

A global database of nitrogen and phosphorus excretion rates of aquatic animals

Michael J. Vanni^{1,*}, Peter B. McIntyre^{2*}, Dennis Allen³, Diane L. Arnott⁴, Jonathan P. Benstead⁵, David J. Berg⁶, Åge Brabrand⁷, Sébastien Brosse⁸, Paul A. Bukaveckas⁹, Adriano Caliman¹⁰, Krista A. Capps¹¹, Luciana S. Carneiro¹⁰, Nanette E. Chadwick¹², Alan D. Christian¹³, Andrew Clarke¹⁴, Joseph D. Conroy¹⁵, Wyatt F. Cross¹⁶, David A. Culver¹⁷, Christopher M. Dalton¹⁸, Jennifer A. Devine¹⁹, Leah M. Domine²⁰, Michelle A. Evans-White²¹, Bjørn A. Faafeng²², Alexander S. Flecker²³, Keith B. Gido²⁴, Claire Godinot²⁵, Rafael D. Guariento²⁶, Susanne Haertel-Borer²⁷, Robert O. Hall²⁸, Raoul Henry²⁹, Brian R. Herwig³⁰, Brendan J. Hicks³¹, Karen A. Higgins³², James M. Hood¹⁷, Matthew E. Hopton³³, Tsutomu Ikeda³⁴, William F. James³⁵, Henrice M. Jansen^{19,36}, Cody R. Johnson^{37,67}, Benjamin J. Koch³⁸, Gary A. Lamberti³⁹, Stephanie Lessard-Pilon⁴⁰, John C. Maerz⁴¹, Martha E. Mather^{1,68}, Ryan A. McManamay⁴², Joseph R. Milanovich⁴³, Dai K.J. Morgan⁴⁴, Jennifer M. Moslemi⁴⁵, Rahmat Naddafi⁴⁶, Jens Petter Nilssen⁴⁷, Marc Pagano⁴⁸, Alberto Pilati⁴⁹, David M. Post⁵⁰, Modi Roopin⁵¹, Amanda T. Rugenski²³, Maynard H. Schaus⁵², Joseph Shostell⁵³, Gaston E. Small²⁰, Christopher T. Solomon⁵⁴, Sean C. Sterrett⁵⁵, Øivind Strand¹⁹, Marjo Tarvainen⁵⁶, Jason M. Taylor⁵⁷, Lisette E. Torres-Gerald⁵⁸, Caroline B. Turner⁵⁹, Jotaro Urabe⁶⁰, Shin-Ichi Uye⁶¹, Anne-Mari Ventelä⁵⁶, Sébastien Villéger⁶², Matt R. Whiles⁶³, Frank M. Wilhelm⁶⁴, Henry F. Wilson⁶⁵, Marguerite A. Xenopoulos⁶⁶, Kyle D. Zimmer²⁰

¹Department of Biology, Miami University, Oxford, OH 45056 USA

²Center for Limnology, University of Wisconsin, Madison, WI 53706 USA

³Baruch Marine Field Lab, University of South Carolina, Georgetown, SC

⁴Centre for Neonatal Research and Education, University of Western Australia, Perth, WA 6009 Australia

⁵Department of Biological Sciences, University of Alabama, Tuscaloosa, AL 35487 USA

⁶Department of Biology, Miami University, Hamilton, OH 45011 USA

⁷Natural History Museum, University of Oslo, P.O. Box 1172 Blindern, NO-0318 Oslo, Norway

⁸Laboratoire Évolution et Diversité Biologique, Université Paul Sabatier, F-31062 Toulouse, France

⁹Department of Biology, Virginia Commonwealth University, Richmond, VA 23284 USA

¹⁰Department of Ecology, Federal University of Rio Grande do Norte, Natal, RN, Brazil

¹¹Odum School of Ecology, University of Georgia, Athens, GA 30602-2202 USA

¹²Department of Biological Sciences, Auburn University, Auburn, AL 36849-5407 USA

¹³School for the Environment, University of Massachusetts Boston, Boston, MA 02125 USA

¹⁴British Antarctic Survey, High Cross, Cambridge CB3 0ET, UK

¹⁵Inland Fisheries Research Unit, Ohio Department of Natural Resources-Division of Wildlife, 10517 Canal Road SE, Hebron, OH 43025 USA

¹⁶Department of Ecology, Montana State University, Bozeman, MT 59717 USA

¹⁷Department of Evolution, Ecology and Organismal Biology, Ohio State University, Columbus, OH 43210 USA

¹⁸The Rivers School, 333 Winter Street, Boston, MA 02493 USA

¹⁹Institute of Marine Research, PO Box 1870 Nordnes, 5817 Bergen, Norway

²⁰Department of Biology, University of St. Thomas, Saint Paul, MN 55105 USA

²¹Department of Biological Sciences, University of Arkansas, Fayetteville, AR 72701 USA

²²Norwegian Institute for Water Research, Gaustadalléen 21, NO-0349 Oslo, Norway

²³Department of Ecology and Evolutionary Biology, Cornell University, Ithaca, NY 14853 USA

²⁴Division of Biology, Kansas State University, Manhattan, KS 66506 USA

²⁵Institut de Radioprotection et de Sûreté Nucléaire, IRSN/PRP-ENV/SERIS/LRD, Laboratoire de Radioprotection de, Cherbourg-Octeville, Rue Max Pol Fouchet, BP10, 50130 Cherbourg-Octeville, France

- 26 Laboratory of Ecology, CCBS, Federal University of Mato Grosso do Sul, Campo Grande, MS, Brazil
- 27 Water Division, Federal Office for the Environment FOEN, CH-3003 Bern, Switzerland
- 28 Department of Zoology and Physiology, University of Wyoming, Laramie, WY 82071 USA
- 29 Department of Zoology, Institute of Biosciences, São Paulo State University, 18618-689 - Botucatu, SP, Brasil
- 30 Fisheries Research, Minnesota Department of Natural Resources, Bemidji, MN 56601 USA
- 31 School of Science, University of Waikato, Private Bag 3105, Hamilton 3240, New Zealand
- 32 North Carolina Department of Environment and Natural Resources, Division of Water Quality, 1617 Mail Service Center, Raleigh, NC 27699 USA
- 33 US Environmental Protection Agency, Office of Research and Development, National Risk Management Research Laboratory, 26 W. Martin Luther King Drive, Cincinnati, OH 45268 USA
- 34 16-3-1001 Toyokawa-cho, Hakodate 040-0065, Japan
- 35 Biology Department, University of Wisconsin-Stout, Menomonie, WI 54751 USA
- 36 Wageningen Marine Research, PO Box 77, 4400 AB Yerseke, The Netherlands
- 37 Department of Watershed Sciences, Utah State University, Logan, UT 84322 USA
- 38 Center for Ecosystem Science and Society, Northern Arizona University, Flagstaff, AZ 86011 USA
- 39 Department of Biological Sciences, University of Notre Dame, Notre Dame, IN 46556 USA
- 40 Smithsonian-Mason School of Conservation, George Mason University, 1500 Remount Road, Front Royal, VA 22360 USA
- 41 Warnell School of Forestry and Natural Resources, University of Georgia, Athens, GA 30602 USA
- 42 Oak Ridge National Laboratory, Environmental Sciences Division, Oak Ridge, TN 37831-6351 USA
- 43 Department of Biology, Loyola University Chicago, Chicago, IL 60660 USA
- 44 Applied and Environmental Sciences, NorthTec, 57 Raumanga Valley Road, Private Bag 9019, Whangarei 0110, New Zealand
- 45 Caravan Lab, 5729 36th Ave. NE, Seattle, WA 98105 USA
- 46 Swedish University of Agricultural Sciences, Department of Aquatic Resources, Institute of Coastal Research, 74242 Öregrund, Sweden
- 47 Free Basic Research, Müller – Sars Society, P.O. Box 5831, N-0308 Oslo, Norway
- 48 Mediterranean Institute of Oceanography, 235 Campus de Luminy Case 901, 13288 Marseille Cedex 09, France
- 49 Departamento de Recursos Naturales, Facultad de Ciencias Exactas y Naturales, Universidad Nacional de La Pampa, Uruguay 151, 6300 Santa Rosa, La Pampa, Argentina
- 50 Department of Ecology and Evolutionary Biology, Yale University, New Haven, CT 06520 USA
- 51 Department of Molecular Cell Biology, Weizmann Institute of Science, Rehovot 7610001, Israel
- 52 Department of Biology, Virginia Wesleyan College, Norfolk, VA 23502 USA
- 53 Math, Science and Technology Department, University of Minnesota-Crookston, Crookston, MN 56716 USA
- 54 Cary Institute of Ecosystem Studies, Millbrook, NY 12545 USA
- 55 US Geological Survey, Patuxent Wildlife Research Center, SO Conte Anadromous Fish Laboratory, Turners Falls, MA 01376 USA
- 56 Pyhäjärvi Institute, Sepäntie 7, FI-27500 Kauttua, Finland
- 57 US Department of Agriculture-Agricultural Research Service, National Sedimentation Laboratory, Water Quality & Ecology Research Unit, 598 McElroy Drive, Oxford, MS 38655 USA
- 58 Cooper Foundation Center for Academic Resources, Nebraska Wesleyan University, Lincoln, NE 68504 USA
- 59 Department of Microbiology and Molecular Genetics, University of Pittsburgh, Pittsburgh, PA 15219 USA
- 60 Graduate School of Life Sciences, Tohoku University, 6-3 Aramaki, Aoba-ku, Sendai, 980-8578 Japan
- 61 Graduate School of Biosphere Science, Hiroshima University, 4-4 Kagamiyama 1 Chome, Higashi-Hiroshima 739-

8528, Japan

⁶²CNRS Laboratoire Biodiversité Marine et ses Usages, Université de Montpellier, F-34095 Montpellier Cedex 5, France

⁶³Department of Zoology and Center for Ecology, Southern Illinois University, Carbondale, IL 62901-6501 USA

⁶⁴Department of Fish and Wildlife Sciences, University of Idaho, Moscow, ID 83844 USA

⁶⁵Agriculture and Agri-Food Canada, Brandon Research Centre, PO Box 1000A, R.R. #3, Brandon, Manitoba R7A 5Y3, Canada

⁶⁶Department of Biology, Trent University, Peterborough, Ontario, K9L 0G2, Canada

⁶⁷ Present address: Polar Field Services Inc., 8100 Shaffer Parkway #100, Littleton, CO 80127 USA

⁶⁸Present address: US Geological Survey, Kansas Cooperative Fish and Wildlife Research Unit, Division of Biology, Kansas State University, Manhattan, KS 66506

*Equal contribution first authors

⁺Corresponding author: yannimj@miamioh.edu

INTRODUCTION

The recycling of nutrients by animals is important at many levels of ecological organization. At the level of the individual, the rates at which animals recycle nutrients (egestion of solids and excretion of dissolved molecules) are important because they can be used to explore theories related to Metabolic Ecology (Gillooly et al. 2001; Brown et al. 2004) and Ecological Stoichiometry (ES, Sterner and Elser 2002). Metabolic Ecology predicts that biological rates of individuals (for example, nutrient excretion rates) are power functions of body size (Gillooly et al. 2001). Body size dependence is usually captured in the formula $B = B_0M^b$, where B is individual metabolic rate (e.g., oxygen consumed, or nitrogen excreted, per individual per unit time), B_0 is a ‘normalization constant,’ M is organism body mass, and b is the ‘scaling coefficient.’ A prominent version of ME, the Metabolic Theory Ecology (MTE), predicts that b is ~ 0.75 for most biological rates (West et al. 1997; Gillooly et al. 2001), although there is controversy regarding the theory and empirical evidence for this value (White and Seymour 2005; Glazier 2010; Isaac and Carbone 2010). The MTE framework also recognizes the importance of temperature; most biological rates increase exponentially with temperature over most of the thermal tolerance range of an organism (Gillooly et al. 2001, Clarke 2004). MTE has focused mostly on the allometry and temperature dependence of metabolic rates (e.g., oxygen consumption). However, the theory is applicable to other biological rates, including excretion rates (Allen and Gillooly 2009). Several studies have examined the allometry of excretion rates, and scaling coefficients vary greatly among taxa and studies (e.g., Hall et al. 2007; Sereda et al. 2010).

Ecological Stoichiometry theory (ES) predicts that nutrient excretion rates and ratios of consumers are functions of the imbalance between the nutrient content of the organism’s body versus that of its food source. ES usually focuses on nitrogen (N) and phosphorus (P), and their ratio (N:P; Sterner and Elser 2002). For example, ES predicts that an animal with a high concentration of P in its body (i.e., low body N:P) will sequester more dietary P to grow, compared to a

counterpart with low body P (high body N:P). As a consequence, the animal with low body N:P will release wastes at a higher N:P than its counterpart with high body N:P. More generally, across individuals or species, body N:P and waste N:P should be negative correlated (Sternner 1990; Sternner and Elser 2002). Differences among animals in body N:P are often driven by differences in the allocation of P-rich structures such as RNA and bone (Elser et al. 1996; Vanni et al. 2002). ES also recognizes the importance of dietary nutrients in driving nutrient excretion rates and ratios; specifically, consumers whose diet is rich in a particular element should release that element at higher rates than a counterpart consuming a diet that is deficient in that element, given similar body elemental compositions (Sternner 1990; Sternner and Elser 2002). Thus, ES also predicts that food N:P will be positively correlated with the N:P of wastes.

Animals can be important agents of nutrient cycling at the ecosystem level. In some ecosystems, the release of potentially limiting nutrients by animals can sustain a substantial proportion of primary production (McNaughton et al. 1997; Vanni 2002; McIntyre et al. 2008). However, the importance of animals in nutrient cycling varies greatly among species and ecosystems (Vanni 2002), in part because in most ecosystems, animal excretion is but one of many fluxes and transformations of nutrients mediated by animals, microbes and physical processes (e.g. Wood et al. 2016). For example, across aquatic ecosystems, nutrient excretion by animal assemblages can support anywhere from <5% to >80% of algal primary production, depending on the ecosystem. Furthermore, the relative importance of different animal groups (e.g., zooplankton, benthic invertebrates, fish) varies greatly among ecosystems (Taylor et al. 2015).

Given the potential ecosystem-level importance of animal-mediated nutrient cycling, and the potential value of data on animal nutrient recycling rates for testing predictions of Metabolic Ecology and Ecological Stoichiometry, a comprehensive compilation of animal nutrient excretion rates can be of great use to the ecological community. The number of studies of animal mediated nutrient cycling has increased greatly in recent decades (Taylor et al. 2015). However, a comprehensive compilation of excretion rates has not been available. Here, we combine published and unpublished data to create a dataset that includes 10,534 observations of N or P excretion rates of animals from freshwater and marine ecosystems worldwide.

This data set was used recently to test predictions of MTE and ES, as described in Vanni and McIntyre (2016). The main findings of that paper are that body size is by far the best predictor of excretion rates, followed by temperature. Whether an animal was a vertebrate or invertebrate was also a significant predictor, with vertebrates excreting both N and P at higher rates than invertebrates, after accounting for body size and temperature. Other predictors based on ecological stoichiometry, such as the N:P ratio of animal bodies or their food resources, explained very little variance in excretion rates or N:P excreted, once body size, temperature, and the vertebrate/invertebrate classification were accounted for. The

temperature dependence of excretion was stronger for N than P, and thus N:P excreted increased with temperature. Allometric scaling coefficients differed for N and P, were significantly less than 0.75 for both elements, and varied greatly among species. While these findings shed light on the variation in excretion rates among animal taxa, the data set should be useful for additional analyses.

METADATA

Note: Metadata follows the format in Table 1 of Michener et al. (1997). Although we exclude Fields that are not applicable, we maintained their numbering system.

Class I. Data set descriptors

A. Data set identity: A global database of nitrogen and phosphorus excretion rates of aquatic animals

B. Data set identification code:

1. Excretion rates and ancillary data: Aquatic_animal_excretion_data.csv

2. Variable descriptors: Aquatic_animal_excretion_variable_descriptions.csv

C. Data set description

1. Originators

Michael J. Vanni^{1,*}, Peter B. McIntyre^{2*}, Dennis Allen³, Diane L. Arnott⁴, Jonathan P. Benstead⁵, David J. Berg⁶, Åge Brabrand⁷, Sébastien Brosse⁸, Paul A. Bukaveckas⁹, Adriano Caliman¹⁰, Krista A. Capps¹¹, Luciana S. Carneiro¹⁰, Nanette E. Chadwick¹², Alan D. Christian¹³, Andrew Clarke¹⁴, Joseph D. Conroy¹⁵, Wyatt F. Cross¹⁶, David A. Culver¹⁷, Christopher M. Dalton¹⁸, Jennifer A. Devine¹⁹, Leah M. Domine²⁰, Michelle A. Evans-White²¹, Bjørn A. Faafeng²², Alexander S. Flecker²³, Keith B. Gido²⁴, Claire Godinot²⁵, Rafael D. Guariento²⁶, Susanne Haertel-Borer²⁷, Robert O. Hall²⁸, Raoul Henry²⁹, Brian R. Herwig³⁰, Brendan J. Hicks³¹, Karen A. Higgins³², James M. Hood¹⁷, Matthew E. Hopton³³, Tsutomu Ikeda³⁴, William F. James³⁵, Henrice M. Jansen^{19,36}, Cody R. Johnson^{37,67}, Benjamin J. Koch³⁸, Gary A. Lamberti³⁹, Stephanie Lessard-Pilon⁴⁰, John C. Maerz⁴¹, Martha E. Mather^{1,68}, Ryan A. McManamay⁴², Joseph R. Milanovich⁴³, Dai K.J. Morgan⁴⁴, Jennifer M. Moslemi⁴⁵, Rahmat Naddafi⁴⁶, Jens Petter Nilssen⁴⁷, Marc Pagano⁴⁸, Alberto Pilati⁴⁹, David M. Post⁵⁰, Modi Roopin⁵¹, Amanda T. Rugenski²³, Maynard H. Schaus⁵², Joseph Shostell⁵³, Gaston E. Small²⁰, Christopher T. Solomon⁵⁴, Sean C. Sterrett⁵⁵, Øivind Strand¹⁹, Marjo Tarvainen⁵⁶, Jason M. Taylor⁵⁷, Lisette E. Torres-Gerald⁵⁸, Caroline B. Turner⁵⁹, Jotaro Urabe⁶⁰, Shin-Ichi Uye⁶¹, Anne-Mari Ventelä⁵⁶, Sébastien Villéger⁶², Matt R. Whiles⁶³, Frank M. Wilhelm⁶⁴, Henry F. Wilson⁶⁵, Marguerite A. Xenopoulos⁶⁶, Kyle D. Zimmer²⁰

¹Department of Biology, Miami University, Oxford, OH 45056 USA

²Center for Limnology, University of Wisconsin, Madison, WI 53706 USA

³Baruch Marine Field Lab, University of South Carolina, Georgetown, SC

⁴Centre for Neonatal Research and Education, University of Western Australia, Perth, WA 6009 Australia

⁵Department of Biological Sciences, University of Alabama, Tuscaloosa, AL 35487 USA

⁶Department of Biology, Miami University, Hamilton, OH 45011 USA

⁷Natural History Museum, University of Oslo, P.O. Box 1172 Blindern, NO-0318 Oslo, Norway

⁸Laboratoire Évolution et Diversité Biologique, Université Paul Sabatier, F-31062 Toulouse, France

- Department of Biology, Virginia Commonwealth University, Richmond, VA 23284 USA
- 10 Department of Ecology, Federal University of Rio Grande do Norte, Natal, RN, Brazil
 - 11 Odum School of Ecology, University of Georgia, Athens, GA 30602-2202 USA
 - 12 Department of Biological Sciences, Auburn University, Auburn, AL 36849-5407 USA
 - 13 School for the Environment, University of Massachusetts Boston, Boston, MA 02125 USA
 - 14 British Antarctic Survey, High Cross, Cambridge CB3 0ET, UK
 - 15 Inland Fisheries Research Unit, Ohio Department of Natural Resources-Division of Wildlife, 10517 Canal Road SE, Hebron, OH 43025 USA
 - 16 Department of Ecology, Montana State University, Bozeman, MT 59717 USA
 - 17 Department of Evolution, Ecology and Organismal Biology, Ohio State University, Columbus, OH 43210 USA
 - 18 The Rivers School, 333 Winter Street, Boston, MA 02493 USA
 - 19 Institute of Marine Research, PO Box 1870 Nordnes, 5817 Bergen, Norway
 - 20 Department of Biology, University of St. Thomas, Saint Paul, MN 55105 USA
 - 21 Department of Biological Sciences, University of Arkansas, Fayetteville, AR 72701 USA
 - 22 Norwegian Institute for Water Research, Gaustadalléen 21, NO-0349 Oslo, Norway
 - 23 Department of Ecology and Evolutionary Biology, Cornell University, Ithaca, NY 14853 USA
 - 24 Division of Biology, Kansas State University, Manhattan, KS 66506 USA
 - 25 Institut de Radioprotection et de Sûreté Nucléaire, IRSN/PRP-ENV/SERIS/LRD, Laboratoire de Radioprotection de, Cherbourg-Octeville, Rue Max Pol Fouchet, BP10, 50130 Cherbourg-Octeville, France
 - 26 Laboratory of Ecology, CCBS, Federal University of Mato Grosso do Sul, Campo Grande, MS, Brazil
 - 27 Water Division, Federal Office for the Environment FOEN, CH-3003 Bern, Switzerland
 - 28 Department of Zoology and Physiology, University of Wyoming, Laramie, WY 82071 USA
 - 29 Department of Zoology, Institute of Biosciences, São Paulo State University, 18618-689 - Botucatu, SP, Brasil
 - 30 Fisheries Research, Minnesota Department of Natural Resources, Bemidji, MN 56601 USA
 - 31 School of Science, University of Waikato, Private Bag 3105, Hamilton 3240, New Zealand
 - 32 North Carolina Department of Environment and Natural Resources, Division of Water Quality, 1617 Mail Service Center, Raleigh, NC 27699 USA
 - 33 US Environmental Protection Agency, Office of Research and Development, National Risk Management Research Laboratory, 26 W. Martin Luther King Drive, Cincinnati, OH 45268 USA
 - 34 16-3-1001 Toyokawa-cho, Hakodate 040-0065, Japan
 - 35 Biology Department, University of Wisconsin-Stout, Menomonie, WI 54751 USA
 - 36 Wageningen Marine Research, PO Box 77, 4400 AB Yerseke, The Netherlands
 - 37 Department of Watershed Sciences, Utah State University, Logan, UT 84322 USA
 - 38 Center for Ecosystem Science and Society, Northern Arizona University, Flagstaff, AZ 86011 USA
 - 39 Department of Biological Sciences, University of Notre Dame, Notre Dame, IN 46556 USA
 - 40 Smithsonian-Mason School of Conservation, George Mason University, 1500 Remount Road, Front Royal, VA 22360 USA
 - 41 Warnell School of Forestry and Natural Resources, University of Georgia, Athens, GA 30602 USA
 - 42 Oak Ridge National Laboratory, Environmental Sciences Division, Oak Ridge, TN 37831-6351 USA
 - 43 Department of Biology, Loyola University Chicago, Chicago, IL 60660 USA
 - 44 Applied and Environmental Sciences, 57 Raumanga Valley Road, Private Bag 9019, Whangarei 0110, New Zealand
 - 45 Caravan Lab, 5729 36th Ave. NE, Seattle, WA 98105 USA
 - 46 Swedish University of Agricultural Sciences, Department of Aquatic Resources, Institute of Coastal Research, 74242 Öregrund, Sweden

- ⁴⁷Free Basic Research, Müller – Sars Society, P.O. Box 5831, N-0308 Oslo, Norway
- ⁴⁸Mediterranean Institute of Oceanography, 235 Campus de Luminy Case 901, 13288 Marseille Cedex 09, France
- ⁴⁹Departamento de Recursos Naturales, Facultad de Ciencias Exactas y Naturales, Universidad Nacional de La Pampa, Uruguay 151, 6300 Santa Rosa, La Pampa, Argentina
- ⁵⁰Department of Ecology and Evolutionary Biology, Yale University, New Haven, CT 06520 USA
- ⁵¹Department of Molecular Cell Biology, Weizmann Institute of Science, Rehovot 7610001, Israel
- ⁵²Department of Biology, Virginia Wesleyan College, Norfolk, VA 23502 USA
- ⁵³Math, Science and Technology Department, University of Minnesota-Crookston, Crookston, MN 56716 USA
- ⁵⁴Cary Institute of Ecosystem Studies, Millbrook, NY 12545 USA
- ⁵⁵US Geological Survey, Patuxent Wildlife Research Center, SO Conte Anadromous Fish Laboratory, Turners Falls, MA 01376 USA
- ⁵⁶Pyhäjärvi Institute, Sepäntie 7, FI-27500 Kauttua, Finland
- ⁵⁷US Department of Agriculture-Agricultural Research Service, National Sedimentation Laboratory, Water Quality & Ecology Research Unit, 598 McElroy Drive, Oxford, MS 38655 USA
- ⁵⁸Cooper Foundation Center for Academic Resources, Nebraska Wesleyan University, Lincoln, NE 68504 USA
- ⁵⁹Department of Microbiology and Molecular Genetics, University of Pittsburgh, Pittsburgh, PA 15219 USA
- ⁶⁰Graduate School of Life Sciences, Tohoku University, 6-3 Aramaki, Aoba-ku, Sendai, 980-8578 Japan
- ⁶¹Graduate School of Biosphere Science, Hiroshima University, 4-4 Kagamiyama 1 Chome, Higashi-Hiroshima 739-8528, Japan
- ⁶²CNRS Laboratoire Biodiversité Marine et ses Usages, Université de Montpellier, F-34095 Montpellier Cedex 5, France
- ⁶³Department of Zoology and Center for Ecology, Southern Illinois University, Carbondale, IL 62901-6501 USA
- ⁶⁴Department of Fish and Wildlife Sciences, University of Idaho, Moscow, ID 83844 USA
- ⁶⁵Agriculture and Agri-Food Canada, Brandon Research Centre, PO Box 1000A, R.R. #3, Brandon, Manitoba R7A 5Y3, Canada
- ⁶⁶Department of Biology, Trent University, Peterborough, Ontario, K9L 0G2, Canada
- ⁶⁷Present address: Polar Field Services Inc., 8100 Shaffer Parkway #100, Littleton, CO 80127 USA
- ⁶⁸Present address: US Geological Survey, Kansas Cooperative Fish and Wildlife Research Unit, Division of Biology, Kansas State University, Manhattan, KS 66506

2. Abstract

Animals can be important in modulating ecosystem-level nutrient cycling, although their importance varies greatly among species and ecosystems. Nutrient cycling rates of individual animals represent valuable data for testing the predictions of important frameworks such as the Metabolic Theory of Ecology (MTE) and ecological stoichiometry (ES). They also represent an important set of functional traits that may reflect both environmental and phylogenetic influences. Over the past two decades, studies of animal-mediated nutrient cycling have increased dramatically, especially in aquatic ecosystems. Here we present a global compilation of aquatic animal nutrient excretion rates. The dataset includes 10,534 observations from freshwater and marine animals of N and/or P excretion rates. These observations represent 491 species, including most aquatic phyla. Coverage varies greatly among phyla and other taxonomic levels. The dataset includes information on animal body size, ambient temperature, taxonomic affiliations, and animal body N:P. This data set was used to test predictions of MTE and ES, as described in Vanni and McIntyre (2016; Ecology).

D. Key words: *Freshwater and marine ecosystems (lakes, rivers and oceans); invertebrates; vertebrates; body size; ecological stoichiometry; metabolic ecology; nitrogen excretion; nutrient cycling; phosphorus excretion; temperature*

Class II. Research origin descriptors

A. Project description

6. Sources of funding

The primary sources of funding used to compile and synthesize these data were an US National Science Foundation (NSF) OPUS (Opportunities for Promoting Understanding through Synthesis) award to M.J.V. (DEB 0918993) and NSF award DEB-1030242 to P.B.M. This data set contains observations from many studies, and therefore many funding sources, information on which is provided in the acknowledgments sections of the individual papers. In addition, some authors wished to further acknowledge the following sources (funding sources, authors of this paper), and/or to clarify the role of their employer in supporting the research:

- Brazilian Council of Research and Technology (CNPq) Research Productivity grants 304621/2015-3: A. Caliman
- Norwegian Council for Scientific and Industrial Research (currently the Research Council of Norway): Å. Brabrand, B.A. Faafeng, and J.P. Nilssen
- NSF DDEP grant 0951432 and the PADI Foundation: K.A. Capps, and A.S. Flecker
- Ohio Chapter of the Nature Conservancy and the Ouachita National Forrest (A.D. Christian and D.J. Berg)
- British Antarctic Survey (A. Clarke)
- Ohio Lake Erie Protection Fund grant SG189-02, United States Environmental Protection Agency grant GL-97590101, and Federal Aid in Sport Fish Restoration Program grant F-69-P (J.D. Conroy and D.A. Culver)
- Carrying Capacity in Norwegian Aquaculture (CANO) – Research Council of Norway, project no. 173537 (H.M. Jansen and Ø. Strand)
- National Research Council Research Associateship to JR Milanovich at the United States Environmental Protection Agency, Office of Research and Development (J.R. Milanovich and M.E. Hopton)
- New Zealand Ministry of Business, Innovation and Employment contract UOWX0505 (D.J.K. Morgan and B.J. Hicks)
- St. Johns River Water Management District (Florida) contract grant SK933AA (M.H. Schaus)
- Maj and Tor Nessling Foundation (M. Tarvainen and A.-M. Ventelä)
- US EPA STAR Fellowship Assistance Agreement No. FP-91694301-1 (J.M. Taylor)
- Japan Society for the Promotion of Science KAKENHI Grant Number 15H02642 (J. Urabe)
- EU BioFresh project, 7th Framework European program, Contract N°226874 (S. Villegier and S. Brosse)
- Biodiversity Grants Program (University of Alberta, and the Alberta Conservation Association); C/BAR grant from the Canadian Circumpolar Institute; NSERC operating grant #89673S (F.M. Wilhelm)
- Canada Natural Sciences and Engineering Research Council (NSERC) Discovery grant (M.A. Xenopoulos) and NSERC Graduate Scholarship (H.F. Wilson)
- This submission was written by the author(s) acting in their own independent capacity and not on behalf of UT-Battelle, LLC, or its affiliates or successors (R.A. McManamay)

B. “Specific subproject” description

1. Site description

- a. Site type
Sites include freshwater and marine ecosystems from across the globe.
- b. Geography
Marine sites include all five oceans: Atlantic, Arctic, Indian, Pacific and Southern. Freshwater sites included every continent except Antarctica (Africa, Asia, Australia, North America and South America). In aggregate, coverage includes those in polar, temperate and tropical ecosystems.
- c. Habitat
Freshwater sites include lakes, rivers and streams, as well as a small amount of data from wetlands. Marine sites include estuaries, coastal ecosystems, and open ocean.
- d. Geology
Sites include oceans and freshwater sites scattered across the globe and hence associated with various geological formations.
- e. Watersheds, hydrology
Sites include dozens of watersheds spread across the world, ranging from essentially no flow (many lakes) to large rivers.

- f. **Site history**
Sites range from pristine (e.g., lakes in Alaska, the Antarctic Ocean) to highly managed (e.g., reservoirs).
- g. **Climate**
Climate range greatly from polar to tropical. For example, the temperature ranged across observations from -1.9 to 33.5°C.

2. **Experimental design**

a. **Design characteristics**

In general, the design includes measuring excretion rates of field caught animals, which are assumed to have been feeding at 'natural' rates typical for that species in that particular ecosystem. These are 'mensurative' experiments (Hurlbert 1984) in which rates are measured but no experimental treatments are imposed on the animals. Our data set does not include any 'basal rate' data from starved animals.

To find appropriate data, we attempted to do comprehensive searches on Web of Science and Google Scholar, using terms such as "nutrient cycling," "nitrogen excretion," "phosphorus excretion," and related search strings. However, these searches returned several thousands of sources, of which only a tiny proportion included excretion rates measured in the field (the majority of excretion rate measurements obtained in this way were lab studies, with animals provided with controlled diets). Searches using more specific terms such as "animal-mediated nutrient*" or "ecological stoichiometry" returned many fewer sources, but these searches missed many earlier papers published before such terms were commonplace. Therefore, to generate our database we started with keyreview/synthesis papers (Andersson et al. 1988, Sterner 1990, Vanni 2002, Hall et al. 2007, Sereda and Hudson 2011), and scoured these for papers they cited and papers that cited them. We repeated this process for each new paper found, until we were confident we had compiled the vast majority of sources. Our search process was completed in early 2014, and newer papers have been incorporated opportunistically.

Once a potential data source was identified, the two first authors (MJV and PBM) contacted its authors to ask for the original data. In nearly every case, this request was accepted. All individuals providing raw data in this manner, and all 'co-owners' of these data are listed as authors of this data paper. In addition, MJV and PBM contacted other scientists who were likely to have unpublished nutrient excretion data, and contributed unpublished data of their own. Approximately 5% of data were obtained from unpublished sources. Finally, in cases where the author of a paper could not be reached or provide data, but individual-level data were depicted explicitly, we digitized data from published graphics using the free software Digitizeit (<http://www.digitizeit.de/>). All data providers and source papers (when applicable) are identified in the data file.

3. **Research methods**

a. **Field/laboratory**

Investigators obtained excretion rates in the field by collecting animals and incubating them in a fixed volume of water for a measured period of time. The incubation water is usually pre-filtered to remove particles that could take up or release nutrients. For example, NH_4 excreted by animals could be rapidly converted to NO_3 via nitrification (e.g., Moulton et al. 2016), so removal of microbes is essential for accurate estimates of animal excretion rates. Nevertheless, some microbial transformations could have occurred in some experiments, which would result in an underestimation of actual excretion rates. Usually incubation containers were placed in the body of water to maintain ambient temperature but in some experiments incubations are done on shore or on a ship (but most these studies also maintained ambient temperature). For most observations in our dataset, an individual animal was incubated by itself in a container. However, it is sometimes not possible to measure rates on single individuals of small species, because they do not excrete enough N or P for accurate measurements. Thus for the smallest animals, a single rate was often measured on several animals (similar in size) incubated together.

Following incubations, animals were weighed to determine their mass. Vertebrates generally were weighed live to determine their wet mass (WM), and in some cases dry mass (DM) was also provided. Most invertebrates were euthanized and dried before weighing for dry mass (DM), and wet mass was not estimated. We placed all measurements in the same mass units by converting to WM to DM whenever necessary (this was necessary only for some vertebrates). In cases where DM was not measured directly, we estimated it by multiplying WM by a conversion factor of 0.25 (i.e. $\text{DM} = 0.25\text{WM}$). This conversion factor reflects our own measurements of both WM and DM on hundreds of fish, and is in keeping with the literature. For mollusks and turtles, mass measurements represent only soft tissue because structural materials are unlikely to be metabolically active. For

all other taxa, the mass of all body tissues was used.

In the lab, water samples from the incubation containers were analyzed for nitrogen (N) and phosphorus (P). N was usually measured as ammonium (NH₄, >98% of observations) and rarely as total N. P was usually measured as soluble reactive P (SRP, 87% of observations), but sometimes as total dissolved P or total P. Excretion rates are calculated as the difference in element mass (N or P) over time from the beginning to the end of the incubations. Nutrient mass at the beginning of an incubation was generally inferred from either incubations with no animal, or samples from a common water source before it was aliquoted into incubation containers. Incubation times typically lasted from 15 minutes to 24 hours, but most incubations were relatively short (median 1.3 h). Thus, the unit of observation in the dataset is considered to be individual excretion of nitrogen (N) or phosphorus (P) per unit time ($\mu\text{g N or P excreted per capita per hour}$).

Some studies reported only mean excretion rates (i.e., the average of what we refer to as observations) rather than data from individual animals. If we could not obtain data on individual observations from the authors, these studies were not included in our dataset.

Our dataset includes a total of 10,534 records (lines of data). However, studies vary in whether they report N excretion rate, P excretion rate, and/or excreted N:P. Thus, the data set contains includes 9822, 8245 and 7513 observations for N excretion rate, P excretion rate, and excreted N:P, respectively. Excreted N:P was sometimes reported by the authors (if both rates were measured) but otherwise we calculated it by dividing N excretion rate by P excretion rate and converting this value to molar units.

b. Instrumentation

N and P were usually measured using standard colorimetric techniques, though NH₄-N was sometimes measured by fluorometry.

c. Taxonomy and systematics

The dataset includes both invertebrate and vertebrate animals. Most aquatic invertebrate phyla are represented, as well as numerous fishes and some amphibians and turtles.

4. *Project personnel*

Many individuals were involved in collection of original data, as shown in the data file

“Aquatic_animal_excretion_data.csv.” The data were compiled, standardized, and organized by the two lead authors, M.J. Vanni and P.B. McIntyre.

Class III. Data set status and accessibility

A. Status

1. *Latest update*

27 November 2016

2. *Latest archive data*

27 November 2016

3. *Metadata status*

27 November 2016

4. *Data verification*

Data were checked in several ways by the two first authors (MJV and PBM). For each variable (e.g., body mass, temperature, excretion rates, etc.), we sorted the data, examined distributions, and looked for outliers or suspicious data points. In a very small proportion of cases, P excretion rates were reported as negative values (this occurs when P excretion rates are very low, such that the change in concentration in incubation chambers is within the range of analytical precision, and concentrations may appear to decline due solely to measurement error). These observations were deleted, precluding us from calculating excreted N:P for these observations.

B. Accessibility

1. *Storage location and medium*

The metadata and data files have been submitted to Ecology.

2. *Contact person*

Michael J. Vanni, Department of Biology, Miami University. vannimj@miamioh.edu; 513-529-3192

3. **Copyright restrictions**

Users are free to use and analyze the data. We request that attribution is given to this presentation of the data, and that any changes to the dataset are detailed. When appropriate, additional attribution to the original data collector is also encouraged.

4. **Proprietary restrictions**

- a. Release date: n/a
- b. Citation: n/a
- c. Disclaimer: n/a

5. **Costs**

There are no costs associated with using these data.

Class IV. Data structural descriptors

A. Data set file

1. **Identity**

Aquatic_animal_excretion_data.csv
Aquatic_animal_excretion_variable_descriptions.csv

2. **Size**

Aquatic_animal_excretion_data.csv: 10,534 observations, 3.4 MB
Aquatic_animal_excretion_variable_descriptions.csv: List of 35 variables, 5 KB

3. **Format and storage mode**

The data are contained in a .csv file downloadable from Ecology or Ecological Archives.

Literature cited

Data source references

- Alves, JM, et al. (2010) Stoichiometry of benthic invertebrate nutrient recycling: interspecific variation and the role of body mass. *Aquat. Ecol* 44:421-430.
- Andersen, V (1989). Phosphate excretion rate of *Salpa fusiformis cuvier* (Tunicata, Thaliacea). *Hydrobiologia* 171:91-97.
- Andre ER, Hecky RE, Duthie HC (2003) Nitrogen and phosphorus regeneration by cichlids in the littoral zone of Lake Malawi, Africa. *J. Gt. Lakes Res* 29:190-201.
- Arnott DL, Vanni MJ (1996) Nitrogen and phosphorus recycling by the zebra mussel (*Dreissena polymorpha*) in the western basin of Lake Erie. *Can. J. Fish. Aquat. Sci* 53:646-659.
- Bamstedt U, Tande KS (1985) Respiration and excretion rates of *Calanus glacialis* in arctic waters of the Barents Sea. *Marine Biology* 87:259-266.
- Barlow JP, Bishop JW (1965) Phosphate regeneration by zooplankton in Cayuga Lake. *Limnol. Oceanogr.* 10:R15-R24.
- Bayne BL, Scullard C (1977) Rates of nitrogen excretion by species of *Mytilus* (Bivalvia - Mollusca). *J. Mar. Biol. Assoc. U.K* 57:355-369.
- Benstead JP, et al. (2010) Biotic and abiotic controls on the ecosystem significance of consumer excretion in two contrasting tropical streams. *Freshw. Biol* 55:2047-2061.
- Brabrand Å, Faafeng BA, Nilssen JPM (1990) Relative importance of phosphorus supply to phytoplankton production -

- fish excretion versus external loading. *Can. J. Fish. Aquat. Sci* 47:364-372.
- Burkhardt S, Lehman JT (1994) Prey consumption and predatory effects of an invertebrate predator (*Bythotrephes*, Cladocera, Cercopagidae) based on phosphorus budgets. *Limnol. Oceanogr* 39:1007-1019.
- Capps KA, Flecker AS (2013) Invasive fishes generate biogeochemical hotspots in a nutrient-limited system. *Plos One*, 8, article 354093.
- Christian AD, Crump BG, Berg DJ (2008) Nutrient release and ecological stoichiometry of freshwater mussels (Mollusca : Unionidae) in 2 small, regionally distinct streams. *J. N. Am. Benthol. Soc* 27:440-450.
- Clarke A, Prothero-Thomas E, Whitehouse MJ (1994) Nitrogen-excretion in the antarctic limpet *Nacella concinna* (Strebel, 1908). *J. Molluscan Stud* 60:141-147.
- Conroy JD, et al. (2005) Soluble nitrogen and phosphorus excretion of exotic freshwater mussels (*Dreissena* spp.): potential impacts for nutrient remineralisation in western Lake Erie. *Freshw. Biol* 50:1146-1162.
- Devine JA, Vanni MJ (2002) Spatial and seasonal variation in nutrient excretion by benthic invertebrates in a eutrophic reservoir. *Freshw. Biol* 47:1107-1121.
- Evans-White MA, Lamberti GA (2005) Grazer species effects on epilithon nutrient composition. *Freshw. Biol* 50:1853-1863.
- Follum OA, Gray JS (1987) Nitrogenous excretion by the sediment-living bivalve *Nucula tenuis* from the Oslofjord, Norway. *Mar. Biol* 96:355-358.
- Fukuhara H, Sakamoto M (1987) Enhancement of inorganic nitrogen and phosphate release from lake sediment by tubificid worms and chironomid larvae. *Oikos* 48:312-320.
- Fukuhara H, Yasuda K (1985) Phosphorus excretion by some zoobenthos in a eutrophic freshwater lake and its temperature dependency. *Japan. J. Limnol* 4:287-296.
- Gardner WS, Briones EE, Kaegi EC, Rowe GT (1993) Ammonium excretion by benthic invertebrates and sediment-water nitrogen flux in the Gulf of Mexico near the Mississippi River outflow. *Estuaries* 16:799-808.
- Gido KB (2002) Interspecific comparisons and the potential importance of nutrient excretion by benthic fishes in a large reservoir. *Trans. Am. Fish. Soc* 131:260-270.
- Godinot C, Chadwick NE (2009) Phosphate excretion by anemonefish and uptake by giant sea anemones: demand outstrips supply. *Bull. Mar. Sci* 85:1-9.
- Gorsky G, Palazzoli I, Fenaux R (1987) Influence of temperature-changes on oxygen-uptake and ammonia and phosphate excretion, in relation to body size and weight, in *Oikopleura dioica* (Appendicularia). *Mar. Biol* 94:191-201.
- Gray JS (1985) Nitrogenous excretion by meiofauna from coral reef sediments – Meco 5. *Mar. Biol* 89:31-35.
- Haertel-Borer SS, Allen DM, Dame RF (2004) Fishes and shrimps are significant sources of dissolved inorganic nutrients in intertidal salt marsh creeks. *J. Exp. Mar. Biol. Ecol* 311:79-99.

- Hall RO, Koch BJ, Marshall MC, Taylor BW, Tronstad LM (2007) How body size mediates the role of animals in nutrient cycling in aquatic ecosystems. In: *Body Size: The Structure and Function of Aquatic Ecosystems* (eds Hildrew, A.G., Raffaelli, D.G. & Edmonds-Brown, R.). Cambridge University Press, Cambridge, U.K., pp 286-305.
- Hall RO, Tank JL, Dybdahl MF (2003) Exotic snails dominate nitrogen and carbon cycling in a highly productive stream. *Front. Ecol. Environ* 1:407-411.
- Henry R, Santos CM (2008) The importance of excretion by *Chironomus* larvae on the internal loads of nitrogen and phosphorus in a small eutrophic urban reservoir. *Braz. J. Biol* 68:349-357.
- Higgins KA, Vanni MJ, Gonzalez MJ (2006) Detritivory and the stoichiometry of nutrient cycling by a dominant fish species in lakes of varying productivity. *Oikos* 114:419-430.
- Ikeda database (<http://eprints.lib.hokudai.ac.jp/dspace/handle/2115/33838>). Data were taken directly from this database, which contains data from the following sources:
- Ikeda T (1985) Metabolic rates of epipelagic marine zooplankton as a function of body-mass and temperature. *Mar. Biol* 85:1-11.
- Ikeda T, Kanno Y, Ozaki K, Shinada A (2001a) Metabolic rates of epipelagic marine copepods as a function of body mass and temperature. *Mar. Biol* 139:587-596.
- Ikeda T, Kanno Y, Ozaki K, Shinada A (2001b) Metabolic rates of epipelagic marine copepods as a function of body mass and temperature (vol 139, pg 587, 2001). *Mar Biol* 139:1020-1020.
- Ikeda T, Sano F, Yamaguchi A (2007) Respiration in marine pelagic copepods: a global-bathymetric model. *Mar. Ecol. Prog. Ser* 339:215-219.
- Ikeda T, Sano F, Yamaguchi A, Matsuishi T (2006) Metabolism of mesopelagic and bathypelagic copepods in the western North Pacific Ocean. *Mar. Ecol. Prog. Ser* 322:199-211.
- Ikeda T, Torres JJ, Hernandez-Leon S, Geiger SP (2000) Metabolism. In: *ICES Zooplankton Methodology Manual* (eds. Harris, R.H., Weibe, P., Lenz, J., Skjoldal H.R. & Huntley M.). Academic Press, p. 455-532.
- Omori M, Ikeda T (1984) *Methods in Marine Zooplankton Ecology*. John Wiley and Sons, California, USA.
- James MR, Weatherhead MA, Ross AH (2001) Size-specific clearance, excretion, and respiration rates, and phytoplankton selectivity for the mussel *Perna canaliculus* at low levels of natural food. *N. Z. J. Mar. Freshw. Res* 35:73-86.
- James WF, et al. (2000) Filtration and excretion by zebra mussels: Implications for water quality impacts in Lake Pepin, upper Mississippi River. *J. Freshw. Ecol.* 15:429-437.
- Jansen HM, Strand O, Verdegem M, Smaal A (2012) Accumulation, release and turnover of nutrients (C-N-P-Si) by the blue mussel *Mytilus edulis* under oligotrophic conditions. *J. Exp. Mar. Biol. Ecol.* 416:185-195.
- Ji, L, et al. (2011) Phosphorus flux by macrobenthic invertebrates in a shallow eutrophic lake Donghu: spatial change.

Knowl. and Manag. Aquat. Ecosyst. 402 (11).

- Johnson CR, Luecke C, Whalen SC, Evans MA (2010) Direct and indirect effects of fish on pelagic nitrogen and phosphorus availability in oligotrophic Arctic Alaskan lakes. *Can. J. Fish. Aquat. Sci.* 67:1635-1648.
- Kiibus M, Kautsky N (1996) Respiration, nutrient excretion and filtration rate of tropical freshwater mussels and their contribution to production and energy flow in Lake Kariba, Zimbabwe. *Hydrobiologia* 331:25-32.
- Kouassi E, Pagano M, Saint-Jean L, Sorbe JC (2006) Diel vertical migrations and feeding behavior of the mysid *Rhopalophthalmus africana* (Crustacea : Mysidacea) in a tropical lagoon (Ebrie, Coted'Ivoire). *Estuar. Coast. Shelf Sci.* 67:355-368.
- Lamarra VAJ (1975) Digestive activities of carp as a major contributor to the nutrient loading of lakes. *Verh. Int. Verein. Limnol.* 19:2461-68.
- Lauritsen DD, Mozley SC (1989) Nutrient excretion by the asiatic clam *Corbicula fluminea*. *J. N. Am. Benthol. Soc.* 8:134-139.
- Martin S, et al. (2006) Respiration, calcification, and excretion of the invasive slipper limpet, *Crepidula fornicata* L.: Implications for carbon, carbonate, and nitrogen fluxes in affected areas. *Limnol. Oceanogr.* 5:1996-2007.
- Mather ME, Vanni MJ, Wissing TE, Davis SA, Schaus MH (1995) Regeneration of nitrogen and phosphorus by bluegill and gizzard shad: Effect of feeding history. *Can. J. Fish. Aquat. Sci.* 52:2327-2338.
- McIntyre PB, et al. (2008) Fish distributions and nutrient cycling in streams: Can fish create biogeochemical hotspots? *Ecology* 89:2335-2346.
- McIntyre PB, Jones LE, Flecker AS, Vanni MJ (2007) Fish extinctions alter nutrient recycling in tropical freshwaters. *Proc. Natl. Acad. Sci. U.S.A.* 104:4461-4466.
- McIntyre PB (2006) The importance of fishes in nutrient cycling in tropical freshwaters: Species, community, and ecosystem perspectives. Dissertation. Cornell University, Ithaca, NY, USA.
- McManamay RA, Webster JR, Valett HM, Dolloff CA (2011) Does diet influence consumer nutrient cycling? Macroinvertebrate and fish excretion in streams. *J. N. Am. Benthol. Soc.* 30:84-102.
- Mellina E, Rasmussen JB, Mills EL (1995) Impact of zebra mussel (*Dreissena polymorpha*) on phosphorus cycling and chlorophyll in lakes. *Can. J. Fish. Aquat. Sci.* 52:2553-2573.
- Meyer JL, Schultz ET (1985) Migrating haemulid fishes as a source of nutrients and organic-matter on coral reefs. *Limnol. Oceanogr.* 30:146-156.
- Milanovich JR, Hopton ME (2014) Stoichiometry of a semi-aquatic plethodontid salamander: Intraspecific variation due to location, size and diet. *Integr. Zool.* 9:613-622.
- Milanovich JR, et al. unpublished data.
- Morgan DKJ, Hicks BJ (2013) A metabolic theory of ecology applied to temperature and mass dependence of N and P

excretion by common carp. *Hydrobiologia* 705:135-145.

- Moslemi JM, Snider SB, MacNeill K, Gilliam JF, Flecker AS (2012) Impacts of an Invasive Snail (*Tarebia granifera*) on Nutrient Cycling in Tropical Streams: The Role of Riparian Deforestation in Trinidad, West Indies. *Plos One*, 7, article 38806.
- Munshaw RG, Palen WJ, Courcelles DM, Finlay JC (2013) Predator-driven nutrient recycling in California stream ecosystems. *Plos One*, 8, article e58542
- Naddafi R, Pettersson K, Eklov P (2008) Effects of the zebra mussel, an exotic freshwater species, on seston stoichiometry. *Limnol. Oceanogr.* 53:1973-1987.
- Paffenhofer GA, Gardner WS (1984) Ammonium release by juveniles and adult females of the sub-tropical marine copepod *Eucalanus pileatus*. *J. Plankton Res.* 6:505-513.
- Pilati A, Vanni MJ (2007) Ontogeny, diet shifts, and nutrient stoichiometry in fish. *Oikos*, 116:1663-1674.
- Post DM, Walters AW (2009) Nutrient excretion rates of anadromous alewives during their spawning migration. *Trans. Am. Fish. Soc.* 138, 264-268.
- Prosch RM, McLachlan A (1984) The regeneration of surf-zone nutrients by the sand mussel, *Donax serra* Roding. *J. Exp. Mar. Biol. Ecol.* 80, 221-233.
- Roopin M, Henry RP, Chadwick NE (2008) Nutrient transfer in a marine mutualism: patterns of ammonia excretion by anemonefish and uptake by giant sea anemones. *Mar Biol* 154:547-556.
- Rugenski AT (2013) Influence of disease-driven amphibian declines on ecosystem structure and function in Panamanian headwater streams. Dissertation. Southern Illinois University, Carbondale, IL, U.S.A.
- Schaus MH, et al. (2010) Impact of the removal of gizzard shad (*Dorosoma cepedianum*) on nutrient cycles in Lake Apopka, Florida. *Freshw. Biol.* 55:2401-2413.
- Schaus MH, et al. (2013) Effect of a size-selective biomanipulation on nutrient release by gizzard shad in Florida (USA) lakes. *Knowl. Manag. Aquat. Ecosyst.* 411:13.
- Schaus MH, et al. (1997) Nitrogen and phosphorus excretion by detritivorous gizzard shad in a reservoir ecosystem. *Limnol. Oceanogr.* 42:1386-1397.
- Sereda JM, Hudson JJ, Taylor WD, Demers E (2008) Fish as sources and sinks of nutrients in lakes. *Freshw. Biol.* 53:278-289.
- Shimauchi H, Uye S (2007) Excretion and respiration rates of the scyphomedusa *Aurelia aurita* from the Inland Sea of Japan. *J. Oceanogr.* 63:27-34.
- Shostell J, Bukaveckas PA (2004) Seasonal and interannual variation in nutrient fluxes from tributary inputs, consumer recycling and algal growth in a eutrophic river impoundment. *Aquat. Ecol.* 38:359-373.

- Small GE, Pringle CM, Pyron M, Duff JH (2011) Role of the fish *Astyanax aeneus* (Characidae) as a keystone nutrient recycler in low-nutrient Neotropical streams. *Ecology* 92:386-397.
- Solomon CT, Olden JD, Johnson PTJ, Dillon, RT Jr, Vander Zanden MJ (2010) Distribution and community-level effects of the Chinese mystery snail (*Bellamya chinensis*) in northern Wisconsin lakes. *Biol. Invasions* 12:1591-1605.
- Srna RF, Baggaley A (1976) Rate of excretion of ammonia by hard clam *Mercenaria-mercenaria* and American oyster *Crassostrea-virginica*. *Mar. Biol.* 36:251-258.
- Sterrett SC, Maerz JC, RA Katz (2015) What can turtles teach us about the theory of ecological stoichiometry? *Freshw. Biol.* 60:443-455.
- Tarvainen M, Ventela AM, Helminen H, Sarvala J. (2005) Nutrient release and resuspension generated by ruffe (*Gymnocephalus cernuus*) and chironomids. *Freshw. Biol.* 50:447-458.
- Tatrai I (1982) Oxygen-consumption and ammonia excretion of herbivorous chironomid larvae in Lake Balaton. *Hydrobiologia* 96:129-135.
- Taylor JM, Back JA, Valenti TW, King RS (2012) Fish-mediated nutrient cycling and benthic microbial processes: can consumers influence stream nutrient cycling at multiple spatial scales? *Freshw. Sci.* 31:928-944.
- Torres LE, Vanni MJ (2007) Stoichiometry of nutrient excretion by fish: interspecific variation in a hypereutrophic lake. *Oikos* 116:259-270.
- Turner CB (2010) Influence of zebra (*Dreissena polymorpha*) and quagga (*Dreissena rostriformis*) mussel invasions on benthic nutrient and oxygen dynamics. *Can. J. Fish. Aquat. Sci.* 67:1899-1908.
- Urabe J (1993) N-cycling and P-cycling coupled by grazers activities - food quality and nutrient release by zooplankton. *Ecology* 74:2337-2350.
- Vanderploeg HA, Laird GA, Liebig JR, Gardner WS (1986) Ammonium release by zooplankton in suspensions of heat-killed algae and an evaluation of the flow-cell method. *J. Plankton Res.* 8:341-352.
- Vaughn CC, Gido KB, Spooner DE (2004) Ecosystem processes performed by unionid mussels in stream mesocosms: species roles and effects of abundance. *Hydrobiologia*, 527:35-47.
- Vanni MJ, Flecker AS, Hood JM, Headworth JL (2002) Stoichiometry of nutrient recycling by vertebrates in a tropical stream: linking species identity and ecosystem processes. *Ecol. Lett.* 5:285-293.
- Villegger S, Ferraton F, Mouillot D, de Wit R (2012a) Nutrient recycling by coastal macrofauna: intra- versus interspecific differences. *Mar. Ecol. Prog. Ser.* 452, 297-303.
- Villegger S, Grenouillet G, Suc V, Brosse S (2012b) Intra- and interspecific differences in nutrient recycling by European freshwater fish. *Freshw. Biol.* 57:2330-2341.
- Whiles MR, Hury AD, Taylor BW, Reeve JD (2009) Influence of handling stress and fasting on estimates of ammonium excretion by tadpoles and fish: recommendations for designing excretion experiments. *Limnol. Oceanogr. Meth.*

- Wilhelm, F.M., Hudson, J.J. & Schindler, D.W. (1999). Contribution of *Gammarus lacustris* to phosphorus recycling in a fishless alpine lake. *Can. J. Fish. Aquat. Sci.*, 56, 1679-1686.
- Wilson HF, Xenopoulos MA (2011) Nutrient recycling by fish in streams along a gradient of agricultural land use. *Glob. Change Biol.* 17:130-139.
- Zimmer KD, Herwig BR, Laurich LM (2006) Nutrient excretion by fish in wetland ecosystems and its potential to support algal production. *Limnol. Oceanogr.* 51:197-207.

Additional references cited

- Allen, A. P., and J. P. Gillooly. 2009. Towards and integration of ecological stoichiometry and the metabolic theory of ecology to better understand nutrient cycling. *Ecology Letters* 12:369-384.
- Andersson, G., W. Graneli, and J. Stenson. 1988. The influence of animals on phosphorus cycling in lake ecosystems. *Hydrobiologia* 170:267-284.
- Brown, J. H., J. F. Gillooly, A. P. Allen, V. M. Savage, and G. B. West. 2004. Toward a metabolic theory of ecology. *Ecology* 85:1771-1789.
- Clarke, A. 2004. Is there a universal temperature dependence of metabolism? *Functional Ecology* 18:252-256.
- Elser, J. J., D. R. Dobberfuhl, N. A. MacKay, and J. H. Schampel. 1996. Organism size, life history and N:P stoichiometry. *BioScience* 46:674-684.
- Gillooly, J. F., J. H. Brown, G. B. West, V. M. Savage, and E. L. Charnov. 2001. Effects of size and temperature on metabolic rate. *Science* 293:2248-2251.
- Glazier, D. S. 2010. A unifying explanation for diverse metabolic scaling in animals and plants. *Biological Reviews* 85:111-138
- Hall R. O., B. J. Koch, M. C. Marshall, B. W. Taylor, and L. M. Tronstad. 2007. How body size mediates the role of animals in nutrient cycling in aquatic ecosystems. Pages 286-305 in A. G. Hildrew, D. G. Raffaelli, and R. Edmonds-Brown, editors. *Body size: The structure and function of aquatic ecosystems*. Cambridge Univ. Press, Cambridge.
- Hurlbert, S. H. 1984. Pseudoreplication and the design of ecological field experiments. *Ecological Monographs* 54:187-211.
- Isaac, N. J. B., and C. Carbone. 2010. Why are metabolic scaling exponents so controversial? Quantifying variance and testing hypotheses. *Ecology Letters* 13:728-735.
- McIntyre, P. B., A. S. Flecker, M. J. Vanni, J. M. Hood, B. W. Taylor, and S. A. Thomas. 2008. Fish distributions and nutrient cycling in streams: Can fish create biogeochemical hotspots? *Ecology* 89:2335-2346.
- McNaughton, S. J., F. F. Banyikwa, and M. M. McNaughton. 1997. Promotion of the cycling of diet-enhancing nutrients by African grazers. *Science* 278:1798-1800.

- Michener, W.K., J. W. Brunt, J. J. Helly, T. B. Kirchner, and S. S. Stafford. 1997. Nongeospatial metadata for the ecological sciences. *Ecological Applications* 7:330-342.
- Moulton, O. M., M. A. Altabet, J. M Beman, L. A. Deegan, J. Lloret, M. K. Lyons, J. A. Nelson, and C. A. Pfister. 2016. Microbial associations with macrobiota in coastal ecosystems: patterns and implications for nitrogen cycling. *Frontiers in Ecology and the Environment* 14:200-208.
- Sereda, J. M., and J. J. Hudson. 2011. Empirical models for predicting the excretion of nutrients (N and P) by aquatic metazoans: taxonomic differences in rates and element ratios. *Freshwater Biology* 56:250-263.
- Sterner, R. W. 1990. The ratio of nitrogen to phosphorus resupplied by herbivores - zooplankton and the algal competitive arena. *American Naturalist* 136:209-229.
- Sterner R. W. and J. J. Elser. 2002. *Ecological stoichiometry: The Biology of elements from molecules to the biosphere.* Princeton Univ Press, Princeton.
- Taylor, J. M., M. J. Vanni, and A. S. Flecker. 2015. Top-down and bottom-up interactions in freshwater ecosystems: Emerging complexities. *In: Hanley, T.C., and K.J. La Pierre, eds: Trophic Ecology: Bottom-Up and Top-Down Interactions across Aquatic and Terrestrial Systems.* Cambridge University Press.
- Vanni, M. J. 2002. Nutrient cycling by animals in freshwater ecosystems. *Annual Review of Ecology and Systematics* 33:341-370.
- Vanni, M. J., A. S. Flecker, J. M. Hood, and J. L. Headworth. 2002. Stoichiometry of nutrient recycling by vertebrates in a tropical stream: linking species identity and ecosystem processes. *Ecology Letters* 5:285-293.
- Vanni, M. J. and P. B. McIntyre. 2016. Predicting nutrient excretion rates of aquatic animals with metabolic ecology and ecological stoichiometry: a global synthesis. *Ecology* 97:3460-3471. DOI: 10.1002/ecy.1582
- West, G. B., J. H. Brown, and B. J. Enquist. 1997. A general model for the origin of allometric scaling laws in biology. *Science* 276:122-126.
- White, C. R. and R. S. Seymour. 2005. Allometric scaling of mammalian metabolism. *Journal of Experimental Biology* 208:1611-1619.
- Wood, J.D., D. Elliott, G. Garman, D. Hopler, W. Lee, S. McIninch, A.J. Porter, and P.A. Bukaveckas. 2016. Autochthony, allochthony and the role of consumers in influencing the sensitivity of aquatic systems to nutrient enrichment. *Food Webs* 7:1-12