Effect of candesartan and metoprolol on myocardial tissue composition during anthracycline treatment: the PRADA trial

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Contemporary anticancer treatment with anthracyclines has significantly improved survival
 in early breast cancer. However, the beneficial effect of anthracyclines must be balanced
 against their well-known cardiotoxicity,¹ which is related to cumulative and peak doses and
 considered irreversible.^{2,3} Histological changes include myofibrillar loss, myocyte death due
 to both apoptosis and necrosis, and interstitial oedema and fibrosis.⁴

Late gadolinium enhancement (LGE) imaging by cardiovascular magnetic resonance (CMR)
fails in depicting diffuse fibrosis,⁵ and LGE is a rare finding in anthracycline cardiotoxicity.^{6,7}
Extracellular volume (ECV) fraction is a relative measure that expresses the relationship
between myocardial ECV and cellular volume, and provides valuable information about
diffuse myocardial changes .⁸ By relating these measurements to mass, the total extracellular
and total cellular volume of the myocardium can be calculated, and this may be especially
useful in longitudinal studies.⁹

13 We have recently shown that adjuvant therapy for early breast cancer with anthracyclines, 14 radiotherapy and/or the monoclonal antibody trastuzumab is associated with a small, but significant decline in left ventricular ejection fraction (LVEF), which is alleviated by the 15 angiotensin receptor blocker candesartan but not by the beta blocker metoprolol.¹⁰ In the 16 17 current longitudinal substudy, we tested the hypotheses that anthracycline therapy for early 18 breast cancer is associated with a dose-dependent increase in myocardial ECV fraction and 19 total ECV as well as a reduction in total myocardial cellular volume. Moreover, we 20 hypothesized that these changes might be prevented by the concomitant administration of 21 candesartan and/or metoprolol.

22

1 Methods

2 Study design and participants

3 Between September 2011 and September 2014, 120 patients scheduled for anthracycline-4 containing adjuvant therapy after surgery for early breast cancer at Akershus University 5 Hospital, Norway, were prospectively included. The Regional Ethics Committee approved 6 the study, and the trial was registered in the ClinicalTrials.gov registry (NCT01434134) prior 7 to study initiation. All participants provided written, informed consent prior to enrolment. 8 The main eligibility criteria were no serious concomitant illness, no prior cardiovascular 9 disease, and no indication or contraindications for the study drugs. Details on study rationale 10 and design, eligibility criteria, patient screening and randomization have been reported previously.^{10,11} The enrolled patients were randomized by a 2 x 2 factorial design to receive 11 12 one of the following treatment combinations: candesartan cilexetil 32 mg q.d. and metoprolol 13 succinate100 mg q.d.; candesartan cilexetil 32 mg q.d. and placebo q.d.; metoprolol succinate 14 100 mg q.d. and placebo q.d.; or placebo and placebo q.d. We registered compliance by 15 counting residual tablets on every second visit, and patients kept a diary to register intake of 16 tablets. Patients were examined with CMR at baseline and after completion of the final cycle 17 of anthracycline-containing therapy with 5-fluorouracil, epirubicin and cyclophosphamide 18 (FEC), before commencement of additional treatment with radiotherapy, taxanes or 19 trastuzumab.

Twenty-four patients underwent baseline CMR before ECV fraction measurements were available in March 2012, and six patients were not examined by CMR at completion of anthracycline therapyTwenty-one patients did not have ECV measurements at baseline or at completion of anthracyclines due to missing haematocrit, artefacts, unenhanced CMR or inactive mapping module (Supplemental Table S1). Accordingly, 69 patients had valid ECV fraction measurements at both baseline and the end of anthracycline treatment and constitute

the study population of the current report. Table 1 shows their baseline characteristics, and
 Supplemental Table S2 shows baseline characteristics of included vs. excluded patients.

3 CMR protocol

4 All CMR examinations were performed on the same 1.5-T MRI scanner (Achieva; Philips 5 Medical Systems, Best, The Netherlands), using a five-element phased-array cardiac coil. 6 Balanced Steady-State-Free-Precession sequences in contiguous, eight mm thick short-axis 7 slices covering the entire ventricles were used to quantify LVEF and LV mass. LGE images 8 were typically acquired with a two-dimensional inversion recovery turbo field echo sequence 9 in short axis covering the ventricles, and phase-sensitive three-dimensional inversion 10 recovery turbo field echo sequences in four chamber and left two chamber axis, starting 10 11 min after intravenous injection of 0.2 mmol/kg Gadolinium-DOTA (Dotarem®, Guerbet, 12 France). Mid-ventricular, short axis images for T1 mapping were acquired before and 15 13 minutes after contrast administration, with single breath-hold, ECG gated, balanced steadystate free precession, modified Look-Locker Imaging (MOLLI)¹² with the following 14 15 acquisition parameters: TR/TE/flip angle were 2.6 ms/1.03ms/35° and acquired and reconstructed voxel size were 1.7x2.1x10mm³ and 1.1x1.1x10mm³, respectively. The 16 17 sampling scheme was 3(2)3(2)5. We used a heart rate-adapted trigger delay. Native and post-18 contrast T1 maps were generated by dedicated, commercially available software (cmr42, 19 version 5.2.0, Circle Cardiovascular Inc., Calgary, Canada). Additional information on scan 20 parameters is provided in the online supplement.

21

22 CMR analysis

All image analyses were performed on cmr42 by a single, board-certified radiologist (SLH)
blinded for treatment allocation and study order. End-diastolic and end-systolic epicardial
and endocardial contours were traced, permitting calculation of LV volumes, EF and mass.

1	Trabeculations and papillary muscles were included in the LV volume and excluded from LV
2	mass estimation. Analyses were performed according to the Society for Cardiovascular
3	Magnetic Resonance guidelines. ¹³ Myocardial native and post-contrast T1 values were
4	obtained by conservatively tracing endo- and epicardial contours on each T1 map to avoid
5	partial volume effects (Figure 1), excluding areas of LGE. Each map was divided into six
6	segments according to the American Heart Association segment model, ¹⁴ providing average
7	LV T1 values for each segment as well as each slice. Source images and goodness-of-fit
8	parametric maps (R^2 maps) were inspected for motion and off-resonance artefacts. The
9	quality of each segment of the T1 maps was assessed, and segments with off resonance
10	artefacts or significant motion artefacts were rejected. Myocardial T1 was calculated as the
11	mean value of segments with matching valid native and post-contrast measurements. Blood
12	T1 was obtained by drawing a region of interest in the LV blood pool, avoiding the papillary
13	muscles. ECV fraction (%) was calculated as
14	(100%-haematocrit(%)) × (Δ R1myocardium/ Δ R1blood), where R1 = 1/T1, ^{15,8}
15	total ECV (ml) was calculated as
16	ECV fraction \times myocardial volume (ml) ⁹
17	and the total cellular volume (ml) as
18	(100%-ECV fraction) \times myocardial volume (ml),
19	where myocardial volume is LV mass (g), divided by the myocardial specific density 1.05
20	g/ml. A sample of 15 examinations was randomly selected for evaluation of intra-observer
21	variability. The same sample was also evaluated by another reader (FvKB) for assessment of
22	inter-observer variability (online supplement).

1 Statistical analysis

2 All statistical analyses were performed according to the intention-to-treat principle using

3 IBM SPSS Statistics version 22.

The Shapiro-Wilks test was used to test normality. Normally distributed data are presented as
mean ± standard deviation, non-normally distributed data as median (25-75 percentiles).
Differences between means, medians or categorical variables were assessed by Student's ttest, the Wilcoxon signed-rank test, the Mann-Whitney U test or Fischer's exact test, as
appropriate. Power calculations for the PRADA study were based on the primary endpoint,
i.e. LVEF. We did not correct for multiple comparisons. All tests were two-sided, and p <

10 0.05 was considered statistically significant.

11

12 **Results**

13 **Baseline characteristics**

14 Baseline characteristics are listed in Table 1. Adjuvant therapy was in accordance with the 15 national guidelines for adjuvant breast cancer treatment in Norway at the time of inclusion. 16 Eleven patients received four cycles containing 100 mg/m² epirubicin (as part of the FEC-100 17 regimen, cumulative doxorubicin equivalent dose 268 mg/m²), representing both the highest 18 peak and cumulative anthracycline dose in this study, while 58 patients received four or six 19 cycles containing 60 mg/m² epirubicin (median cumulative doxorubicin equivalent dose of 20 161 (161,241 mg/m²). Two patients had small areas of LGE with a non-ischemic pattern 21 (mean percentage of LV myocardium $3.0 \pm 1.0\%$).

22

23 Changes during anthracycline therapy

24 Median time from baseline to end of anthracycline CMR examination was 13.0 weeks (95 %

25 confidence interval [CI] 13.7,15.4). During anthracycline therapy, LVEF decreased 3.1%

1 (95% CI -5.2, -1.0, p=0.006) in the placebo-placebo group. The decline in LVEF was most 2 marked in patients receiving the highest anthracycline doses, and in patients who did not 3 receive candesartan (Table 2 and 3). No patient developed symptomatic heart failure. 4 The ECV fraction of the total study cohort increased from 27.5±2.7% to 28.6±2.9% (mean 5 change 1.2% [95% CI 0.4, 1.9]; p= 0.002). The ECV fraction and total ECV increased 6 significantly more in patients receiving the highest anthracycline doses than in patients who 7 received lower doses: 3.4% (95% CI 1.2, 5.5) vs. 0.7% (95% CI 0.0, 1.5); p=0.006 and 1.9 ml 8 (95% CI 0.4, 3.5) vs. 0.1 ml (95% CI -0.6, 0.8); p=0.040. 9 There was no impact of candesartan on change in ECV fraction. However, in patients 10 receiving candesartan total cellular volume decreased significantly more than in those not 11 receiving candesartan (-3.5 ml [95% CI -4.7, -2.2] vs. -0.6 ml [95% CI -2.1, 0.9]; p=0.003). 12 Details of changes from baseline to the end of anthracycline therapy are shown in Table 2 13 and 3 and in Figure 2. 14 There was no difference in change in LVEF, ECV fraction, total ECV or total cellular volume 15 between patients who received or did not receive metoprolol (online supplementary Table 16 S3). We did not observe any new areas of LGE, nor did the LGE observed at baseline 17 increase (mean change 0.0% [95% CI -0.7, 0.7]; p=0.968). There was no apparent effect of 18 coincidental statin use on any measures (data not shown). One patient assigned to 19 candesartan and two patients assigned to metoprolol were not compliant to the intervention. 20 The per protocol analysis did not differ significantly from the intention to treat analysis, 21 online supplement Table S4 and S5. 22

1 **Discussion**

The new and important findings of this study are that adjuvant anthracycline therapy for early breast cancer is associated with dose-dependent subclinical structural myocardial changes detectable by serial CMR that are modified by neuroendocrine blockade . Accordingly, treatment with higher doses of the anthracycline epirubicin was associated with significantly greater increase in ECV fraction and in total ECV than lower doses. Moreover, patients who received candesartan experienced a significant decrease in myocardial total cellular volume.
(Figure 3) Finally, none of the patients developed focal fibrosis.

9

10 ECV fraction and total ECV

Previous longitudinal studies have shown that traditional CMR markers such as LGE ^{6,7,16} 11 and T2 ratio¹⁷ do not reliably detect cardiotoxicity caused by chemotherapy. Our findings 12 13 suggest that changes in ECV fraction and total ECV may be more sensitive markers. ECV 14 fraction is elevated in a number of cardiomyopathies and has been shown to correlate strongly with histologically determined diffuse interstitial fibrosis.^{18,19} However, as it is a 15 16 relative measure, increasing ECV fraction in longitudinal studies may be caused by either expansion of the extracellular matrix due to oedema or fibrosis, or by reduction of myocyte 17 18 volume. Therefore, the calculation of the total ECV and cellular volumes by relating ECV 19 fraction to mass is especially useful in longitudinal studies. A study of 23 hypertensive 20 patients showed no change in ECV fraction from baseline to six months after renal 21 denervation, but a significant decrease in LV mass and total ECV suggesting that the 22 observed LV mass reduction was due to reversion of both myocyte hypertrophy and interstitial myocardial fibrosis.²⁰ Another study demonstrated that LV hypertrophy regression 23 24 six months post aortic valve replacement in severe aortic stenosis was due to cell volume reduction rather than fibrosis resolution.²¹ However, sparse data concerning the effect of 25

1 adjuvant anti-cancer therapy are available. Two small, cross sectional studies of patients with 2 a history of anthracycline treatment demonstrated higher ECV fraction values compared to that observed in healthy controls²² and that increased ECV fraction several years after anti-3 cancer therapy correlated with higher cumulative anthracycline doses.⁶ Both studies were 4 5 conducted in heterogeneous patient populations with various cancer entities and several years 6 after anthracycline exposure, and did not examine acute changes. A recent longitudinal study 7 showed that patients receiving anthracyclines experienced an increase in ECV fraction of two 8 percent points from baseline to three months, whereas non-anthracycline regimens were not associated with increased ECV fraction.²³ None of these studies report on total extracellular 9 10 or cellular volumes, and whether reduced cellular volume contributed to the increase in ECV 11 fraction is not known. Accordingly, our prospective study is the first to investigate 12 longitudinal changes in ECV fraction and total ECV during anthracycline treatment in a 13 homogenous patient population.

14

15 Anthracycline therapy

Anthracycline cardiotoxicity is dose-dependent,²⁴ and we observed that in patients who 16 received the highest cumulative and peak doses of anthracycline, ECV fraction increased by 17 3.4 percent points which is comparable to increases of 2 to 4 percent points reported in 18 myocarditis.^{25,26} Whether this expansion was caused by oedema or fibrosis, cannot be 19 20 determined by the current data, but a recent study demonstrated oedema by CMR and 21 histopathology five weeks after the initiation of high dose anthracycline treatment in mice, followed by fibrosis at 10 weeks.²⁷ Increased native T1 has been demonstrated both in 22 23 myocardial oedema and diffuse fibrosis, and in a recent report of 56 patients with various 24 cancers, anthracycline treatment was associated with a small but statistically significant increase in native T1 from baseline to three months after initiation of anthracycline therapy.²³ 25

1 We did not observe a significant change in native T1. One possible explanation is that native 2 T1 measures a composite signal from myocytes and extracellular space, and the different 3 effect of anthracyclines and candesartan on these compartments may attenuate signal 4 differences. Also, numeric differences in diffuse fibrosis may be small and a wide overlap in native T1 between patients groups, especially at 1.5 T, may limit statistical power.²⁸ In line 5 with previous studies^{7,29} decline in systolic function was generally small, indicating that 6 7 clinical heart dysfunction from treatment with contemporary anthracycline doses is unusual 8 in the short term. Long-term follow-up of the study patients will be required to determine 9 whether these early changes in total ECV will predict later decline in LV function, and 10 whether the total ECV over time will increase as a marker of diffuse fibrosis in late 11 anthracycline cardiotoxicity.

12 **Candesartan treatment**

13 Anthracyclines are known to cause myofibrillar loss and cell death. A cross-sectional CMR study has shown an inverse association between anthracycline dose and LV mass in patients 14 with established anthracycline-cardiotoxicity,¹⁶ and a longitudinal echocardiographic study of 15 115 paediatric patients treated with median 352 mg/m^2 doxorubicin showed progressive 16 reduction of LV mass as assessed by M-mode echocardiography after end of therapy.³⁰ The 17 18 renin–angiotensin–aldosterone system is thought to be an important mediator in the pathogenesis of LV remodelling after anthracycline cardiotoxicity,³¹ and we found that 19 patients who received candesartan experienced less decline in systolic function than patients 20 21 who did not receive candesartan. Contrary to our hypothesis, however, candesartan, but not 22 anthracycline treatment was associated with a decline in cellular volume. Angiotensin II directly stimulates protein synthesis in myocytes,³² causes myocyte hypertrophy³³ and 23 induces increase in LV mass independent of pressure overload.³⁴ Inhibition of these effects 24 by candesartan may contribute to the observed change in total cellular volume, and our 25

1 findings are in line with echocardiographic studies showing that candesartan reduces LV mass and attenuate myocardial remodelling.^{35,36} As candesartan treatment was associated 2 3 with preserved LV systolic function and no significant increase in total ECV during 4 anthracycline therapy, the reduction in LV mass observed in the candesartan group may be 5 related to attenuation of the growth promoting effects of angiotensin II on cardiomyocytes 6 rather than cardiomyocyte death and replacement fibrosis. The current results demonstrate 7 that inhibition of the renin-angiotensin-aldosterone system in the setting of anthracycline 8 therapy has complex actions on cardiomyocyte structure and function. Also, they underscore 9 the important point that an increasing ECV fraction in longitudinal studies does not 10 necessarily equate expansion of the total extracellular space, and that changes in LV cellular 11 volumes must be taken into account. The recent ESC position paper on cancer treatments and 12 cardiovascular toxicity propose CMR mapping techniques and ECV fraction measurements as a future potential tool for detection of diffuse myocardial fibrosis in cardiotoxicity.³⁷ Our 13 14 study indicates that CMR measurements of the myocardial tissue composition may be useful 15 in longitudinal interventional studies aiming to attenuate remodelling. The long-term clinical 16 implication of the effects of candesartan, with relative preservation of LV systolic function 17 and a modest reduction of cellular volume, remains to be determined.

18

19 Strengths and Limitations

Strengths of our study include the serial CMR evaluation in a homogenous, previously
healthy breast cancer patient population and the single scanner design that minimizes
variability. Although modestly sized, the current analysis encompasses more patients than
previous studies analysing the impact of anthracycline treatment on ECV fraction and is also
the first to report longitudinal changes in total ECV and cellular volume. Some limitations of
our study merit comments. Not all of the 120 patients included in the intervention study had

ECV fraction measurements, but the patients who were eligible for the current study showed
no marked differences from those who were not. Also, we did not explore ECV
heterogeneity, but there were no new areas of focal fibrosis by LGE. Finally, the duration of
follow-up in the current report did not extend beyond the end of anthracycline therapy.

5

6 Conclusions

7 In this longitudinal CMR study, we show that higher doses of the anthracycline epirubicin are

8 associated with greater increase in ECV fraction and total ECV, indices of oedema and

9 diffuse myocardial fibrosis. Treatment with candesartan, which alleviated a reduction in LV

10 systolic function, is associated with a greater decline in total cellular volume than no

11 candesartan treatment while metoprolol did not affect myocardial composition.

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9

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1 **References**

2 1. Ewer MS, Ewer SM. Cardiotoxicity of anticancer treatments. Nat Rev Cardiol. 3 2015 Sep;12(9):547-58. 4 2. Iarussi D, Indolfi P, Casale F, Martino V, Di Tullio MT, Calabro R. Anthracycline-5 induced cardiotoxicity in children with cancer: strategies for prevention and 6 management. *Paediatr Drugs*. 2005;**7**(2):67-76. 7 Ewer MS, Lippman SM. Type II chemotherapy-related cardiac dysfunction: time 3. 8 to recognize a new entity. *J Clin Oncol*. 2005 May 1;23(13):2900-2. 9 4. Butany J, Ahn E, Luk A. Drug-related cardiac pathology. J Clin Pathol. 2009 10 Dec;62(12):1074-84. 11 Mewton N, Liu CY, Croisille P, Bluemke D, Lima JA. Assessment of myocardial 5. 12 fibrosis with cardiovascular magnetic resonance. J Am Coll Cardiol. 2011 Feb 13 22;57(8):891-903. 14 Tham EB, Haykowsky MJ, Chow K, Spavor M, Kaneko S, Khoo NS, et al. Diffuse 6. 15 myocardial fibrosis by T1-mapping in children with subclinical anthracycline 16 cardiotoxicity: relationship to exercise capacity, cumulative dose and remodeling. J 17 Cardiovasc Magn Reson. 2013;15:48. 18 Drafts BC, Twomley KM, D'Agostino R, Lawrence J, Avis N, Ellis LR, et al. Low to 7. 19 Moderate Dose Anthracycline-Based Chemotherapy Is Associated With Early 20 Noninvasive Imaging Evidence of Subclinical Cardiovascular Disease. JACC Cardiovasc 21 *Imaging*. 2013. 22 Moon J, Messroghli D, Kellman P, Piechnik S, Robson M, Ugander M, et al. 8. 23 Myocardial T1 mapping and extracellular volume quantification: a Society for 24 Cardiovascular Magnetic Resonance (SCMR) and CMR Working Group of the European 25 Society of Cardiology consensus statement. J Cardiovasc Magn Reson. 2013;15(1):92. Schelbert EB, Messroghli DR. State of the Art: Clinical Applications of Cardiac T1 26 9. 27 Mapping. Radiology. 2016 Mar;278(3):658-76. 28 10. Gulati G, Heck SL, Ree AH, Hoffmann P, Schulz-Menger J, Fagerland MW, et al. 29 Prevention of cardiac dysfunction during adjuvant breast cancer therapy (PRADA): a 2 x 2 factorial, randomized, placebo-controlled, double-blind clinical trial of candesartan 30 31 and metoprolol. *Eur Heart J.* 2016 Jun 1;37(21):1671-80. 32 Heck SL, Gulati G, Ree AH, Schulz-Menger J, Gravdehaug B, Røsjø H, et al. 11. Rationale and Design of the Prevention of Cardiac Dysfunction during an Adjuvant 33 34 Breast Cancer Therapy (PRADA) Trial. Cardiology. 2012;123(4):240-7. 35 Messroghli DR, Greiser A, Frohlich M, Dietz R, Schulz-Menger J. Optimization and 12. 36 validation of a fully-integrated pulse sequence for modified look-locker inversion-37 recovery (MOLLI) T1 mapping of the heart. J Magn Reson Imaging. 2007 38 Oct;26(4):1081-6. 39 Schulz-Menger J, Bluemke DA, Bremerich J, Flamm SD, Fogel MA, Friedrich MG, et 13. 40 al. Standardized image interpretation and post processing in cardiovascular magnetic 41 resonance: Society for Cardiovascular Magnetic Resonance (SCMR) board of trustees 42 task force on standardized post processing. J Cardiovasc Magn Reson. 2013;15:35. 43 Cerqueira MD, Weissman NJ, Dilsizian V, Jacobs AK, Kaul S, Laskev WK, et al. 14. 44 Standardized myocardial segmentation and nomenclature for tomographic imaging of 45 the heart. A statement for healthcare professionals from the Cardiac Imaging Committee 46 of the Council on Clinical Cardiology of the American Heart Association. *Circulation*. 47 2002 Jan 29;105(4):539-42.

1 15. White SK, Sado DM, Fontana M, Banypersad SM, Maestrini V, Flett AS, et al. T1 2 Mapping for Myocardial Extracellular Volume Measurement by CMRBolus Only Versus 3 Primed Infusion Technique. JACC Cardiovasc Imaging. 2013 Apr 5. 4 Neilan TG, Coelho-Filho OR, Pena-Herrera D, Shah RV, Jerosch-Herold M, Francis 16. 5 SA, et al. Left ventricular mass in patients with a cardiomyopathy after treatment with 6 anthracyclines. Am J Cardiol. 2012 Dec 1;110(11):1679-86. 7 Jordan JH, D'Agostino RB, Jr., Hamilton CA, Vasu S, Hall ME, Kitzman DW, et al. 17. 8 Longitudinal assessment of concurrent changes in left ventricular ejection fraction and 9 left ventricular myocardial tissue characteristics after administration of cardiotoxic 10 chemotherapies using T1-weighted and T2-weighted cardiovascular magnetic resonance. *Circ Cardiovasc Imaging*. 2014 Nov;7(6):872-9. 11 12 Hong Y, Park C, Kim Y, Hur J, Lee H-J, Hong S, et al. Extracellular volume fraction 18. 13 in dilated cardiomyopathy patients without obvious late gadolinium enhancement: comparison with healthy control subjects. The International Journal of Cardiovascular 14 15 *Imaging*. 2015 2015/06/01;**31**(1):115-22. de Meester de Ravenstein C, Bouzin C, Lazam S, Boulif J, Amzulescu M, Melchior J, 16 19. 17 et al. Histological Validation of measurement of diffuse interstitial myocardial fibrosis 18 by myocardial extravascular volume fraction from Modified Look-Locker imaging 19 (MOLLI) T1 mapping at 3 T. J Cardiovasc Magn Reson. 2015;17:48. 20 Doltra A, Messroghli D, Stawowy P, Hassel JH, Gebker R, Leppänen O, et al. 20. 21 Potential Reduction of Interstitial Myocardial Fibrosis With Renal Denervation. Journal 22 of the American Heart Association. 2014 December 19, 2014;3(6). 23 Flett AS, Sado DM, Quarta G, Mirabel M, Pellerin D, Herrey AS, et al. Diffuse 21. 24 myocardial fibrosis in severe aortic stenosis: an equilibrium contrast cardiovascular 25 magnetic resonance study. Eur Heart J Cardiovasc Imaging. 2012 Oct; 13(10):819-26. 26 22. Neilan TG, Coelho-Filho OR, Shah RV, Feng JH, Pena-Herrera D, Mandry D, et al. 27 Myocardial Extracellular Volume by Cardiac Magnetic Resonance Imaging in Patients 28 Treated With Anthracycline-Based Chemotherapy. *The American Journal of Cardiology*. 29 2013 3/1/;111(5):717-22. 30 Meléndez GC, Jordan JH, D'Agostino RB, Vasu S, Hamilton CA, Hundley WG. 23. 31 Progressive 3-Month Increase in LV Myocardial ECV After Anthracycline-Based 32 Chemotherapy. JACC Cardiovasc Imaging. 2016. Sawyer DB, Peng X, Chen B, Pentassuglia L, Lim CC. Mechanisms of anthracycline 33 24. 34 cardiac injury: can we identify strategies for cardioprotection? Prog Cardiovasc Dis. 35 2010 Sep-Oct; 53(2):105-13. 36 Luetkens JA, Homsi R, Dabir D, Kuetting DL, Marx C, Doerner J, et al. 25. 37 Comprehensive Cardiac Magnetic Resonance for Short - Term Follow - Up in Acute 38 Myocarditis. Journal of the American Heart Association. 2016;5(7). 39 von Knobelsdorff-Brenkenhoff F, Schuler J, Doganguzel S, Dieringer MA, Rudolph 26. 40 A, Greiser A, et al. Detection and Monitoring of Acute Myocarditis Applying Quantitative 41 Cardiovascular Magnetic Resonance. *Circ Cardiovasc Imaging*. 2017 Feb;**10**(2). 42 Farhad H, Staziaki PV, Addison D, Coelho-Filho OR, Shah RV, Mitchell RN, et al. 27. 43 Characterization of the Changes in Cardiac Structure and Function in Mice Treated With 44 Anthracyclines Using Serial Cardiac Magnetic Resonance Imaging. Circ Cardiovasc 45 Imaging. 2016;9(12). Germain P, El Ghannudi S, Jeung M-Y, Ohlmann P, Epailly E, Roy C, et al. Native T1 46 28. 47 Mapping of the Heart – A Pictorial Review. *Clinical Medicine Insights Cardiology*.

48 2014;**8**(Suppl 4):1-11.

- 1 Narayan HK, Finkelman BS, French B, Plappert T, Hyman D, Smith AM, et al. 2 Detailed Echocardiographic Phenotyping in Breast Cancer Patients: Associations with 3 Ejection Fraction Decline, Recovery, and Heart Failure Symptoms over 3 Years of 4 Followup. Circulation. 2017. 5 Lipshultz SE, Lipsitz SR, Sallan SE, Dalton VM, Mone SM, Gelber RD, et al. Chronic 30. 6 progressive cardiac dysfunction years after doxorubicin therapy for childhood acute 7 lymphoblastic leukemia. *J Clin Oncol.* 2005 Apr 20;23(12):2629-36. 8 Toko H, Oka T, Zou Y, Sakamoto M, Mizukami M, Sano M, et al. Angiotensin II type 31. 9 1a receptor mediates doxorubicin-induced cardiomyopathy. *Hypertens Res.* 10 2002;25(4):597 - 603. 32. Baker KM, Aceto JF. Angiotensin II stimulation of protein synthesis and cell 11 12 growth in chick heart cells. Am J Physiol. 1990 Aug; 259(2 Pt 2): H610-8. 13 33. Zaman MA, Oparil S, Calhoun DA. Drugs targeting the renin-angiotensinaldosterone system. *Nat Rev Drug Discov*. 2002 08//print;**1**(8):621-36. 14 Susic D, Nuñez E, Frohlich ED, Prakash O. Angiotensin II Increases Left 15 34. 16 Ventricular Mass Without Affecting Myosin Isoform mRNAs. Hypertension. 1996 August 17 1, 1996;28(2):265-8. Ariff B, Zambanini A, Vamadeva S, Barratt D, Xu Y, Sever P, et al. Candesartan-18 35. 19 and Atenolol-Based Treatments Induce Different Patterns of Carotid Artery and Left 20 Ventricular Remodeling in Hypertension. *Stroke*. 2006;**37**(9):2381-4. 21 Wang Z, Niu Q, Peng X, Li M, Liu K, Liu Y, et al. Candesartan cilexetil attenuated 36. 22 cardiac remodeling by improving expression and function of mitofusin 2 in SHR. Int J
- 23 *Cardiol.* 2016 7/1/;**214**:348-57.
- 24 Zamorano JL, Lancellotti P, Rodriguez Muñoz D, Aboyans V, Asteggiano R, 37.
- 25 Galderisi M, et al. 2016 ESC Position Paper on cancer treatments and cardiovascular
- 26 toxicity developed under the auspices of the ESC Committee for Practice Guidelines The
- 27 Task Force for cancer treatments and cardiovascular toxicity of the European Society of
- 28 Cardiology (ESC). *Eur Heart J.* 2016;**37**(36):2768-801.
- 29

29.

	Candesartan	Candesartan	Placebo	Placebo	
	Metoprolol	Placebo	Metoprolol	Placebo	p-value
Ν	18	20	13	18	
Age at recruitment (years)	49.8 ± 8.9	52.6 ± 10.2	49.2 ± 8.1	50.3 ± 9.6	0.725
Systolic blood pressure (mmHg)	120.5 (119.8, 131.5)	132.0 (116.0, 144.8)	135.0 (127.5, 140.0)	131.5 (120.0, 141.3)	0.213
Diastolic blood pressure (mmHg)	80.0 (70, 92)	80.0 (75.0, 87.8)	80.0 (75.0, 92.0)	80.0 (77.3, 91.3)	0.741
Heart rate (beats/min)	66.5 (63.5, 71)	66.5 (59.0, 73.0)	65.0 (61.0, 77.5)	64.0 (57.0, 73.3)	0.897
Body mass index kg/m ²	24.2 (22.4, 25.4)	25.7 (21.8, 28.8)	28.6 (22.5, 31.1)	24.2 (21.4, 28.2)	0.364
Blood haemoglobin (g/dL)	13.3 ± 0.8	13.3 ± 1.0	13.2 ± 0.6	13.3 ± 0.8	0.991
Serum creatinine (mg/dl)	0.73 (0.64, 0.86)	0.71 (0.68, 0.76)	0.73 (0.69, 0.86)	0.72 (0.68, 0.80)	0.527
LVEF (%)*	62.4 ± 4.4	61.7 ± 4.8	64.4 ± 5.6	63.4 ± 3.4	0.340
LV mass (g)	79.9 ± 14.3	89.0 ± 15.0	79.2 ± 15.0	85.8 ± 14.6	0.160
LV mass (g/m ^{1.7})	33.9±5.7	36.9±5.4	33.4±5.2	35.3±5.4	0.223
Current smokers	3/18 (16.7%)	5/20 (25%)	2/13 (15.4%)	4/18 (22.2%)	0.913
Hypertension	1/18 (5.6%)	3/20 (15.0%)	1/13 (7.7%)	0/18 (0.0%)	0.442
Diabetes	0/18 (0.0%)	1/20 (5.0%)	1/13 (7.7%)	0/18 (0.0%)	0.555
Late gadolinium enhancement	0/18 (0.0%)	0/20 (0.0%)	1/13 (7.7%)	1/18 (5.6%)	0.444
Concomitant statin use	2/18 (11.1%)	1/20 (5.0%)	1/13 (7.7%)	4/18 (22.2%)	0.424
Concomitant diuretic use	0/18 (0.0%)	4/20 (20.0%)	0/13 (0.0%)	0/18 (0.0%)	0.014
Doxorubicin equivalent mg/m ²	160.8 (160.8,241.2)	160.8 (160.8,241.2)	160.8 (160.8,201.0)	201.0 (160.8,268.0)	0.410

Table 1: Baseline characteristics and cancer treatment of the study population

Data are expressed as mean ± SD (ANOVA), median (25, 75 percentile) (Kruskal-Wallis h test) or numbers (percent) (Fischers exact

test). * LVEF denotes left ventricular ejection fraction

		Ν	Baseline	After anthracyclines	Change (95%CI)	p-value	Between groups difference (95%CI)	p- value
LVEF, % [*]	All	69	62.8 ± 4.6	61.1 ± 4.4	-1.7 (-2.8, -0.6)	0.003		
	No candesartan	31	63.9 ± 4.4	60.7 ± 4.1	-3.2 (-4.9, -1.5)	0.001		
	Candesartan	38	62.0 ± 4.5	61.5 ± 4.7	-0.5 (-1.9, 0.9)	0.454	-2.7 (-4.8, -0.5)	0.015
ECV fraction, % †	All	69	27.5 ± 2.7	28.6 ± 2.9	1.2 (0.4, 1.9)	0.002		
	No candesartan	31	27.6 ± 2.6	28.6 ± 2.8	1.0 (-0.1, 2.2)	0.072		0.790
	Candesartan	38	27.4 ± 2.7	28.7 ± 3.1	1.2 (0.3, 2.2)	0.013	-0.2 (-1.6, 1.3)	
T1, ms	All	69	1005 ± 32	1011 ± 33	6 (-2, 15)	0.147		
	No candesartan	31	1004 ± 33	1011 ± 32	7 (-6, 19)	0.292		
	Candesartan	38	1005 ± 31	1011 ± 35	6 (-6, 18)	0.323	1 (-17, 18)	0.949
Total ECV, ml	All	69	22.0 ± 4.4	22.4 ± 4.2	0.4 (-0.3, 1.1)	0.240		
	No candesartan	31	21.8 ± 4.5	22.6 ± 3.9	0.8 (-0.1, 1.7)	0.070		
	Candesartan	38	22.1 ± 4.3	22.1 ± 4.4	0.1 (-0.9, 1.0)	0.906	0.7 (-0.6, 2.1)	0.266
Total cellular	All	69	58.0 ± 10.7	55.8 ± 9.9	-2.2 (-3.2, -1.2)	< 0.001		
volume, ml	No candesartan	31	57.3 ± 10.4	56.7 ± 9.9	-0.6 (-2.1, 0.9)	0.448		
	Candesartan	38	58.6 ± 11.1	55.1 ± 9.9	-3.5 (-4.7, -2.2)	< 0.001	2.9 (1.0, 4.8)	0.003

Table 2: Change from baseline to completion of anthracycline therapy. No candesartan treatment vs. candesartan treatment

Values are mean \pm SD. ^{*}LVEF denotes left ventricular ejection fraction; [†] ECV extracellular volume.

		Ν	Baseline	After anthracyclines	Change (95%CI)	p-value	Between groups difference (95%CI)	p-value
	Lower [*]	58	62.6 ± 4.2	61.6 ± 4.3	-1.1 (-2.2, 0.1)	0.063	4.1 (1.2, 6.9)	0.006
LVEF, %	Higher	11	63.9 ± 6.3	58.8 ± 4.6	-5.1 (-8.4, -1.9)	0.006		
ECV ^{††}	Lower	58	27.5 ± 2.6	28.3 ± 2.6	0.7 (0.0, 1.5)	0.049	-2.6 (-4.5, -0.8)	0.006
fraction, %	Higher	11	27.3 ± 3.1	30.6 ± 3.7	3.4 (1.2, 5.5)	0.006		
	Lower	58	1004 ± 32	1012 ± 34	8 (-3, 18)	0.138	8 (-4, 21)	0.197
T1, ms	Higher	11	1006 ± 31	1006 ± 31	-1 (-9, 8)	0.880		
	Lower	58	22.1 ± 4.5	22.2 ± 4.3	0.1 (-0.6, 0.8)	0.785		0.040
Total ECV, ml	Higher	11	21.0 ± 3.7	23.0 ± 3.2	1.9 (0.4, 3.5)	0.020	-1.8 (-3.6, -0.1)	
Total cellular	Lower	58	58.4 ± 11.3	56.4 ± 9.9	-1.9 (-3.1, -0.8)	0.001		0.000
volume, ml	Higher	11	55.9 ± 7.1	52.5 ± 9.4	-3.4 (-5.8, -1.0)	0.010	1.5 (-1.3, 4.2)	0.290

Table 3: Change from baseline to completion of anthracycline therapy according to anthracycline dose

Values are mean \pm SD. ^{*}Lower denotes 40 mg/kg² x 4 or 6; higher denotes 67 mg/kg² x 4 doxorubicin equivalent, [†] LVEF denotes left ventricular ejection fraction; ^{††} ECV extracellular volume.

Figure Legends

Figure 1

Example of native and post contrast T1 maps with endocardial, epicardial and blood volume contouring

Figure 2

Change in extracellular volume (ECV) fraction (%), total ECV (ml), and total cellular volume

(ml) from baseline to end of anthracycline treatment. Lower denotes 40 mg/kg² x 4 or 6;

higher denotes 67 mg/kg² x 4 doxorubicin equivalent

Figure 3

Increased ECV fraction may be caused by increased total extracellular volume or reduced total cellular volume. ECV denotes extracellular volume.







Change during anthracycline therapy



* Between group difference < 0.05

Figure 3

High dose anthracyclines



Normal myocardium

Candesartan



Increased ECV fraction through increased total ECV



Increased ECV fraction through decreased total cellular volume