}}$  ( $\text{mg C m}^{-2} \text{d}^{-1}$ )/DOC ( $\text{mg C m}^{-2}$ )) to increased DOC:TP ratio.

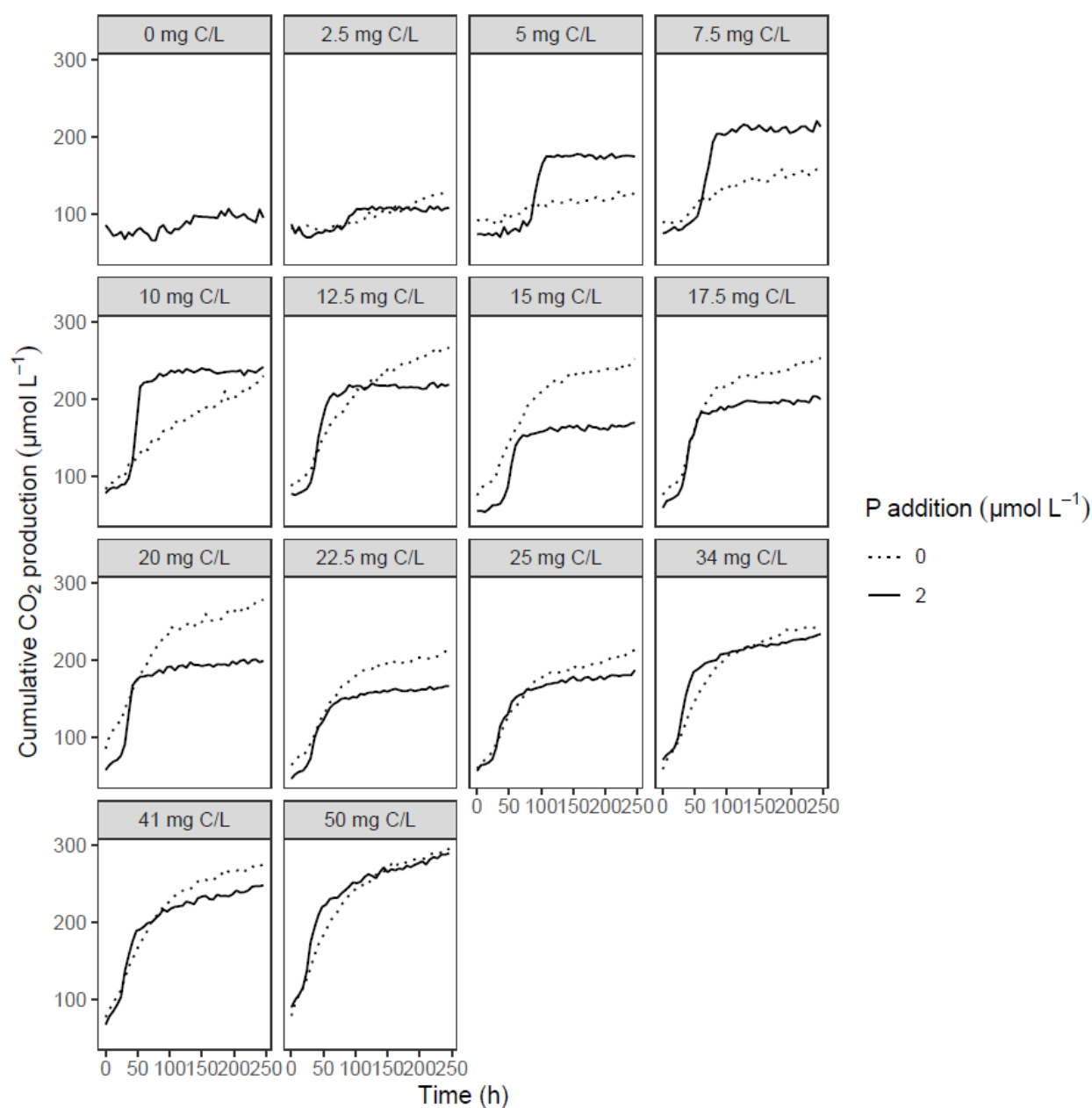


Figure S4. Cumulative CO<sub>2</sub> production over incubation time. The figure is divided into boxes of DOC additions starting from top left with no additions to bottom right with 50 mg C L<sup>-1</sup>. Dotted and solid lines represent treatments without and with P additions, respectively.

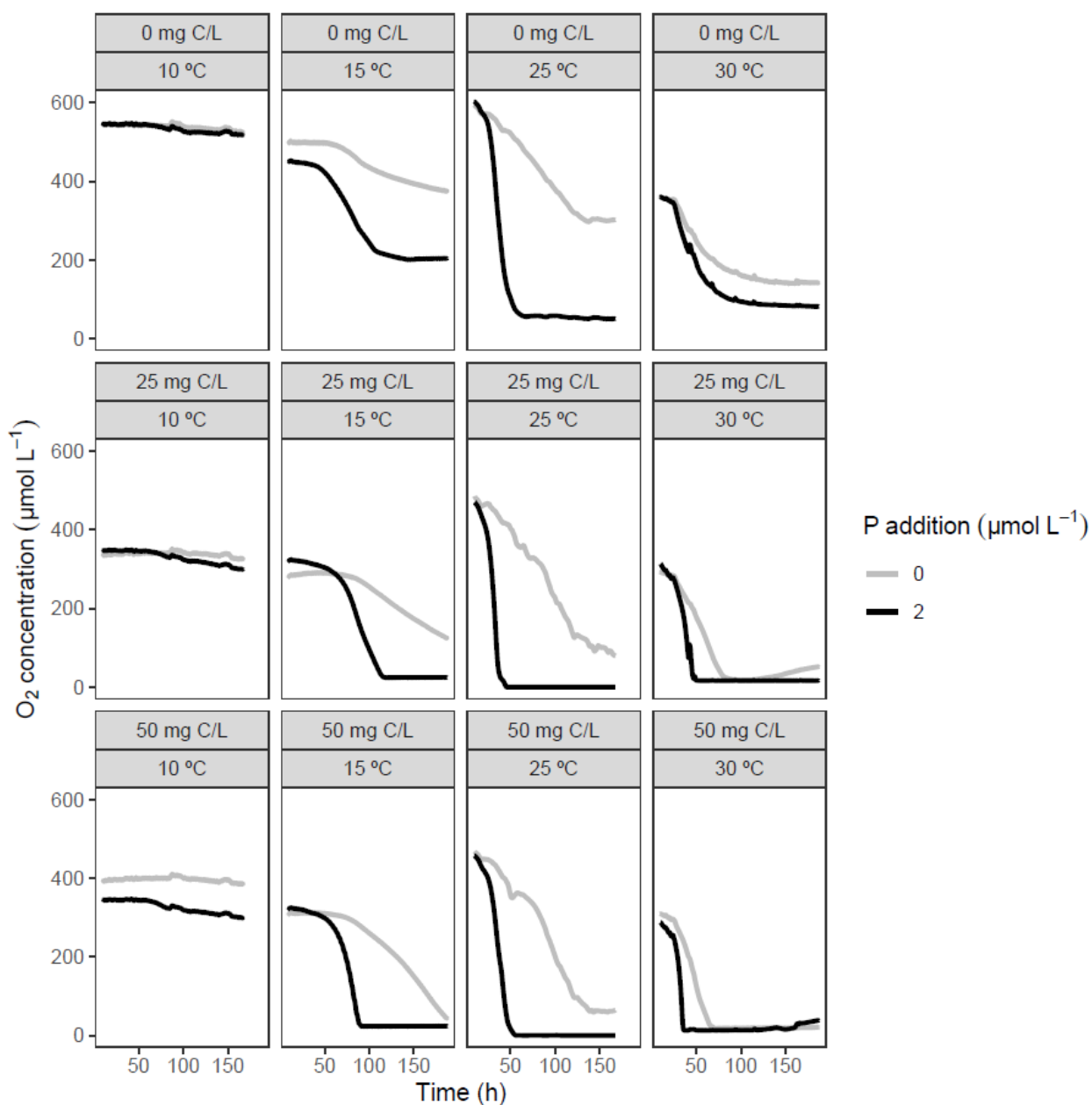


Figure S5. Cumulative O<sub>2</sub> consumption over incubation time. The figure is divided into boxes of DOC (top row: no addition; middle row: 25 mg C L<sup>-1</sup>; bottom 50 mg C L<sup>-1</sup>), and temperature (from 10 °C in the left column to 30 °C in the right column). The colors represent treatments without (red) and with (blue) P additions, respectively.

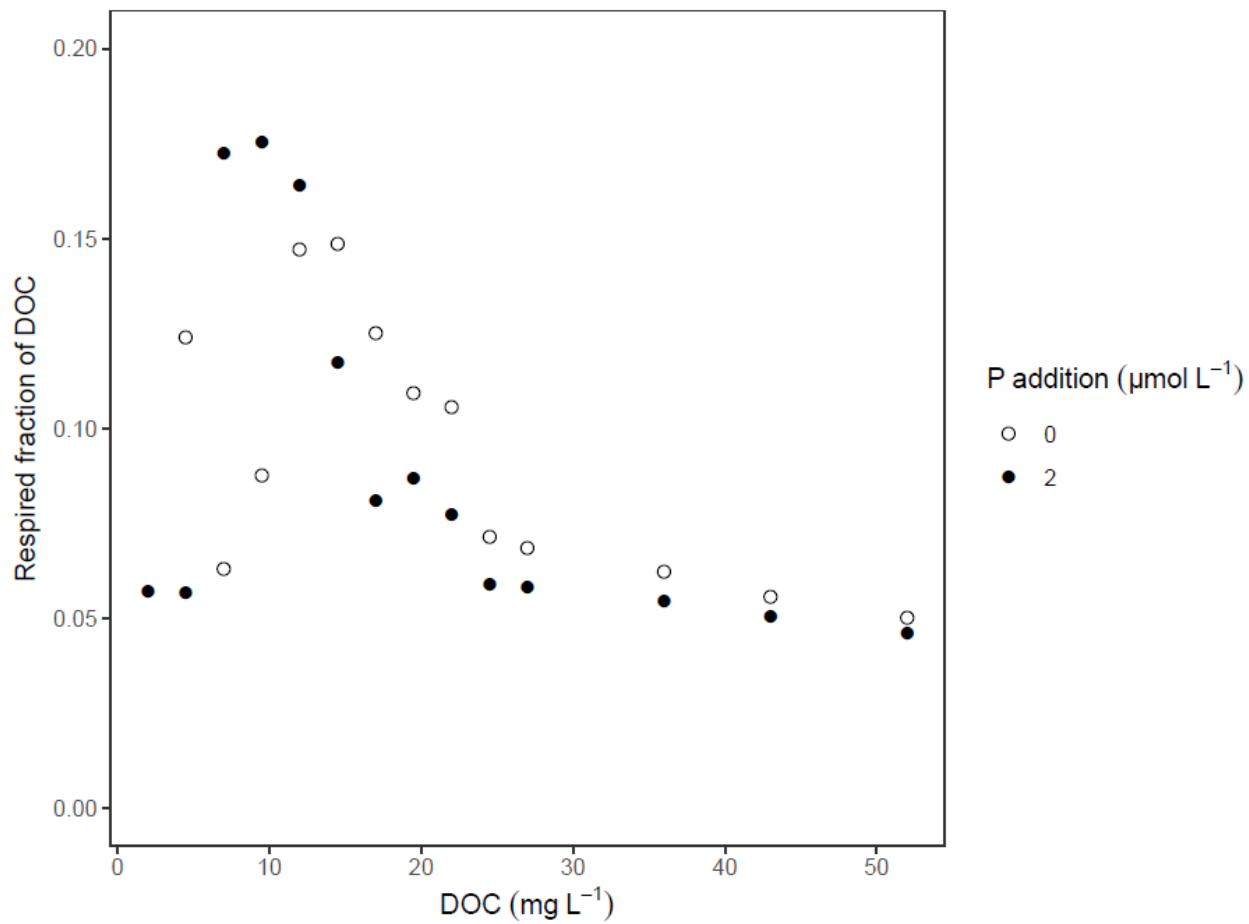


Figure S6. The respired fraction of the added DOC versus DOC concentration (mg L<sup>-1</sup>) open circles are treatments without P and filled circles are treatments with 2 μmol L<sup>-1</sup> P addition.

**Table S1.** Results of a generalized additive model (GAM) from the two experiments using non-parametric smoothers.

Response variable	Factor	edf*	F-value	R <sup>2</sup> <sub>adj</sub>	Dev. Expl.#	Effect <sup>§</sup>
<b>Experimental set-up 1</b>						
total CO <sub>2</sub> production	s(DOC):P0	7.372	23.98	0.935	97.5 %	P*
	s(DOC):P2	7.731	20.27			
16S rRNA gene copy number	s(DOC):P0	1.493	7.273	0.98	98.9 %	P***
	s(DOC):P2	8.931	56.04			
copy number specific respiration	s(DOC):P0	7.548	3.748	0.813	88.2 %	P***
	s(DOC):P2	1.000	0.703			
maximum CO <sub>2</sub> production rate	s(DOC):P0	2.825	26.46	0.91	93.6 %	P***
	s(DOC):P2	3.689	26.89			
<b>Experimental set-up 2</b>						
maximum O <sub>2</sub> consumption rate	s(temp):P0:DOC0	1.000	6.49	0.844	84.8%	P*** DOC*
	s(temp):P2:DOC0					
	s(temp):P0:DOC25	1.990	92.98			
	s(temp):P2:DOC25					
	s(temp):P0:DOC50	1.000	15.04			
	s(temp):P2:DOC50					
		1.983	104.18			
		1.608	11.13			
		1.982	89.71			

\* edf is the estimated degree of freedom accounting for the smoothing function.

# Deviance explained by the model with all factors.

§ Significance of P-level (experimental set-up 1) and P and DOC levels (experimental set-up 2) on the response variable. Significant codes: \*\*\* <0.001; \*\* <0.01; \* <0.05.

**Table S2.** Results from analysis of covariance (ANCOVA) using data from the two experiments.

<b>Variable tested</b>	<b>explanatory</b>	<b>Estimate</b>	<b>t-value</b>	<b>p-value</b>	<b>R<sup>2</sup><sub>adj</sub></b>	<b>p-value</b>
<b>Experimental set-up 1</b>						
total CO <sub>2</sub> production	DOC	2.946	5.650	<0.001	0.553	<0.001
	P	-17.90	-1.227	0.232		
16S rRNA gene copy number	DOC	1.93 × 10 <sup>8</sup>	2.305	0.0301	0.642	<0.001
	P	1.57 × 10 <sup>10</sup>	6.690	<0.001		
copy number specific respiration	DOC	-16.1	-0.811	0.425	0.626	<0.001
	P	0.374	-6.730	<0.001		
maximum CO <sub>2</sub> production rates	DOC	0.028	4.724	<0.001	0.531	<0.001
	P	0.539	3.257	<0.005		
<b>Experimental set-up 2</b>						
maximum O <sub>2</sub> consumption rate	DOC	0.554	1.784	0.077	0.563	<0.001
	P	0.365	11.415	<0.001		
	Temperature	3.903	7.270	<0.001		



# Paper III





## RESEARCH LETTER

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## Key Points:

- Photoprocessed DOC is consumed at higher bacterioplankton RQ than nonphotoprocessed DOC
- Bacterioplankton RQ depends on the chemical composition of the assimilated organic substrate pool
- Faulty assumption of RQ = 1 in light-exposed water greatly underestimates bacterioplankton respiration

## Supporting Information:

- Supporting Information S1

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## Impact of photochemical processing of DOC on the bacterioplankton respiratory quotient in aquatic ecosystems

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**Abstract** Many studies assume a respiratory quotient (RQ = molar ratio of CO<sub>2</sub> produced to O<sub>2</sub> consumed) close to 1 when calculating bacterioplankton respiration. However, evidence suggests that RQ depends on the chemical composition of the respired substrate pool that may be altered by photochemical production of oxygen-rich substrates, resulting in elevated RQs. Here we conducted a novel study of the impact of photochemical processing of dissolved organic carbon (DOC) on RQ. We monitored the bacterial RQ in bioassays of both ultraviolet light irradiated and nonirradiated humic lake water, using optic gas-pressure sensors. In the experimentally irradiated samples the average RQ value was significantly higher (3.4–3.5 [ $\pm 0.4$  standard error (SE)]) than that in the dark controls (1.3 [ $\pm 0.1$  SE]). Our results show that the RQ is systematically higher than 1 when the bacterial metabolism in large part is based on photoproducts. By assuming an RQ of 1, bacterioplankton respiration in freshwater ecosystems may be greatly underestimated.

### 1. Introduction

Most inland waters worldwide are supersaturated with carbon dioxide (CO<sub>2</sub>), driving a globally significant ( $\sim 1\text{--}2\text{ Pg C yr}^{-1}$ ) CO<sub>2</sub> flux to the atmosphere [Cole *et al.*, 1994; Raymond *et al.*, 2013]. Several processes contribute to this CO<sub>2</sub>, including groundwater input [Humborg *et al.*, 2010], sediment respiration [Gudasz *et al.*, 2010], and photochemical mineralization of dissolved organic carbon (DOC) [Koehler *et al.*, 2014], but often bacterioplankton respiration (BR; release of CO<sub>2</sub> per unit water volume and time) fueled by DOC is dominating the CO<sub>2</sub> production in the water column [Jansson *et al.*, 2000; Sobek *et al.*, 2003; Yang *et al.*, 2015]. It has been estimated that BR could be the single most important organic carbon mineralization process in the biosphere [Williams and del Giorgio, 2005]. However, there are methodological uncertainties in the measurement of BR, leading to a lack of understanding of the carbon cycle, especially in freshwaters [Humborg *et al.*, 2010]. A key uncertainty in the determination of BR is the assumption of a respiratory quotient (RQ), i.e., a certain fixed amount of respiratory CO<sub>2</sub> produced per unit measured O<sub>2</sub> consumption [Gaarder and Gran, 1927].

Throughout the history of BR assessments, indirect assessments based on dissolved oxygen (O<sub>2</sub>) consumption measurements [Winkler, 1888] have been favored over direct CO<sub>2</sub> production measurements [del Giorgio *et al.*, 2011; Marchand *et al.*, 2009]. The O<sub>2</sub> method is advantageous because it can be performed on the aqueous phase, without gas extraction, and it avoids the need to correct for changing equilibria within the carbonic acid system (CO<sub>2</sub>  $\leftrightarrow$  HCO<sub>3</sub><sup>-</sup>  $\leftrightarrow$  CO<sub>3</sub><sup>2-</sup>). Moreover, the predominant analytical techniques for O<sub>2</sub> are typically cost efficient, accurate, and easy to use (Winkler, electrode, and optode methods) or in some cases extremely highly resolved, e.g., using membrane inlet mass spectroscopy [Kana *et al.*, 1994]. Therefore, still today the RQ remains a required conversion factor to transform respiration rates measured as O<sub>2</sub> consumption into carbon units.

Although generically assumed at a fixed value close to 1, true bacterioplankton RQ can vary greatly [Berggren *et al.*, 2012; Cimblaris and Kalff, 1998; Romero-Kutzner *et al.*, 2015]. For example, the RQ is affected by biochemical pathways of metabolism, where anabolic processes (cell growth) contribute to higher RQs than catabolic respiration alone [Berggren *et al.*, 2012; Dilly, 2003]. More importantly, RQ is strongly dependent on the composition of the assimilated organic substrate [Berggren *et al.*, 2012; Romero-Kutzner *et al.*, 2015], especially the O and H content [Dilly, 2001]. Assimilation of organic substrates with large O content and high O:H ratio requires less O<sub>2</sub> from the surroundings, and hence, respiration is performed at a relatively high RQ.

Photochemical processes in aquatic ecosystems may represent the most important source of oxygen-rich low molecular weight (LMW) compounds available for consumption by bacterioplankton [Bertilsson and

**Table 1.** Stoichiometry of Complete Aerobic Oxidation of Selected Compounds

Compound	Chemical Equation	CO <sub>2</sub> Produced	O <sub>2</sub> Consumed	RQ
Methane	$\text{CH}_4 + 2 \text{O}_2 \rightarrow \text{CO}_2 + 2 \text{H}_2\text{O}$	1	2	0.5
Glucose	$\text{C}_6\text{H}_{12}\text{O}_6 + 6 \text{O}_2 \rightarrow 6 \text{CO}_2 + 6 \text{H}_2\text{O}$	6	6	1
Succinic acid <sup>a</sup>	$\text{C}_4\text{H}_6\text{O}_4 + 3.5 \text{O}_2 \rightarrow 4 \text{CO}_2 + 3 \text{H}_2\text{O}$	4	3.5	1.14
Citric acid <sup>a</sup>	$\text{C}_6\text{H}_8\text{O}_7 + 4.5 \text{O}_2 \rightarrow 6 \text{CO}_2 + 4 \text{H}_2\text{O}$	6	4.5	1.33
Glycolic acid <sup>a</sup>	$\text{C}_2\text{H}_4\text{O}_3 + 1.5 \text{O}_2 \rightarrow 2 \text{CO}_2 + 2 \text{H}_2\text{O}$	2	1.5	1.33
Malic acid <sup>a</sup>	$\text{C}_4\text{H}_6\text{O}_5 + 3 \text{O}_2 \rightarrow 4 \text{CO}_2 + 3 \text{H}_2\text{O}$	4	3	1.33
Tartaric acid <sup>a</sup>	$\text{C}_4\text{H}_6\text{O}_6 + 2.5 \text{O}_2 \rightarrow 4 \text{CO}_2 + 3 \text{H}_2\text{O}$	4	2.5	1.6
Formic acid <sup>a</sup>	$\text{CH}_2\text{O}_2 + 0.5 \text{O}_2 \rightarrow \text{CO}_2 + \text{H}_2\text{O}$	1	0.5	2
Oxalic acid <sup>a</sup>	$\text{C}_2\text{H}_2\text{O}_4 + 0.5 \text{O}_2 \rightarrow 2 \text{CO}_2 + \text{H}_2\text{O}$	2	0.5	4

<sup>a</sup>LMW organic acids detected in this study.

Tranvik, 1998]. Several of the most common LMW compounds that are released from reactions between DOC and ultraviolet (UV) sunlight are theoretically oxidized at RQs between 1.3 and 4 (see examples in Table 1). Although photooxidation is known to be an important process in natural freshwater systems, e.g., directly contributing to up to about 10% of the CO<sub>2</sub> efflux from lakes and reservoirs to the atmosphere [Koehler *et al.*, 2014], its potential role as RQ regulator has to our knowledge never been addressed. If partial photooxidized DOC compounds represent a major source of carbon for inland water bacteria, then the natural RQs are likely higher than 1 and the CO<sub>2</sub> production is systematically underestimated even on a global scale [Williams and del Giorgio, 2005].

The commonly applied RQ of 1 corresponds to complete oxidation of glucose (Table 1), but it is also close to the theoretical RQ for complete oxidation of bulk natural DOC [Dilly, 2001]. However, bacterioplankton do not normally make use of the bulk pool of DOC, especially not humic fractions, but instead use small selected fractions that are easy to assimilate [Berggren *et al.*, 2007]. These compounds contain various proportions of O and H, resulting in a potential range in RQ from 0.7 to 0.8 for phytoplankton-derived DOC [Berggren *et al.*, 2012] to far above 1 for many photoproducted organic acids (Table 1). Thus, the theoretical RQ for complete oxidation of bulk DOC pools may tell us little about the actual RQs, resulting in errors when estimating BR.

In this study, we therefore test the impact of UV radiation on bacterioplankton RQ hypothesizing that photochemically processed DOC is used at a higher RQ than nonirradiated DOC because bacteria assimilate highly oxidized photoproducts such as organic acids. To test this hypothesis, we conducted biological incubations of both irradiated and nonirradiated samples of natural lake water, simultaneously measuring the changes in O<sub>2</sub> and CO<sub>2</sub> to obtain the RQ. To confirm that the photochemical processes contributed to increased concentrations of LMW organic acids (LMWOAs), we also measured concentrations of common LMWOAs in all samples, before and after UV treatment. We expected that the bacterioplankton RQ would be significantly higher than 1 after UV treatment and around 1 in nonirradiated samples (obtained from natural humic lake waters).

## 2. Methods

### 2.1. Study Site and Sampling

Samples were collected from four brown-water, humic lakes in northern Sweden during 14 and 21 August 2014 (for lake data see supporting information Table S1). The catchments are dominated by coniferous forest, mainly Norway spruce (*Picea abies*, L.; 80%) and *Sphagnum* peatmires (20%). The lakes can be considered representative for unproductive humic lakes in the boreal zone, characterized by low productivity and high degrees of CO<sub>2</sub> supersaturation [Jansson *et al.*, 2000]. We can assume that DOC in the samples was dominated by allochthonous sources [Karlsson *et al.*, 2012]. In the laboratory, the water was sterile filtered through a 0.22 μm pore size filter (Sartorius Stedim Biotech, NY) and kept cold 4°C until further analyses.

### 2.2. Study Design

Before the experiments, the samples were transferred into 300 mL acid washed Erlenmeyer flasks and sparged with synthetic air (N<sub>2</sub> and O<sub>2</sub>) to remove CO<sub>2</sub> supersaturation. The samples were then inoculated with 1 mL of a fresh unfiltered mix of water from the study lakes and their inlets, in equal proportions. The

filled flasks were closed using ground glass joints, leaving no headspace, and additionally sealed with parafilm to avoid diffusion of CO<sub>2</sub>. Biological incubations were carried out in the dark with both irradiated water and ambient lake water controls in a climate chamber (PU-3 J high performance chamber, ESPEC, Japan) keeping a stable temperature of 20°C (±0.1°C). In each experiment, one sample per lake was incubated.

The experiment consisted of three parts. (1) *Irradiated*: the water was first irradiated during 48 h followed by 48 h of biological incubation. (2) *Biological + irradiated*: the samples were first incubated in the dark for 48 h followed by 48 h of UV treatment and then finally biologically incubated for 48 h. (3) *Dark control*: the samples were biologically incubated in the dark for 144 h. The longer incubation time was chosen to mirror the time of the second treatment, controlling that a possible change in RQ was due to bacterial consumption of photochemical processed DOC and not incubation time.

### 2.3. Biological Incubations and Bacterial Respiration

Depending on the alkalinity and pH in the system, a certain share of the respiratory CO<sub>2</sub> dissociates in the water, forming bicarbonate and carbonate ions [Stumm and Morgan, 1996]. To cover the total CO<sub>2</sub> (TCO<sub>2</sub> = CO<sub>2</sub> + HCO<sub>3</sub><sup>-</sup> + CO<sub>3</sub><sup>2-</sup>) production during BR measurements, the impact of the carbonate system on CO<sub>2</sub> hence needs to be corrected for. Either the CO<sub>2</sub> concentrations can be measured and used to calculate the pH-induced changes in ionization fractions, or the change in pH during a bioassay can be measured and used directly to calculate the TCO<sub>2</sub> production (see supporting information Text S1 for details). In this study, we measured BR through changes in pH (decreasing as CO<sub>2</sub> is produced) in the water samples using noninvasive optical pH sensor spots placed in the bottom of each flask with a SensorDish Reader (SDR: resolution: 0.05 units; PreSens GmbH, Regensburg, Germany). Each lake was considered a replicate. In order to obtain the same starting point pH was standardized to the range 6.5–7, by adding 0.2 M NaOH. Bacterial O<sub>2</sub> consumption was measured with an “oxy-10 system” (resolution: best: ± 0.14 μM at 2.83 μM and worst: ± 1.14 μM at 283 μM; PreSens GmbH, Regensburg, Germany), recording the concentration of dissolved oxygen every 5 min with optic sensor spots glued on to the inside of the flasks [Marchand et al., 2009].

Bacterial communities have a short establishment time and previous studies have shown that a total of 48 h of incubation time is sufficient to acquire enough data to analyze the bacterial respiration [Berggren et al., 2012; Cimleris and Kalff, 1998]. To let the sensors equilibrate and reach a stable reading and allow the bacteria to establish, we waited 10–12 h before starting any gas measurements. The impact of photoproduced hydrogen peroxide and other reactive oxygen species could be neglected in our measurements due to their relatively short half-life under nonsterile, aerobic conditions [Tranvik and Kokalj, 1998], assuming they disappeared during the sensor equilibration time. Since the measurement range of the optical pH sensors was 5.5–8.5, some data points in irradiated samples with high rates of CO<sub>2</sub> production were lost, as the incubation pH in these samples sometimes reached very low values (pH < 5.5).

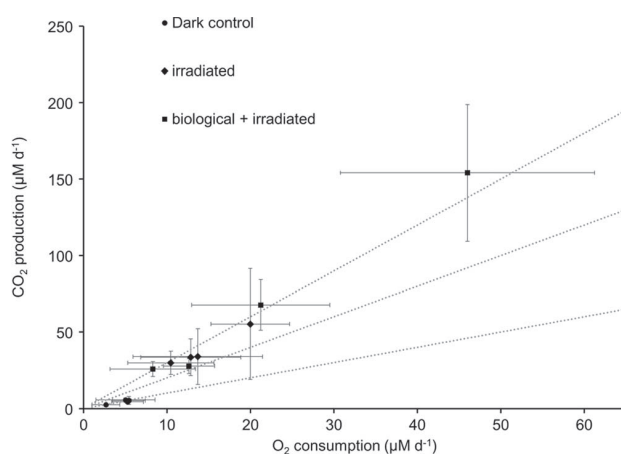
### 2.4. Irradiation Incubations

Before irradiation the water samples were transferred to 1000 mL quartz glass bottles (Chemglass Life Sciences, Vineland, NJ). The samples were thereafter placed vertically on a spinning disk (0.67 rpm) under UV lamps and incubated for 48 h in a climate chamber kept at 20 ± 1°C. The radiation was within 3.64–6.89 Wm<sup>-2</sup> for UV-A and 0.06–0.1 Wm<sup>-2</sup> for UV-B, which is roughly representative to average daytime radiation during summer in northern Sweden. Since the disk was spinning, all samples received the same amount of light. Photochemical processing produces CO<sub>2</sub>, alters the gas equilibration and may damage the microorganisms [Anesio and Granéli, 2003]. Therefore, inoculation was performed again following each UV treatment.

### 2.5. Chemical Analyses

Determination of LMWOAs was made using a liquid ion chromatography-ionspray tandem mass spectrometry system (IC-MS). The system consisted of a Dionex (Sunnyvale, CA, USA) ICS-2500 liquid chromatography system and an Applied Biosystems (Foster City, CA, USA) 2000 Q-trap triple quadrupole mass spectrometer. The method is described in further detail in Ström et al. [2012].

For IC-MS analysis of LMWOA, subsamples of ~ 10 mL were sampled before start of each irradiation treatment (data not shown) and twice during each biological treatment, at the beginning and end, respectively. To replenish the water in the Ehrlenmeyer flasks after subsampling, water treated identically and in parallel to the water samples being monitored were incubated at the side and used for this purpose. The subsamples



**Figure 1.** Plot of the daily increase in  $\text{CO}_2$  concentration versus the daily decrease in  $\text{O}_2$  concentration (in absolute numbers). Error bars represent the standard error and grey dotted reference lines from below represent RQs of 1, 2, and 3, respectively.

the known alkalinity in the water samples, using the equations from *Stumm and Morgan* [1996]. We thus accounted for all  $\text{CO}_2$  production in the samples, i.e., both the share that was transformed into carbonates and the share that stayed as dissolved  $\text{CO}_2$ . The concentrations of the different species were calculated using well-established temperature-dependent empirical equilibration constants (the uncertainty in these constants was considered negligible). The pH measurements were averaged over 5 h and  $\text{TCO}_2$  concentrations were calculated for each 5 h pH value with the standard deviation (SD) representing the spread in the measurements (Figure 1). The RQ was then calculated as the  $\text{TCO}_2$  production divided by the  $\text{O}_2$  consumption during the five hour intervals in each sample and incubation. The standard error (SE) of the RQ was calculated as the SD divided by the square root of the number of observations.

The calculated RQs were compared through a repeated measures analysis of variance (ANOVA) followed by a Tukey HSD post hoc test to compare the RQs of the different treatments. In order to test whether the RQs obtained were significantly different from 1, a one sample *t* test (two tailed) was performed.

The concentrations of the different LMWOAs that would give rise to a theoretical  $\text{RQ} > 1$  were summed and converted to concentrations of carbon per liter ( $\mu\text{M C}$ ) in the samples for each incubation period. A correlation analysis between the concentrations and the RQ values was then performed. All statistical tests were performed using R [R Development Core Team, 2015].

### 3. Results

The pH decreased significantly over time in all bioassays, allowing us to calculate the release of  $\text{CO}_2$  from BR. In bioassays with irradiated water, the  $\text{CO}_2$  production was remarkably high. With a range between lakes of  $27\text{--}52\ \mu\text{M d}^{-1}$  in the first trial (irradiated) and of  $25\text{--}130\ \mu\text{M d}^{-1}$  in the second trial (biological and irradiated), the  $\text{CO}_2$  production was one order of magnitude higher than the nonirradiated dark controls (range  $2.6\text{--}6.6\ \mu\text{M d}^{-1}$ ). Therefore, although intended to be 48 h, some of these bioassays had to be aborted after 20–30 h (depending on the alkalinity of the lake water; Table S1 and Figure S1 in the supporting information) as the lower limit of the pH sensors was approached.

Similar to the pattern for  $\text{CO}_2$ , the  $\text{O}_2$  consumption rates were consistently higher in irradiated samples than in nonirradiated dark controls. The highest rates were measured in the second trial (biological + irradiated) ranging between  $8.7$  and  $46.0\ \mu\text{M d}^{-1}$  (Figure 1). Following the initial equilibration time, the  $\text{O}_2$  consumption rates were stable and linear. Similarly, the  $\text{CO}_2$  production rates were mostly stable throughout the experiment, although showing some increase toward the end of the incubation time in bioassays with especially high rates (Figure S1 in the supporting information).

were filtered through  $0.22\ \mu\text{m}$  pore size filters, prerinsed with 30 mL of distilled water to remove any LMWOA contaminants, and deep frozen until further analysis.

Analyses of DOC were performed on  $0.45\ \mu\text{m}$  filtered water samples (30 mm Polyethersulfone (PES) membrane filters, Thermo scientific, MA, USA) using a TOC-VCN (Shimadzu, Japan). Alkalinity ( $\mu\text{eq L}^{-1}$ ) was defined and measured as the amount HCl ( $\mu\text{mol}$ ) that was consumed per liter sample water during titration with 0.1 M HCl to an endpoint of pH 4.5.

### 2.6. Calculations and Statistics

The  $\text{TCO}_2$  production was calculated from the change in pH together with

**Table 2.** RQ Values With Standard Deviations and Organic Acids Carbon Concentration During the Different Treatments and for All Four Lakes<sup>a</sup>

Lake and Treatment	RQ ( $\pm$ SE)	Organic Acids <sup>b</sup> ( $\mu$ M C)
<i>Övre Björntjärn</i>		
Dark control	1.2 ( $\pm$ 0.1)	5.6
Irradiated	4.1 ( $\pm$ 1.0)	8.0
Biological + irradiated	3.9 ( $\pm$ 0.9)	8.0
<i>Nedre Björntjärn</i>		
Dark control	1.0 ( $\pm$ 0.1)	6.1
Irradiated	3.5 ( $\pm$ 0.4)	11.5
Biological + irradiated	2.3 ( $\pm$ 0.2)	7.6
<i>Lillsjöleden</i>		
Dark control	1.6 ( $\pm$ 0.2)	5.7
Irradiated	3.5 ( $\pm$ 0.5)	10.3
Biological + irradiated	3.7 ( $\pm$ 0.7)	10.5
<i>Stortjärn</i>		
Dark control	1.3 ( $\pm$ 0.2)	6.1
Irradiated	3.1 ( $\pm$ 0.6)	7.8
Biological + irradiated	4.1 ( $\pm$ 0.5)	6.4
<i>All Lakes</i>		
Dark control	1.3 ( $\pm$ 0.1)	
Irradiated	3.5 <sup>a</sup> ( $\pm$ 0.4)	
Biological + irradiated	3.4 <sup>a</sup> ( $\pm$ 0.4)	

<sup>a</sup>The analysis was performed using repeated measures ANOVA ( $F_{2,6} = 32.32$ ,  $p < 0.001$ ) with a Tukey HSD post hoc test. RQ in irradiated samples were significantly higher than in dark controls (dark control—irradiated  $p < 0.0001$ ,  $df = 28$ ; dark control—biological + irradiated  $p < 0.0001$ ,  $df = 26$ ).

<sup>b</sup>Organic acid concentrations represent the mean of two measurements performed during different time points of the bioassays. Concentrations were not systematically different between these time points.

plankton RQ and the concentration of LMWOAs ( $r^2 = 0.718$ ;  $p < 0.01$ ;  $n = 12$ ; Figure S2 in the supporting information). However, the apparent production of LMWOA during UV treatment (difference in concentrations measured before and after) was relatively low compared to the large amount of CO<sub>2</sub> production that was recorded during biological incubation of irradiated samples. In the nonirradiated dark controls, the concentration of LMWOAs was 5.6–6.1  $\mu$ M C, while in the irradiated samples the ranges were 7.7–11.2  $\mu$ M C in the first trial (irradiated) and 6.5–10.5  $\mu$ M C in the second trial (biological + irradiated) (Table 2 and Figure S2 in the supporting information). These organic acid concentrations corresponded to 19–48% of the cumulative CO<sub>2</sub> production in nonirradiated samples and 6–24% of the corresponding production in irradiated water.

#### 4. Discussion

Our results clearly show that the bacterioplankton respiratory quotient (RQ) is elevated during degradation of photochemically processed DOC compared to dark controls. The RQ in the same lake water samples shifted from slightly above 1 before irradiation to exceeding 3 after UV light treatment of the water. We further found that the irradiation greatly boosted the CO<sub>2</sub> production rates during biological incubations (approximately tenfold increase, on average, compared to nonirradiated dark control incubations), which implies that photoaltered DOC was strongly dominating as substrate after irradiation. In agreement with our hypothesis, we observed relatively high concentrations of LMWOAs in UV light treated water, in parallel to the elevated RQs, and there was an overall positive correlation between RQ and the total LMWOA concentration. These results imply that the estimates of the contribution to CO<sub>2</sub> by bacterioplankton are greatly underestimated assuming an RQ of 1 in naturally light-exposed aquatic ecosystems.

From a mass balance perspective, however, the elevated RQ values found in irradiated samples could only partially be explained by bacterial use of LMWOAs, because the measured photoproduction of LMWOAs was too small to account for the photostimulated CO<sub>2</sub> release. Moreover, we systematically observed photostimulated RQs above 3, which is higher than the expected theoretical RQs—mostly below 2—for oxidation of the LMWOAs that were produced (Table 1). Although other studies have observed higher photoproduction

Dark controls showed an average RQ of 1.3, significantly different from 1 (one sample  $t$  test,  $t = 2.96$ ,  $p = 0.004$ ). Furthermore, the RQ in irradiated samples was significantly ( $p < 0.0001$ ) higher than the dark controls, with mean values of 3.5 ( $\pm$ 0.4 SE) and 3.4 ( $\pm$ 0.4 SE) for water irradiated directly (irradiated) and water irradiated after biological incubation (biological + irradiated) respectively (Table 2). Thus, also in the second trial (biological + irradiated) we could see a shift in the RQ from before to after UV treatment.

A number of LMWOAs that theoretically are oxidized at an RQ above 1 were found in the IC-MS analysis (Table 1). The concentrations of these acids were slightly higher in irradiated than in nonirradiated samples (Table 2) indicating that there was a production of LMWOAs due to photochemical processing of DOC. There was a significant positive correlation between bacterio-



rates of LMWOAs than we did [Bertilsson and Tranvik, 2000], it is evident from our results that analyses of organic acids alone provide limited possibilities for explaining the large amounts of CO<sub>2</sub> released during biological processing of irradiated water (Figure S2 in the supporting information). Thus, the change in RQ that we observed was not only dependent on LMWOA production but most likely also by production and modification of a broader range of DOC molecules. This conclusion is supported by the previous study by Miller and Moran [1997] which observed biological degradation of photoaltered DOC exceeding the detected LMWOA production by 90%. Therefore, a recommendation for future RQ studies is to combine LMWOA measurements with chemical characterizations of the bulk DOC pool and to assess changes in the oxidation state of the entire substrate pool by performing a redox balance.

In highly humic (dark) lakes, natural UV light only penetrates a few centimeters into the water column and the majority of photochemical reactions hence take place at the surface layers. Nonetheless, photochemically produced carboxylic acids have been shown to be a major source of labile DOC over the entire mixed zone [Bertilsson and Tranvik, 1998]. The dark control RQ exceeded 1 ( $1.3 \pm 0.1$ ), which is in agreement with the average RQ of a number of humic rich lakes in Québec [Berggren et al., 2012] and may reflect the downward mixing of photochemically altered DOC in the lakes.

It should be noted that all substrates that are assimilated are not oxidized by the microorganisms into CO<sub>2</sub> but instead used for production of biomass [Biddanda et al., 1994; del Giorgio and Cole, 1998]. In soil ecosystems, increases in microbial growth under glucose amended conditions have resulted in elevated RQs up to 1.5 [Dilly, 2003]. In lakes, bacterial consumption of photoprocessed DOC has been shown to result in enhanced microbial biomass production [Amado et al., 2015; Anesio et al., 2005], which could give a positive marginal effect on RQ similar to what has been observed in soils. However, in this study bacterial production was not measured and it can only be speculated that enhanced biomass production might have partly contributed to the high RQ values observed in UV treated compared to dark samples. Nonetheless, considering the vastly elevated RQs that we observed, a shift in substrate pool due to partial photooxidation is more likely to have had an overriding impact on the bacterial RQ than physiological processes that tend to have only marginal effects [Romero-Kutzner et al., 2015].

The consistently much higher RQ values that we observed in the irradiated samples reflect a mechanism that should be present in all light-exposed DOC-rich aquatic ecosystems around the globe. The relative importance of photomineralization to partial photooxidation of DOC is dependent on the age of the compounds and the intensity of the incoming solar radiation [Cory et al., 2013, 2014; Vähätalo et al., 2003]. Photochemical mineralization of DOC to CO<sub>2</sub> represents up to about 10% of the total CO<sub>2</sub> emissions from lakes and reservoirs globally [Koehler et al., 2014] and studies of photochemical processing of DOC have reported partial photooxidation possibly be as important as photomineralization [Bertilsson and Tranvik, 2000]. In water samples from a coastal marine environment, Miller and Moran [1997] found DOC losses from photochemical gas formation to be approximately equal to the losses from microbial consumption of labile photoproducts. Applying an underestimated RQ value would thus lead to large amounts of CO<sub>2</sub> from BR being missed, given that labile photoproducts require little dissolved oxygen to degrade. There is extensive evidence of microbial consumption of photoproduced LMW compounds [Bertilsson and Tranvik, 2000; Judd et al., 2007]. In long term bioassays (years), simulating water residence times in lakes, Vähätalo and Wetzel [2008] showed that exposure to solar radiation followed by bacterial degradation depleted all humic DOC (99.7%). It must thus be expected that bacterioplankton in natural waters will consume photoproducts and most likely, according to our results, at an RQ significantly exceeding 1.

## 5. Summary, Conclusions, and Recommendations

Biological incubations of water with photochemically processed DOC lead to enhanced RQ values compared to nonirradiated dark controls. The RQ was consistently higher than 1 in irradiated samples and the CO<sub>2</sub> production rates exceeded the rates in nonirradiated dark controls several fold. In fact, we could see a clear shift in RQ from slightly above 1 before to exceeding 3 after UV treatment. The elevated RQ values may partly be explained by bacterial consumption of single photoproduced LMWOAs but, more likely, also by a general shift in oxidation state of the bulk substrate pool. Our results indicate that by the use of a fixed RQ value of 1, bacterioplankton respiration in inland waters may be globally underestimated. Therefore, we cannot recommend the a priori assumption of an RQ value for a lake to estimate the BR from dissolved oxygen



consumption. For future respiration studies we suggest to either determine the RQ or to measure the CO<sub>2</sub> production directly instead of measuring the O<sub>2</sub> consumption. The possible loss of information due to less accurate and more cumbersome methods for measuring CO<sub>2</sub> compared to measuring O<sub>2</sub> is most probably not as severe as the uncertainty accompanying the assumption of a faulty RQ value.

#### Acknowledgments

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*[Geophysical Research Letters]*

Supporting Information for

**[Impact of photo-chemical processing of DOC on the bacterial respiratory quotient  
in aquatic ecosystems]**

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**Contents of this file**

1. Methods S1
2. Table S1
3. Figure S1
4. Figure S2

**Description**

In Methods S1 we describe how the pH and alkalinity of the lake water samples was used to assess the carbonic system and to calculate total CO<sub>2</sub> release from bacterial respiration. Table S1 contains information of the lake characteristics, such as DOC concentrations, alkalinity, lake area and depth. In Figure S1, the cumulative CO<sub>2</sub> production is plotted against the cumulative O<sub>2</sub> consumption for each lake and bioassay. Finally, Figure S2 shows the relationship between RQ and the total concentrations of organic acids during the bioassays.

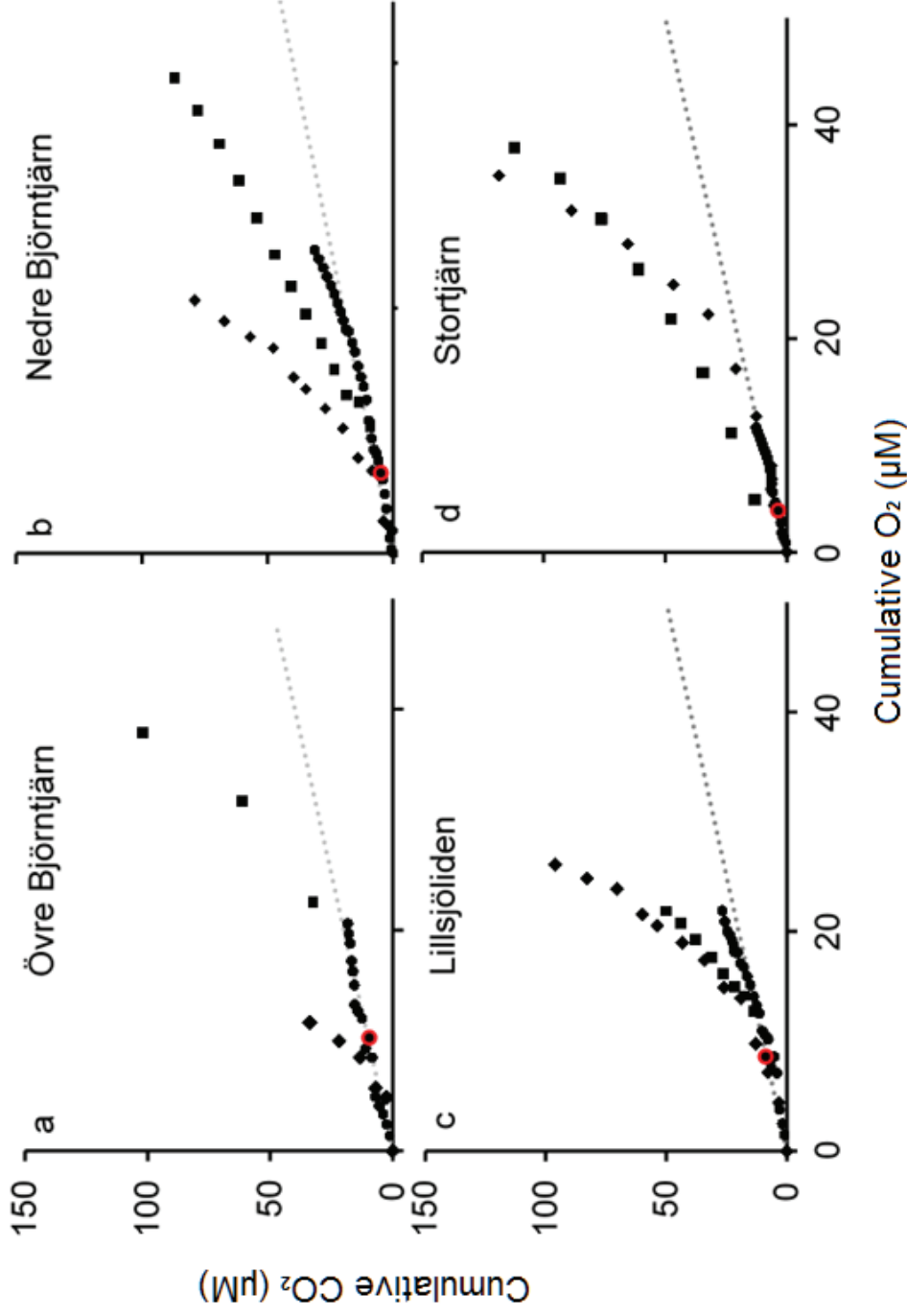
## Supporting Methods S1 – TCO<sub>2</sub> calculations from pH measurements

The carbonate system was modeled from the known pH, alkalinity and temperature of the samples, using the equations in Stumm and Morgan [1996]. The release of CO<sub>2</sub> by bacterial respiration (BR) causes formation of carbonate acid (H<sub>2</sub>CO<sub>3</sub>), in turn dissociating into HCO<sub>3</sub><sup>-</sup>, CO<sub>3</sub><sup>2-</sup> and H<sup>+</sup>, lowering the pH of the system. Therefore, the BR was calculated as the change in TCO<sub>2</sub> (CO<sub>2</sub> + HCO<sub>3</sub><sup>-</sup> + CO<sub>3</sub><sup>2-</sup>) concentrations over time. In this study we used closed systems with no headspace, implying that any change in pCO<sub>2</sub> was caused by mineralization of DOC to CO<sub>2</sub>. We considered that the release of CO<sub>2</sub> to the water did not affect the alkalinity since dissociation of CO<sub>2</sub> produces the same amount of negative and positive charges [Stumm and Morgan, 1996]. We further assumed that dissolved organic carbon (DOC) did not contribute significantly to the alkalinity. Thus, alkalinity was assumed to be constant during the bioassays, equaling the total charge of bicarbonate and carbonate ions. The concentrations and the ionization fractions of the different dissolved inorganic carbon species in the solution were obtained using existing empirical, temperature dependent constants of high accuracy [Stumm and Morgan, 1996].

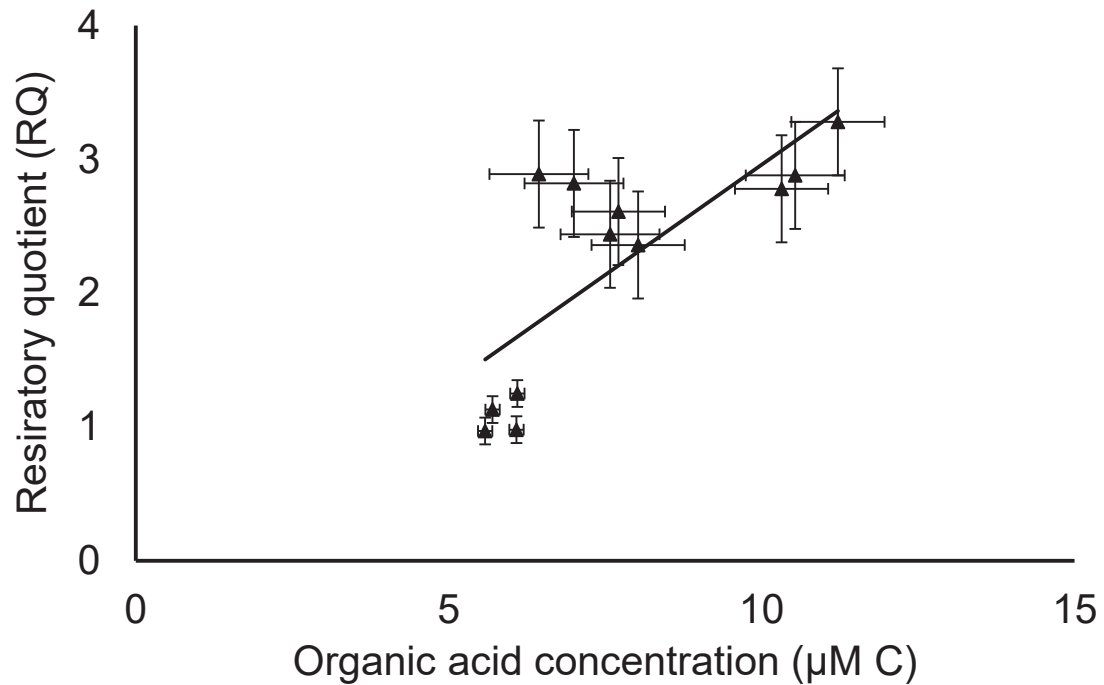
Although the pH sensors had been recently calibrated by the manufacturer, we noted a slight systematic difference between measured output values and the true pH values for a range of buffer solutions, possibly caused by the optical properties of our incubation flasks. Therefore we used the measurements from the buffer solutions (pH 5.5, 6.5, 7.5, and 8.5) to perform a four point linear recalibration ( $y = 0.9608 * x + 0.5826$ ;  $R^2 = 0.9942$ ).

**Table S1.** Study site information. Variables are self-explanatory or described elsewhere in the paper.

Lake name	Location	Elevation (m.a.s.l.)	Lake area (ha)	Lake depth, mean/max (m)	Lake peri- meter (m)	Catchment area (ha)	DOC (mM C)	Alkalinity ( $\mu\text{eq L}^{-1}$ )	CDOM <sub>440nm</sub> ( $\text{m}^{-1}$ )
Övre Björntjärn	64°12'N 18°78'E	336	4.8	4.0/8.0	1178	284.0	1.8	37	19.1
Nedre Björntjärn	64°12'N 18°78'E	336	3.2	6.0/9.7	893	324.9	2.3	75	17.0
Lillsjöleden	63°85'N 16°62'E	317	0.8	3.8/5.2	407	25.4	1.6	300	11.7
Stortjärn	64°26'N 19°76'E	284	3.9	2.7/6.7	1089	81.7	2.1	80	21.7



**Figure S1.** The cumulative CO<sub>2</sub> production plotted against the cumulative O<sub>2</sub> consumption in the bioassays. Circles represent 'dark controls', diamonds the 'irradiated' treatment and squares the 'biological + irradiated' samples (see main paper for explanations). The first 48 h of dark incubation in the 'biological + irradiated' treatment are overlapping with the dark controls before to after UV-treatment. The grey dotted line is the 1:1 reference line representing an RQ of 1.



**Figure S2.** Correlation ( $r^2 = 0.718$ ;  $p < 0.01$ ;  $n = 12$ ) between respiratory quotient (RQ) and the total concentration of carbon in measured organic acids. Symbols show means across the four study lakes and error bars display  $\pm 1$  standard errors of the means (see Table 2 in main paper).





# Paper IV



1 **Drivers and variability of CO<sub>2</sub>:O<sub>2</sub> saturation along a gradient from boreal to Arctic**  
2 **lakes**

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12

13 **Abstract**

14 Lakes are significant players for the global climate since they sequester terrestrially derived  
15 dissolved organic carbon (DOC), and emit greenhouse gases like CO<sub>2</sub> to the atmosphere.  
16 However, the differences in environmental drivers of CO<sub>2</sub> concentrations are not well  
17 constrained along latitudinal and thus climate gradients. Our aim here is to provide a better  
18 understanding of net heterotrophy and gas balance at the catchment scale. We assessed water  
19 chemistry and concentrations of dissolved O<sub>2</sub> and CO<sub>2</sub>, as well as the CO<sub>2</sub>:O<sub>2</sub> ratio in three  
20 groups of lakes separated by steps of approximately 10 degrees latitude in South-Eastern  
21 Norway (near 60 °N), sub-Arctic lakes in the northernmost part of the Norwegian mainland  
22 (near 70 °N) and high-Arctic lakes on Svalbard (near 80 °N). Across all regions, CO<sub>2</sub>  
23 saturation levels varied more (6 – 1374 %) than O<sub>2</sub> saturation levels (85 – 148 %) and hence  
24 CO<sub>2</sub> saturation governed the CO<sub>2</sub>:O<sub>2</sub> ratio. The boreal lakes were generally undersaturated  
25 with O<sub>2</sub>, while the sub-Arctic and high-Arctic lakes ranged from O<sub>2</sub> saturated to  
26 oversaturated. Regardless of location, the majority of the lakes were CO<sub>2</sub> supersaturated. In  
27 the boreal lakes the CO<sub>2</sub>:O<sub>2</sub> ratio was mainly related to DOC concentration, in contrast to the

28 sub-Arctic and high-Arctic localities, where conductivity was the major statistical  
29 determinant. While the southern part is dominated by granitic and metamorphic bedrock, the  
30 sub-Arctic sites are scattered across a range of granitic to sedimentary bed rocks, and the  
31 majority of the high-Arctic lakes are situated on limestone, resulting in contrasting lake  
32 alkalinities between the regions. DOC dependency of the CO<sub>2</sub>:O<sub>2</sub> ratio in the boreal region  
33 together with low alkalinity suggests that in-lake heterotrophic respiration was a major source  
34 of lake CO<sub>2</sub>. Contrastingly, the conductivity dependency indicates that CO<sub>2</sub> saturation in the  
35 sub-Arctic and high-Arctic lakes was to a large part explained by DIC input from catchment  
36 respiration and carbonate weathering.

37

38

39 **Keywords:** Aquatic Biogeochemistry, CO<sub>2</sub> saturation, O<sub>2</sub> saturation, Lake Heterotrophy,  
40 Geology, Dissolved Organic Carbon

41

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47

48

## 49 **Introduction**

50 Oxygen (O<sub>2</sub>) and carbon dioxide (CO<sub>2</sub>) are key gases for life on Earth. Their ratio is mainly  
51 determined by the balance between heterotrophic and autotrophic processes in the biosphere.  
52 Under net autotrophic conditions, CO<sub>2</sub> is sequestered in both terrestrial and aquatic  
53 ecosystems, thus decreasing atmospheric CO<sub>2</sub> concentrations and greenhouse effects on  
54 global climate. Lakes, and notably boreal lakes, are key players in this context since they  
55 convert a significant part of terrestrially derived organic carbon to CO<sub>2</sub> and CH<sub>4</sub> [1, 2]. A  
56 large share of CH<sub>4</sub> may also be converted to CO<sub>2</sub> by methanotrophs in the water column [3].  
57 Simultaneous and complementary biological processes thus drive variations in O<sub>2</sub> and CO<sub>2</sub>  
58 concentrations [4, 5] and this coupling has led to the assumption that O<sub>2</sub> and CO<sub>2</sub> can be used  
59 interchangeably when studying metabolism of aquatic ecosystems. O<sub>2</sub> is advantageous over  
60 CO<sub>2</sub> because it can be measured directly in the aqueous phase without gas extraction.  
61 Therefore, indirect assessments of CO<sub>2</sub> based on dissolved O<sub>2</sub> are often favored over direct  
62 CO<sub>2</sub> measurements [6, 7]. However, combined CO<sub>2</sub> and O<sub>2</sub> measurements have shown  
63 decoupling over time [8-10]. This may in part be explained by decoupling of production and  
64 consumption of these gases across the aquatic-terrestrial interface and thus reflect the  
65 catchment type of the aquatic ecosystem [9]. Other explanations for the deviations in CO<sub>2</sub>:O<sub>2</sub>  
66 coupling could be inputs of CO<sub>2</sub>-rich water and anaerobic CO<sub>2</sub> cycling [11]. Simultaneous  
67 measurements of CO<sub>2</sub> and O<sub>2</sub> concentrations thus increase the understanding of lake  
68 ecosystem functioning. Ratios of CO<sub>2</sub>:O<sub>2</sub> may in turn give a better indication of lake net  
69 heterotrophy than CO<sub>2</sub> or O<sub>2</sub> saturation alone, since it encompasses the net balance between  
70 autotrophic and heterotrophic and well as physical and chemical processes. It will also  
71 provide insight in spatial drivers of CO<sub>2</sub> emissions that could allow a space-for-time approach  
72 to address future changes in e.g. climate, hydrology and forest cover.

73

74 Biological oxidation of dissolved organic matter (DOM) is a main source of aquatic CO<sub>2</sub> [12-  
75 14] while photochemical oxidation plays a smaller role [14, 15]. Other sources may be CO<sub>2</sub>  
76 from catchment processes like root exudation or supersaturated groundwater (Raymond et al.,  
77 2013). Depending on the pH and buffering capacity of the water, inorganic carbon is either  
78 present as CO<sub>2</sub> or reacts with water forming bicarbonate or carbonate. Lake chemistry and  
79 notably pH thus plays a key role in determining the CO<sub>2</sub> concentration in the water. Similarly,  
80 elevated temperatures will reduce concentrations while not necessarily the level of saturation  
81 or the CO<sub>2</sub>:O<sub>2</sub> ratio, as the dissolution of both gases depends similarly on temperature.  
82 The Boreal zone is characterized by extensive land-water interfaces. Forests with large  
83 above- and below-ground C-pools as well as abundant bogs and wetlands, export terrestrial  
84 DOM and dissolved inorganic carbon (DIC) to waters, making boreal aquatic ecosystems an  
85 especially important component of the global carbon cycle [2]. In certain areas also  
86 agricultural areas may be important sources of organic C. Climate change, changes in land  
87 use, most notably afforestation, and the recovery from acidification [16] have led to increased  
88 export of terrestrial DOM and a shift in water color towards brown in many boreal  
89 freshwaters [17, 18]. This “browning” may affect lake metabolism in various ways.  
90 Terrestrially derived DOM provides energy and nutrients to heterotrophs, stimulating  
91 bacterial metabolism and CO<sub>2</sub> production [19, 20]. Nutrients associated to DOM may also  
92 benefit autotrophs. However, the beneficial effect of enhanced nutrient loadings is overridden  
93 by enhanced light attenuation inhibiting primary production as DOM concentration increases  
94 [20, 21]. This trade-off between positive and negative effects of increasing DOM inputs on  
95 photosynthesis may yield a unimodal response in primary production with a maximum around  
96 5-10 mg DOC l<sup>-1</sup> [22, 23].  
97 In contrast to boreal lakes, Arctic lakes have generally unforested catchments with less  
98 developed soils, yet there is a gradient from sub-Arctic to high-Arctic sites. Still they are

99 generally supersaturated with and net emitters of CO<sub>2</sub> to the atmosphere [24]. Lake  
100 abundancy is higher in the Arctic than in any other region of the world with high complexity  
101 in lake type ranging from clear, pristine, mountain lakes to brown, DOM rich thermokarst  
102 lakes formed by thawing permafrost [25-27]. Climate warming in the Arctic has led to  
103 increased biomass and production of terrestrial plants and shrubs, so called Arctic greening,  
104 leading to increased loads of DOM to Arctic lakes [28-30]. At the same time, thawing  
105 permafrost results in inputs of old organic carbon to Arctic lakes that may undergo microbial  
106 oxidation [31-33]. Increased DOM loadings in Arctic lakes are thus expected to result in  
107 enhanced CO<sub>2</sub> production and emission [30, 34, 35].

108 Autochthonous (in-lake) production is however not the only source of DIC. Lateral flux of  
109 inorganic carbon produced in the catchment may account for a sizeable share of lake CO<sub>2</sub>,  
110 especially in small lakes with short retention times and long ice-free seasons [36]. In-lake  
111 DOM mineralization together with DIC inputs make most lakes worldwide supersaturated  
112 with- and net emitters of CO<sub>2</sub> to the atmosphere [1, 37].

113 Whether a lake is a net conduit of CO<sub>2</sub> is however not necessarily a sign of net heterotrophy  
114 but could also reflect that catchment derived CO<sub>2</sub> exceeds photosynthetic uptake. Likewise,  
115 the magnitude of atmospheric CO<sub>2</sub> uptake in a net autotrophic system may be reduced by  
116 inputs of terrestrial CO<sub>2</sub>. There are conflicting reports of whether CO<sub>2</sub> produced in aquatic  
117 environments via DOM mineralization or exported from terrestrial environments is the main  
118 regulator of lake CO<sub>2</sub> flux [36, 38-40]. Some of the contradictory findings likely depend on  
119 climate, local hydrology, catchment slopes, water retention time, and not the least catchment  
120 properties like lake size or fraction and type of forest, bogs and wetlands [41, 42].

121 In this paper, we aim to gain understanding of lake CO<sub>2</sub> source, whether it is dominated by in-  
122 lake processes or by allochthonous inputs and whether there is a latitudinal difference in  
123 drivers of lake CO<sub>2</sub>:O<sub>2</sub> ratio, notably related to forested or unforested catchments. To do so,

124 we couple surface CO<sub>2</sub> and O<sub>2</sub> concentrations in 103 Norwegian lakes to environmental  
125 variables along a geographical gradient ranging from the boreal zone in southern Norway (58°  
126 N) through sub-Arctic northern Norway (69° N) to the high Arctic at Svalbard (79° N). The  
127 gradient reflects different catchment properties varying from dense spruce forest, via open  
128 birch forest to totally unforested catchments with thin soils in the high Arctic. We believe that  
129 this wide spatial gradient across climatic regions and catchment properties could provide  
130 insights relevant to larger parts of both the boreal and the arctic biome, yet with the proviso  
131 that there clearly may be pronounced regional differences within this vast area.

132

### 133 **Materials and methods**

#### 134 *Study lakes*

135 During fall 2019, 73 lakes in South-Eastern Norway (Boreal) and 22 Arctic lakes on Svalbard  
136 (high-Arctic) were sampled and a number of water chemistry parameters and dissolved gases  
137 were measured. In fall 2020, additionally 14 sub-Arctic lakes in the Finnmark county  
138 (Northern Norway; sub-Arctic) were sampled (Fig. 1). Sampling was performed at, or shortly  
139 after, fall overturn, to secure a maximum vertical homogenization of the water masses and  
140 gases. Since onset of fall differs with latitude, the samplings were performed during October,  
141 September and August for the three clusters of lakes (Boreal, sub-Arctic and high-Arctic,  
142 respectively), reflecting their geographical position from south to north. The boreal, southern  
143 cluster of lakes is within the coniferous forest zone but covers locations differing in size and  
144 altitude. The sub-Arctic lakes have sparsely forested catchments with birch, less topographic  
145 variation and larger areas of bogs and wetlands, while the high Arctic sites are treeless and  
146 also with very sparsely developed soils. Granitic and metamorphic bedrock dominate in the  
147 southern zone, yielding low alkalinity lakes (Fig. S1). The catchments of these lakes are also  
148 characterized by well-developed soils. The sub-Arctic lakes are situated on granitic to



149 sedimentary rocks (slate) giving rise to a wide range in alkalinity. The sub-Arctic lakes were  
150 chosen to span a geographical gradient from the southern inland to the costal northeast,  
151 reflecting a gradient in biome domination from taiga to Arctic tundra. A high proportion of  
152 the high-Arctic lakes are situated on limestone bedrock, with elevated alkalinity (Fig. S1).  
153 The high-Arctic lakes are mostly small with catchments devoid of soil and vegetation beyond  
154 scrubs and mosses, covering a gradient from rocky terrain from glacier fronts to shore sites,  
155 the latter influenced by birds and with some vegetation [43].

156

### 157 *Sampling and field sample preparations*

158 The lakes were sampled in early fall, after mixing but before ice-on. Sampling was performed  
159 from the shore using a sampling rod with a beaker, collecting surface water in a bucket. At the  
160 sampling sites, *in situ* measurements of temperature (T), pH and electrical conductivity (EC)  
161 were taken using a portable multimeter (Hach HQ40D). Samples were processed *in situ* for  
162 further analysis of lake chemistry: 50 ml of unfiltered water was taken for the analysis of total  
163 organic carbon (TOC), total nitrogen (TN) and total phosphorus (TP). For analysis of DOC,  
164 dissolved nitrogen (DN) and phosphorus (DP), surface water from the bucket was filtered in  
165 the field through Sterivex 0.22  $\mu\text{m}$  pore size filters (Millipore). All samples were frozen until  
166 further analysis.

167 Dissolved  $\text{CO}_2$ ,  $\text{O}_2$  and argon (Ar; all sites except Svalbard) were analyzed using the  
168 headspace method after acidification [44]. The samples were prepared in the field by filling  
169 30 ml water in a syringe, creating an in-situ headspace with air (20 ml) and adding 0.6 ml of  
170 3% HCl ( $\approx 1\text{M}$ ). Syringes were closed and equilibrated by shaking the syringes for 3 min. *In*  
171 *situ* lake temperature was used as equilibration temperature and caution was exercised to not  
172 warm the syringes during shaking. The equilibrated headspace gas was then injected into He-

173 washed and pre-evacuated glass vials crimp sealed with butyl rubber septa, and then kept at  
174 room temperature until analysis by gas chromatography.

175

#### 176 *Laboratory analysis*

177 TOC and DOC were measured by infrared CO<sub>2</sub> detection after catalytic high temperature  
178 combustion (Shimadzu TOC-VWP analyser). TP was measured on an Autoanalyser (Thermo  
179 Finnigan EA 1112 series flash elemental analyser) as phosphate after wet oxidation with  
180 peroxodisulfate. TN was measured on unfiltered samples by detecting nitrogen monoxide by  
181 chemiluminescence using a TNM-1 unit attached to the Shimadzu TOC-VWP analyser.

182 Concentrations of CO<sub>2</sub>, O<sub>2</sub> and Ar were determined by automated gas chromatography (GC)  
183 analysis with He back-flushing as described by Yang et al. [13]. The level of saturation of  
184 each gas relative to equilibrium with ambient air was calculated from measured  
185 concentrations using Henry's law with temperature-dependent solubility constants. In the sites  
186 where Ar was measured, O<sub>2</sub> was normalized to Ar saturation for a more stable and accurate  
187 estimate of O<sub>2</sub> saturation [45].

188

#### 189 *Statistical analysis*

190 All data analysis was performed using the open-source software R version 3.4.1 [46]. We use  
191 Spearman's rank correlation as a measure of association between continuous variables. To  
192 calculate DIC from pH and CO<sub>2</sub> at the *in situ* temperature, we used the AquaEnv package  
193 [47]. For the statistical modelling, we used the mgcv package [48] to fit generalized additive  
194 models (gam) of the gaussian family to the dependent variables. To test the dependency of  
195 CO<sub>2</sub>:O<sub>2</sub> ratio and O<sub>2</sub> saturation, we used a gam model with smoothers on each of the  
196 explanatory variables DOC (mg L<sup>-1</sup>), TP (µg L<sup>-1</sup>), TN (mg L<sup>-1</sup>), and conductivity (EC; µS cm<sup>-1</sup>).  
197 Predictive variable selection was done by applying additional shrinkage on the null space

198 of the penalty with the `select=TRUE` argument in the `mgcv::gam` function, as recommended  
199 by Marra and Wood [49]. The resulting models have all smoothers that are not necessary for  
200 the fit as close to zero as possible.

201

## 202 **Results**

203 Saturation levels of O<sub>2</sub> varied with geographical location (Fig. 2). By contrast, CO<sub>2</sub> saturation  
204 covered a much wider range with both undersaturated and supersaturated lakes in the sub-  
205 Arctic and saturation or supersaturation in the boreal and the high-Arctic lakes (Table 1; Fig.  
206 2).

207 The majority of the boreal lakes were supersaturated with CO<sub>2</sub> and undersaturated with O<sub>2</sub>,  
208 with a weak but significantly negative correlation between CO<sub>2</sub> and O<sub>2</sub> saturation levels ( $\rho = -$   
209  $0.42$ ,  $p < 0.001$ ). The sub-Arctic and high-Arctic lakes, which were all saturated or  
210 supersaturated with O<sub>2</sub> showed deviating patterns of CO<sub>2</sub> saturation with no significant  
211 correlation between CO<sub>2</sub> and O<sub>2</sub> saturation levels (Fig. S2). Across all three latitudinal zones,  
212 lake O<sub>2</sub> saturation levels showed lower variation than CO<sub>2</sub> saturation levels. Hence, the  
213 variation in CO<sub>2</sub>:O<sub>2</sub> ratio was mainly governed by CO<sub>2</sub> saturation (Fig. 2).

214

### 215 *General patterns among nutrients and conductivity*

216 While high Arctic lakes had lower DOC concentrations than boreal lakes on the mainland,  
217 there were no major differences in DOC concentrations among the boreal and the sub-Arctic  
218 mainland lakes (Table 1). In the boreal lakes, DOC concentrations correlated positively with  
219 TN and total conductivity ( $\rho = 0.48$  and  $\rho = 0.45$ , respectively; Fig. S3). The relation between  
220 DOC and TP was also positive but weak. Conductivity ranged between 6 and 243  $\mu\text{S cm}^{-1}$   
221 with a median value at 24  $\mu\text{S cm}^{-1}$  (Table 1) and correlated best with TN ( $\rho = 0.72$ ; Fig. S3).

222 Similar to the boreal lakes, DOC was positively correlated to TN in both the sub-Arctic and in  
223 the high-Arctic lakes ( $\rho = 0.42$  and  $\rho = 0.65$ , respectively). However, there was no significant  
224 relation between DOC and conductivity in the high-Arctic while in the sub-Arctic, the relation  
225 was weak but negative (Fig. S3). DOC and pH correlated positively in the high-Arctic ( $\rho =$   
226  $0.65$ ) and in the sub-Arctic lakes there was a positive correlation between conductivity and pH  
227 ( $\rho = 0.59$ ). TP was generally lower in the high-Arctic than in the boreal and sub-Arctic lakes  
228 and 12 of the 22 lakes in the high-Arctic had TP concentrations below detection limit.  
229 Conductivity in the high-Arctic lakes was generally higher and had a wider range than in the  
230 boreal lakes (Table 1).  
231 In all lakes, there was a strong positive correlation between conductivity and DIC  
232 concentrations (Fig. S3). In the boreal lakes, DIC concentrations were generally low.  
233 Contrastingly, in the Arctic the relationship between DOC and DIC was either negative (sub-  
234 Arctic) or non-existing (high-Arctic) and thus inorganic compounds are the main sources of  
235 alkalinity in the arctic lakes. Including all lakes in the full model with DOC, TP, TN, and  
236 conductivity as independent variables, the model explained 42 % of the deviance, with  
237 conductivity, TP and DOC concentration as significant predictors (Table 2). Since the  
238 variables predicting the  $\text{CO}_2:\text{O}_2$  ratio differed with geographic location, the models were also  
239 run for each zone separately.

240

#### 241 *Boreal zone*

242 As stated, the  $\text{CO}_2$  and  $\text{O}_2$  saturation were negatively related with each other in the boreal  
243 lakes. A model with DOC as the only explanatory variable explained 35 % of the deviance.  
244 The full model, including TN, TP, and conductivity, improved the deviance explained to 73 %  
245 (Table 2; Fig. 3a). DOC and conductivity were the strongest predictors, both having positive

246 effects on the CO<sub>2</sub>:O<sub>2</sub> ratio. O<sub>2</sub> saturation in the boreal lakes was negatively related to DOC  
247 concentrations and positively related to TN (Fig. S3).

248

#### 249 *High-and sub-Arctic lakes*

250 CO<sub>2</sub>:O<sub>2</sub> ratios in the Arctic lakes were primarily driven by conductivity (positive) and TN  
251 (negative) with the models explaining 75 % and 98 % of the deviance for the sub-Arctic and  
252 the high-Arctic, respectively (Table 2; Fig. 3b and 3c). In the high-Arctic, DOC was a weak  
253 ( $p = 0.04$ ), mainly negative predictor of CO<sub>2</sub>:O<sub>2</sub> ratio while TP was below detection limit in  
254 most lakes and excluded from the analysis. In the sub-Arctic, neither DOC nor TP were  
255 significant predictors of CO<sub>2</sub>:O<sub>2</sub> ratio.

256 All lakes in the Arctic were saturated with O<sub>2</sub>. In the sub-Arctic, O<sub>2</sub> saturation was at 100 %  
257 in all lakes without variation among lakes and, hence, no correlation was found between O<sub>2</sub>  
258 saturation and any other variable. In the high-Arctic, O<sub>2</sub> saturation spanned a wider range  
259 from saturation to oversaturation. The only variable that correlated with O<sub>2</sub> saturation was  
260 conductivity ( $\rho = -0.48$ ). Modelling the CO<sub>2</sub>:O<sub>2</sub> ratio in the high-Arctic lakes with  
261 conductivity as the only independent variable explained 69 % of the deviance. In the sub-  
262 Arctic, 70 % of the deviance was explained with conductivity as the only explanatory  
263 variable.

264

## 265 **Discussion**

### 266 *O<sub>2</sub> and CO<sub>2</sub> saturation*

267 We analyzed saturation levels of O<sub>2</sub> and CO<sub>2</sub> as well as CO<sub>2</sub>:O<sub>2</sub> ratios in surface lake waters  
268 of three different regions of Norway to assess regional differences in lake metabolism related  
269 to catchment properties and key water quality parameters. The study revealed striking  
270 differences between boreal on the one hand, and sub-Arctic and high-Arctic sites on the other

271 hand. The boreal lakes ( $n = 73$ ) all had catchments dominated by coniferous forests, primarily  
272 spruce and pine and the terrestrial C-fixation by forests enrich also lake water with DOC [17,  
273 18, 50], which in turn promote the biogenic production of  $\text{CO}_2$  from bacterial respiration [1, 2,  
274 14, 38]. The positive relationship between  $\text{CO}_2$  saturation and DOC concentration indicates  
275 in-lake  $\text{CO}_2$  production by DOC mineralization, but could also reflect a negative relation  
276 between DOC and retention time, and that more  $\text{CO}_2$  is flushed in from the catchment in  
277 systems with low retention times. On the other hand, short retention times are associated with  
278 inputs of highly reactive DOC [51], yielding enhanced  $\text{CO}_2$  production rates. The two  
279 explanations are thus not mutually exclusive, and in the high-Arctic sites, the  $\text{CO}_2:\text{O}_2$ -ratio  
280 was primarily driven by input of allochthonous  $\text{CO}_2$ .

281 The corresponding negative relationship between DOC concentration and  $\text{O}_2$  saturation  
282 suggests enhanced microbial  $\text{O}_2$  consumption, which may go along with a decrease in primary  
283 production when DOC inputs rise [21, 52]. The net heterotrophy that is prevailing in boreal  
284 DOC-rich lakes is attributed both to reduced photosynthesis in the water column and  
285 enhanced microbial respiration [53]. Together with the negative relation between  $\text{CO}_2$  and  $\text{O}_2$   
286 saturation, such findings indicate that in-lake processes are the main drives of the  $\text{CO}_2:\text{O}_2$   
287 balance and that the level of net heterotrophy increases with increasing DOC concentration in  
288 the boreal lakes.

289 Besides biological processes and air-water exchange, lateral input via groundwater and  
290 surface run-off contribute to lake  $\text{CO}_2$  [54, 55]. Groundwater input has been shown to  
291 correlate well with conductivity, as water in contact with bedrock is likely to become enriched  
292 in ions and minerals. The positive relationship between  $\text{CO}_2$  saturation and conductivity may  
293 thus indicate lateral input of  $\text{CO}_2$  [56]. Further, lake pH may regulate the  $\text{CO}_2$  concentration  
294 with a higher proportion of the DIC staying as free  $\text{CO}_2$  at low pH values while at higher pH  
295 values, more  $\text{CO}_2$  enters the carbonate cycle, forming carbonate and bicarbonate. Boreal lakes

296 are low in alkalinity (Fig. S1) and sensitive to changes in pH. Therefore, besides serving as a  
297 substratum for heterotrophic bacteria, enhanced input of humic acids may result in a decline  
298 in pH and an additional increase in CO<sub>2</sub> saturation [57].

299

### 300 *The role of nutrients*

301 Phosphorus is the major limiting nutrient of bacterioplankton and has been previously shown  
302 to be a strong driver of lake CO<sub>2</sub> supersaturation [13, 58, 59]. The influence of TP on the  
303 CO<sub>2</sub>:O<sub>2</sub> ratio in the boreal lakes was positive up to about 15 µg DOC l<sup>-1</sup>, above which the  
304 effect declined and became negative. However, there were only few observations with high  
305 TP concentrations resulting in wide confidence interval in the second half of the curve (Fig.  
306 3a). The ratio of CO<sub>2</sub>:O<sub>2</sub> could also be affected by the bacterial carbon use efficiency which  
307 depends on the nutrient to C ratio of the substrate with high ratios allowing to allocate more C  
308 to growth while with low ratios, bacteria may dispose excess C as enhanced respiration [60,  
309 61]. Accordingly, CO<sub>2</sub> saturation was negatively related to the TP:DOC ratio in boreal lakes  
310 (Fig. S4), suggesting enhanced rates of CO<sub>2</sub> production in lakes where DOC concentrations  
311 were high in relation to TP concentrations.

312 Primary production in boreal lakes has been shown to be strongly affected by nitrogen  
313 availability in areas receiving low input of N [62] and the positive effect of TN on O<sub>2</sub>  
314 saturation likely reflects increased primary production with increased TN concentrations.

315

### 316 *Different dynamics in high-latitude lakes*

317 A main result of our study was the difference between boreal and Arctic catchments in terms  
318 of CO<sub>2</sub> and thus CO<sub>2</sub>:O<sub>2</sub>-ratio. In contrast to the boreal lakes, DOC concentration did not  
319 appear to drive the CO<sub>2</sub>:O<sub>2</sub> ratio in Arctic lakes, and there was no significant difference in  
320 CO<sub>2</sub> saturation between lakes with high and low DOC concentrations. This was consistent

321 both for sub-Arctic and high-Arctic sites despite the difference in soils and bogs, and points to  
322 absence or presence of coniferous forests in the catchment as a major determinant. This is also  
323 in support of a recent comparison between boreal (forested) and Arctic sites [41]. The  
324 majority of the studied Arctic lakes were saturated or oversaturated with both O<sub>2</sub> and CO<sub>2</sub>  
325 without any relation between O<sub>2</sub> and CO<sub>2</sub> saturation level. These lakes are generally shallow  
326 and low in productivity. Hence, O<sub>2</sub> saturation is most likely a result of downward mixing  
327 rather than primary production.

328 Among the significant predictors of the CO<sub>2</sub>:O<sub>2</sub> ratio in the Arctic, conductivity was clearly  
329 predominant (Table 2; Fig. 3b and 3c). This suggests that the majority of lake CO<sub>2</sub> in the  
330 Arctic entered from the surrounding mineral soils [63] and was not produced through in-lake  
331 DOC mineralization. For some high-Arctic sites, conductivity can also be coupled to their  
332 proximity to the sea. The lakes closest to the sea are also the lakes most influenced by  
333 vegetation and most frequently visited by birds, yielding enhanced DOC and nutrient input  
334 via bird feces [43].

335 While CO<sub>2</sub> saturation increased with conductivity, we observed no correlation between  
336 conductivity and DOC concentration in neither the sub-Arctic nor the high-Arctic. Instead,  
337 conductivity was closely correlated with DIC concentrations and total alkalinity in both Arctic  
338 regions. In the sub-Arctic, CO<sub>2</sub> saturation spanned from undersaturated to supersaturated.

339 Saturation level was closely related to conductivity and thus also to total alkalinity. Alkalinity  
340 in turn could be coupled to bedrock, indicating weathering to be a source of lake DIC.

341 Likewise, the relatively high alkalinity together with the lime-rich bedrock suggests carbonate  
342 weathering to be a DIC source also in the high-Arctic lakes [41, 64].

343 In the high-Arctic lakes, O<sub>2</sub> saturation correlated negatively with conductivity. While all lakes  
344 were supersaturated with O<sub>2</sub>, there was more variation in CO<sub>2</sub> saturation with 7 out of 22  
345 lakes being undersaturated or close to saturation and the rest supersaturated. The seven CO<sub>2</sub>



346 undersaturated lakes were all high in DOC and TN and highly influenced by birds. Many high  
347 latitude lakes are naturally poor in nutrients and although there was no significant relation  
348 between TN and O<sub>2</sub> saturation, additions of N via birds may stimulate primary production  
349 with a concomitant drawdown of CO<sub>2</sub>.

350 The sub-Arctic lakes were all close to O<sub>2</sub> saturation with little variation among lakes (95 % -  
351 99 %), meaning that the variability in CO<sub>2</sub>:O<sub>2</sub> ratio was determined by CO<sub>2</sub> alone. Most  
352 studies suggest that arctic lakes are net heterotrophic, as they are net emitters of CO<sub>2</sub> to the  
353 atmosphere [2, 65]. The majority of Arctic lakes in our study was indeed supersaturated and  
354 net emitters of CO<sub>2</sub> at the time of sampling. However, the uncoupling of CO<sub>2</sub>:O<sub>2</sub> ratio from  
355 DOC and of CO<sub>2</sub> and O<sub>2</sub> saturation indicate an allochthonous CO<sub>2</sub> source rather than in-lake  
356 heterotrophic respiration. The high O<sub>2</sub> saturation levels together with a dominance of  
357 allochthonous DIC suggest autotrophy rather than heterotrophy in the arctic lakes studied  
358 here. In contrast to other parts of the arctic where permafrost cover vast areas, permafrost in  
359 the Scandinavian arctic is generally found in palsas or in mountainous areas. Although  
360 allochthonous DIC may govern the CO<sub>2</sub> in the arctic lakes in this study, heterotrophic  
361 mineralization of DOC may thus have an impact on CO<sub>2</sub> saturation in arctic lakes affected by  
362 permafrost thawing.

363

### 364 *Conclusion*

365 Based on a large number of (mostly) boreal lakes, Larsen et al [38] claimed DOC to be a  
366 universal predictor of lake pCO<sub>2</sub>, while groundwater influx was a minor contributor.

367 However, including also sub- and high-Arctic lakes, we found a clear distinction in drivers of  
368 CO<sub>2</sub> saturation along a latitudinal gradient. The boreal lakes followed the expected pattern  
369 with both CO<sub>2</sub> and O<sub>2</sub> saturation being largely dependent on DOC concentrations and relating  
370 negatively to each other, suggesting enhanced net heterotrophy with increased DOC inputs. In

371 the Arctic lakes, despite the differences between sub-Arctic and high-Arctic sites, there was  
372 no correlation between DOC and CO<sub>2</sub>, yet these sites were also to a large degree  
373 supersaturated with CO<sub>2</sub> and thus could be considered net heterotrophic. However, most of  
374 these lakes were also saturated or supersaturated with O<sub>2</sub>, indicative of low respiratory  
375 activity in agreement with generally nutrient-poor conditions and low levels of primary  
376 production. This is supported by the positive correlation between CO<sub>2</sub>:O<sub>2</sub> ratio and  
377 conductivity, while the influence of DOC concentration was weak or non-significant. This  
378 may suggest that the major share of CO<sub>2</sub> in these lakes is of allochthonous origin, likely from  
379 organic carbon mineralization and carbonate weathering in the catchment soils, entering via  
380 groundwater flow [41, 56]. This points to fundamentally different links between DOC and  
381 CO<sub>2</sub> in boreal and Arctic catchments, pinpointing the fundamental role of coniferous forests  
382 for organic and inorganic carbon dynamics in lakes.

383

384

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389

### 390 **Author contribution**

391 Lina Alleesson and Dag Hessen conceived the idea. Lina Alleesson, Peter Dörsch and Nicolas  
392 Valiente Parra performed material preparation and data collection. All authors were involved  
393 in the analysis of data and final writing and approved the submitted version.

394

### 395 **Data availability statement**

396 All data will be made available upon request, or if demanded, stored in a UiO repository (Git-  
397 HUB)  
398

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548

## Figures

Figure 1

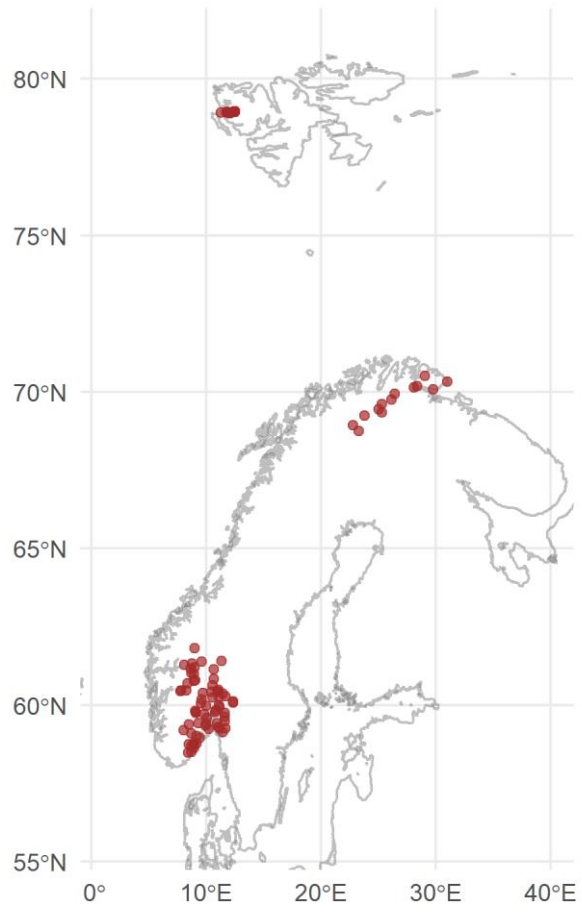


Figure 2

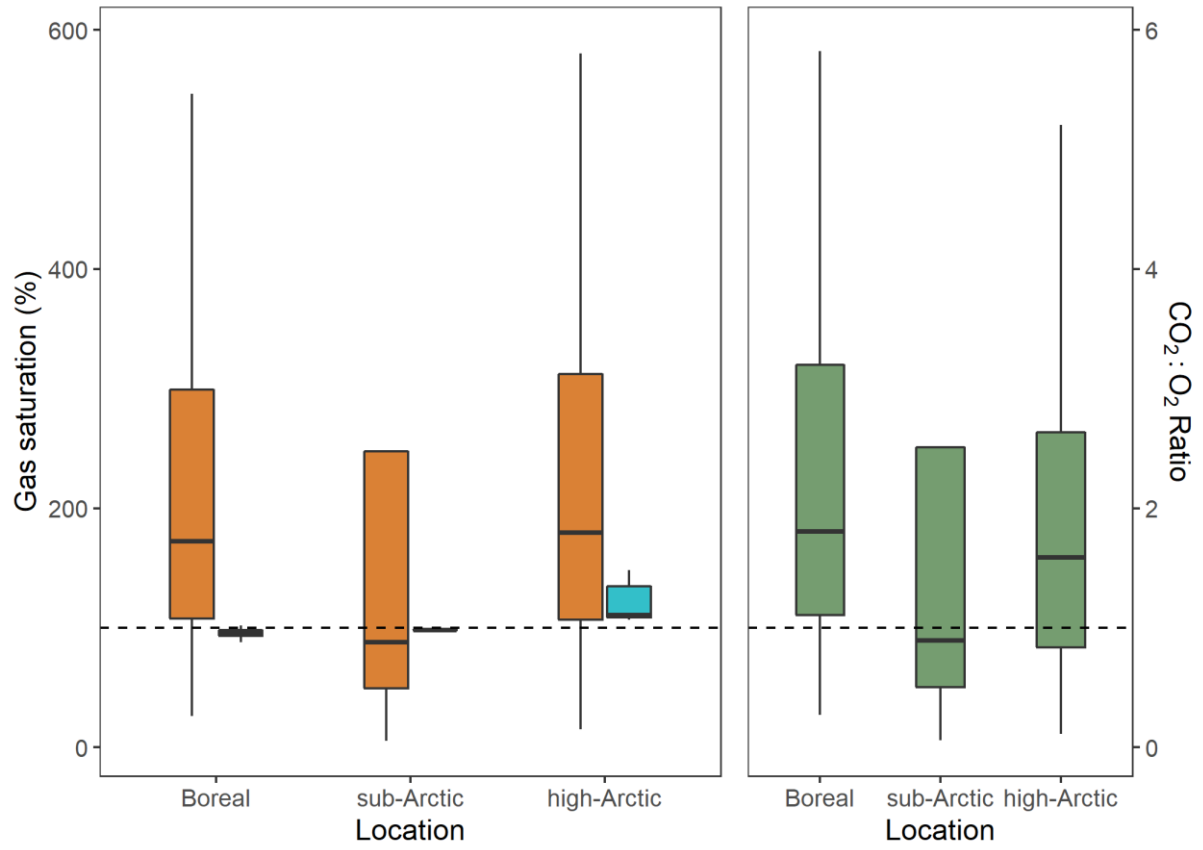
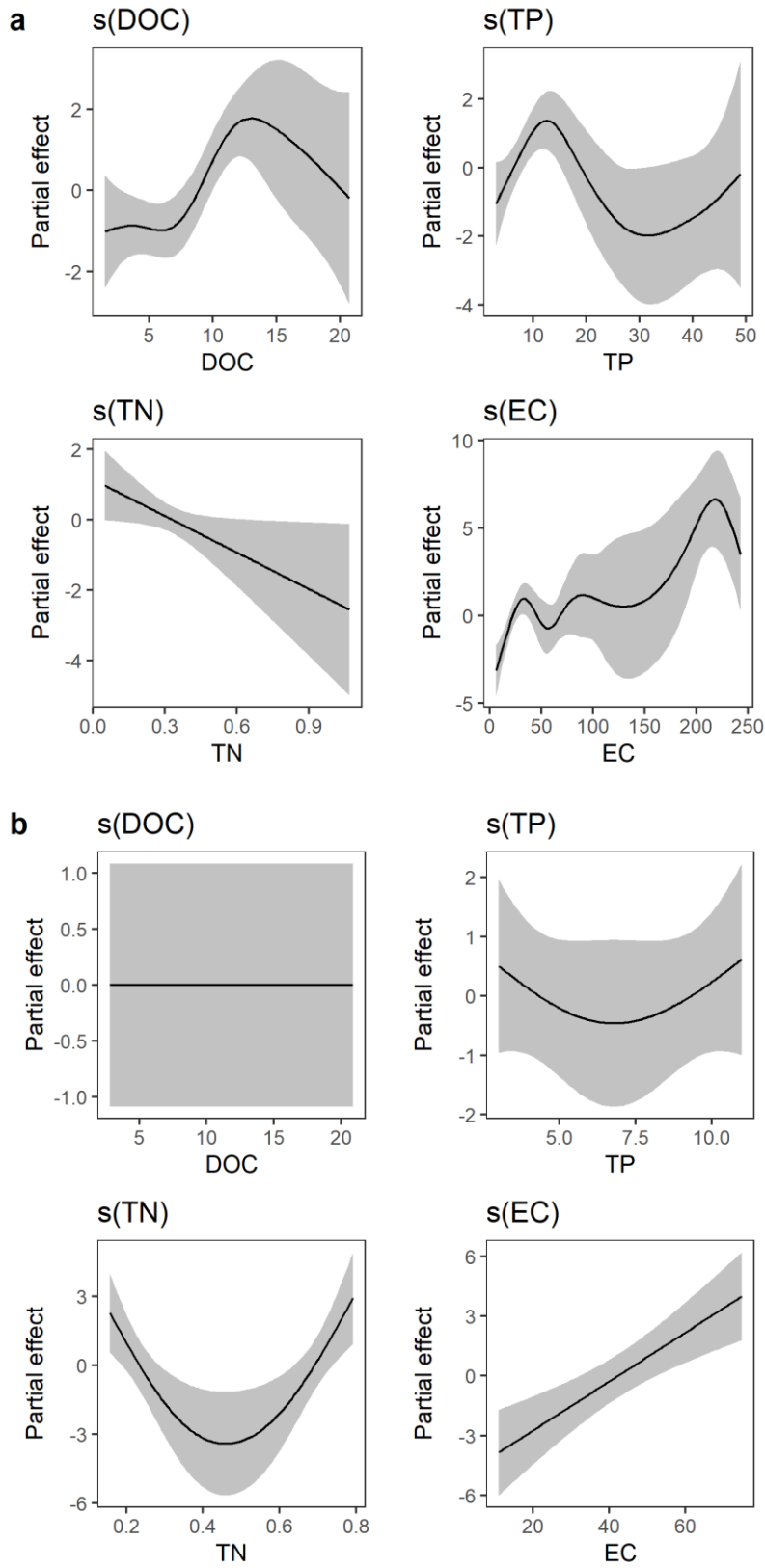
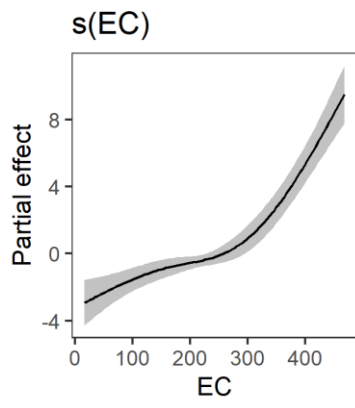
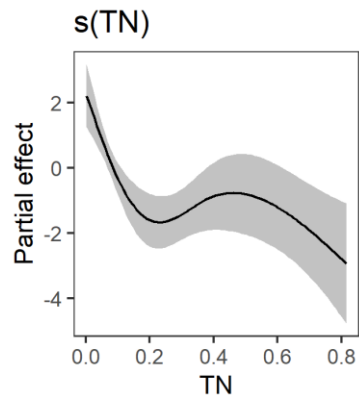
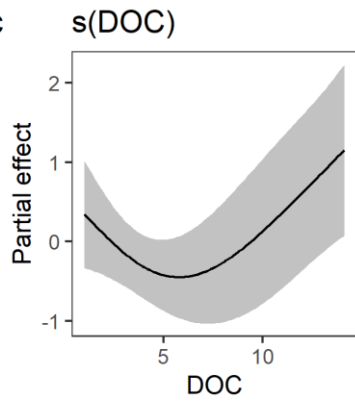




Figure 3



**c**



**Table 1.** Statistics of the lake chemistry variables used in the analyses.

	Boreal	Sub-Arctic	High-Arctic
CO <sub>2</sub> saturation range (%; median; average)	26 – 1374 (175; 260)	6 – 1218 (88; 262)	15 – 1121 (180; 245)
O <sub>2</sub> saturation range (%; median; average)	85 – 102 (96; 95)	95 – 99 (98; 98)	107 – 148 (111; 120)
DOC range (mg l <sup>-1</sup> ; median; average)	1.5 – 20.7 (7.3; 7.7)	2.8 – 20.9 (6.6; 8)	1 – 14 (4; 4.9)
TP range (µg l <sup>-1</sup> ; median; average)	3 – 49 (9; 12)	3 – 11 (6; 6.5)	2 – 19 (3; 6.2) **
TN range (mg l <sup>-1</sup> ; median; average)	0.05 – 1.07 (0.27; 0.33)	0.16 – 0.79 (0.27; 0.35)	0.003 – 0.8 (0.08; 0.2)
Conductivity range (µS cm <sup>-1</sup> ; median; average)	6 – 243 (24.5; 42)	11 – 74.8 (40; 44.4)	16.5 – 1410 (212; 269)

\*\*12 out of 22 lakes with concentrations below detection limit.

**Table 2.** Model outputs of the general additive models (CO<sub>2</sub>:O<sub>2</sub>~ s(DOC) + s(TP) + s(TN) + s(EC)) for the different regions. Significance codes: \*\*\* p < 0.001, \*\* p < 0.01, \* p < 0.05, non-sig p > 0.05.

	Variance explained (%)	Significant predictors
All lakes	42	EC***, DOC*, TP*, TN <sup>non-sig</sup>
Boreal lakes	73	DOC***, TP**, EC**, TN*
Sub- Arctic lakes	75	EC***, TN**, DOC <sup>non-sig</sup> , TP <sup>non-sig</sup>
High-Arctic lakes	94	EC***, TN**, DOC*

## Figure captions

**Fig. 1** Map of the site locations

**Fig. 2** Box plot of lake CO<sub>2</sub> saturation (red), O<sub>2</sub> saturation (blue), and CO<sub>2</sub>:O<sub>2</sub> ratio (green) in the different geographical regions. Dashed line represents the 100 % saturation (to the left) and a CO<sub>2</sub>:O<sub>2</sub> ratio of 1 (to the right)

**Fig. 3** Result plots of the generalized additive models (gams) predicting CO<sub>2</sub>:O<sub>2</sub> saturation ratio for the three regions. In the Boreal lakes (a), the best predictor was DOC concentration followed by conductivity. In both the sub-Arctic (b) and high-Arctic (c), conductivity was the best predictor for CO<sub>2</sub>:O<sub>2</sub> saturation ratio

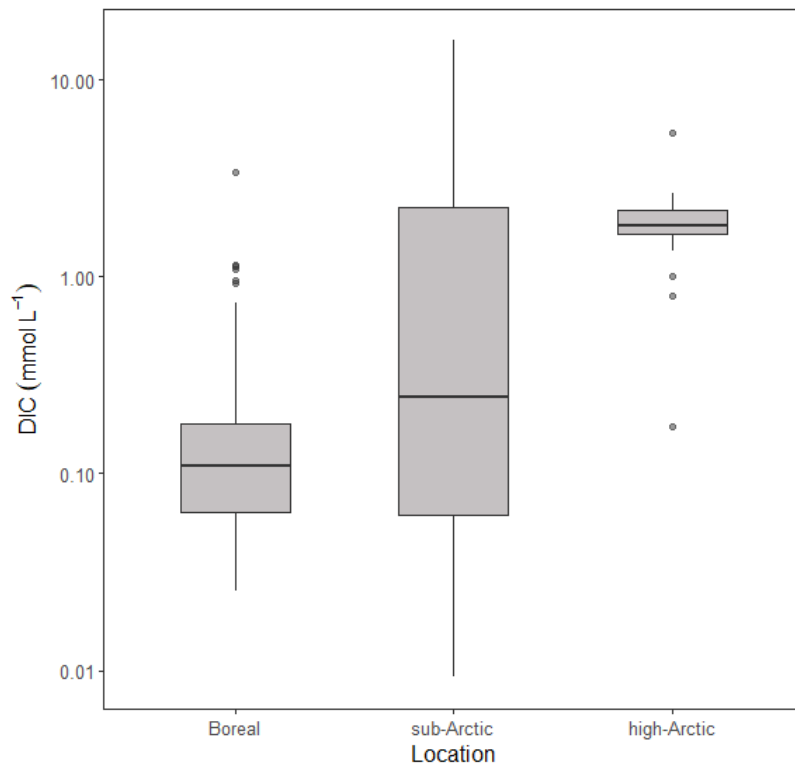


Fig. S1 Boxplot of total DIC concentration in the three regions. In the Boreal part, where granitic and metamorphic bedrock dominates, the alkalinity is generally low. The wide variability in alkalinity in the sub-Arctic is a result of the variability in bedrock from granitic to sedimentary (slate). In the high-Arctic, the majority of the lakes are situated on limestone bedrock giving elevated alkalinity.

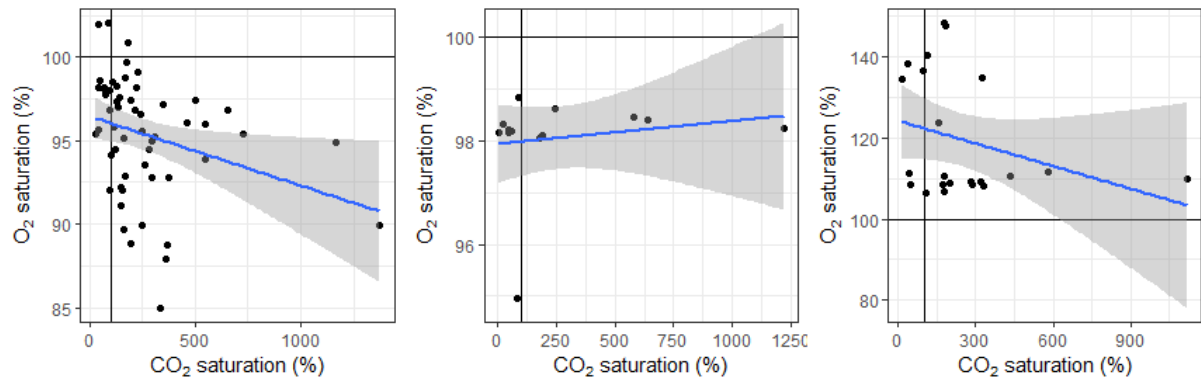
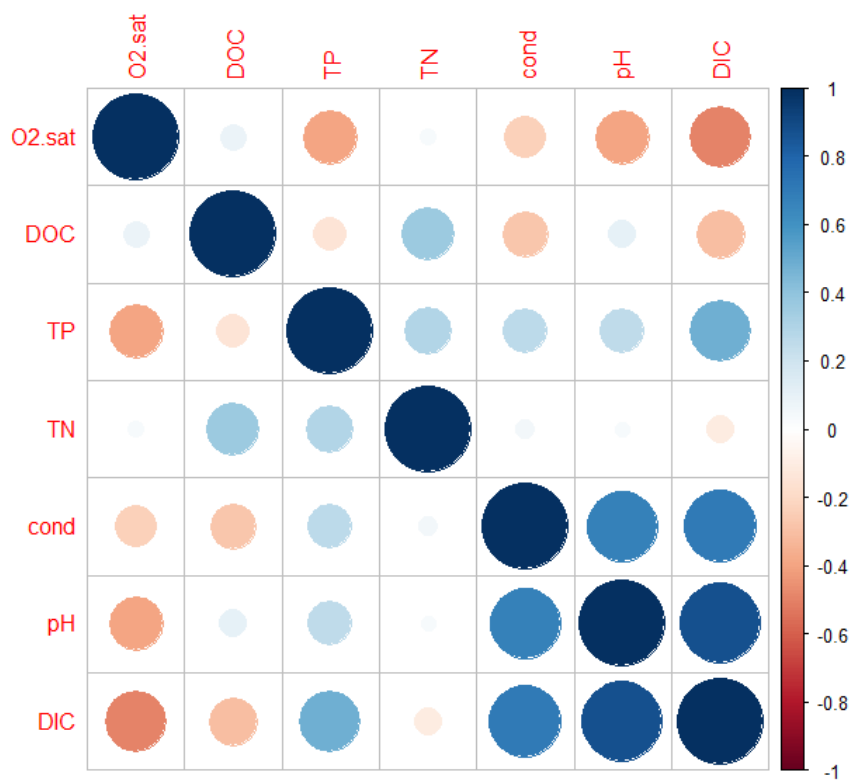
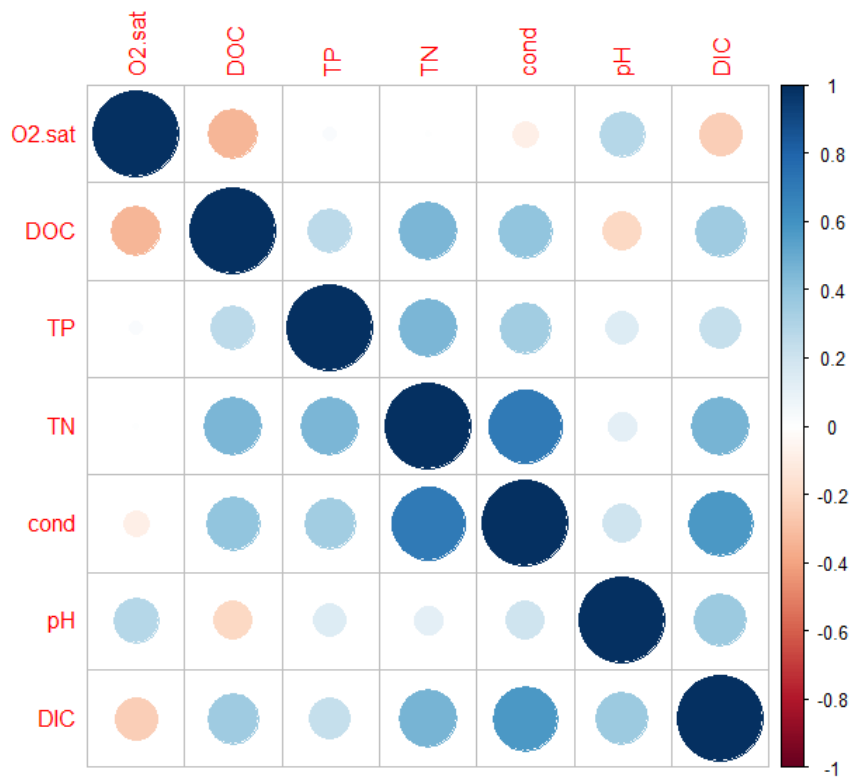


Fig. S2 Scatterplots of O<sub>2</sub> saturation vs. CO<sub>2</sub> saturation. From left to right Boreal lakes, sub-Arctic lakes, high-Arctic lakes. Although quite some scattering, there was a weak but significant negative relation between O<sub>2</sub> saturation and CO<sub>2</sub> saturation in the boreal lakes ( $\rho = -0.42$ ). The relation was stronger in lakes with CO<sub>2</sub> saturation below 500 %. In the northern lakes (sub-Arctic and high-Arctic) where all lakes were saturated or subsaturated with O<sub>2</sub>, there was no significant relation between O<sub>2</sub> and CO<sub>2</sub> saturation. Note differences in axes, lines indicate 100 % saturation.



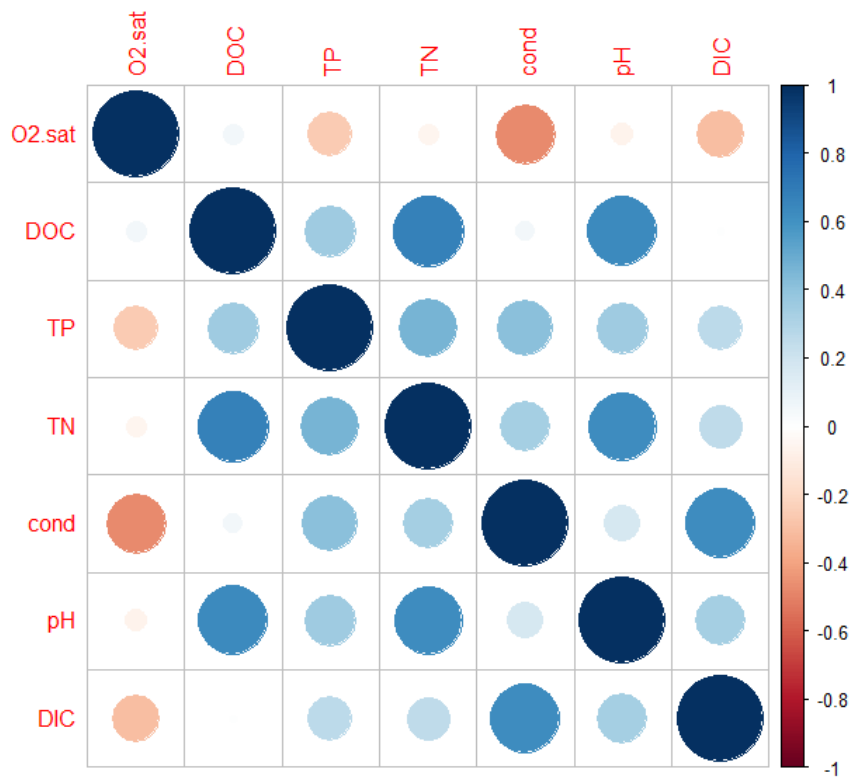


Fig. S3 Spearman's correlation matrices of lake chemistry variables and O<sub>2</sub> saturation. Top to bottom: Boreal lakes, sub-Arctic lakes, high-Arctic lakes.



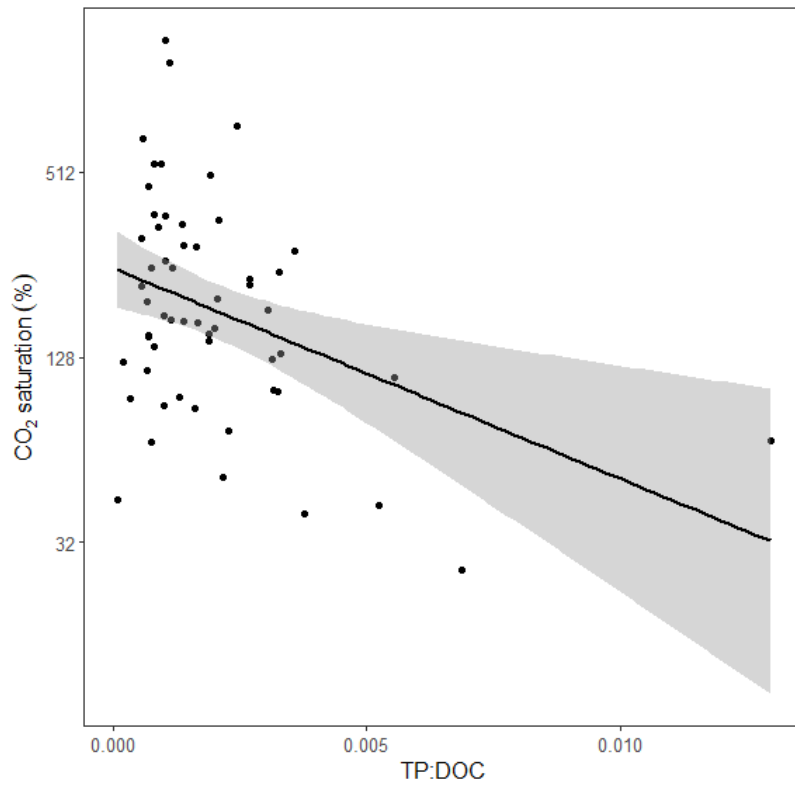


Fig. S4 CO<sub>2</sub> saturation vs TP:DOC ratio in the boreal lakes. The CO<sub>2</sub> saturation level decreases as TP:DOC increases.