

Altered β_{1-3} -adrenoceptor influence on α_2 -adrenoceptor-mediated control of catecholamine release and vascular tension in hypertensive rats

Torill Berg*

Division of Physiology, Department of Molecular Medicine, Institute of Basic Medical Sciences, University of Oslo, Oslo, Norway

OPEN ACCESS

Edited by:

James J. Galligan,
Michigan State University, USA

Reviewed by:

Deborah H. Damon,
University of Vermont, USA
Brendan J. Canning,
Johns Hopkins School of Medicine,
USA

*Correspondence:

Torill Berg,
Division of Physiology, Department of
Molecular Medicine, Institute of Basic
Medical Sciences, University of Oslo,
Sognsvannsveien 9, PO Box 1103,
Blindern, 0317 Oslo, Norway
torill.berg@medisin.uio.no

Specialty section:

This article was submitted to
Autonomic Neuroscience,
a section of the journal
Frontiers in Physiology

Received: 09 January 2015

Accepted: 31 March 2015

Published: 20 April 2015

Citation:

Berg T (2015) Altered
 β_{1-3} -adrenoceptor influence on
 α_2 -adrenoceptor-mediated control of
catecholamine release and vascular
tension in hypertensive rats.
Front. Physiol. 6:120.
doi: 10.3389/fphys.2015.00120

α_2 - and β -adrenoceptors (AR) reciprocally control catecholamine release and vascular tension. Disorders in these functions are present in spontaneously hypertensive rats (SHR). The present study tested if α_2 AR dysfunctions resulted from altered α_2 AR/ β AR interaction. Blood pressure (BP) was recorded through a femoral artery catheter and cardiac output by an ascending aorta flow probe. Total peripheral vascular resistance (TPR) was calculated. Norepinephrine release was stimulated by a 15-min tyramine-infusion, which allows presynaptic release-control to be reflected as differences in overflow to plasma. Surgical stress activated some secretion of epinephrine. L-659,066 (α_2 AR-antagonist) enhanced norepinephrine overflow in normotensive controls (WKY) but not SHR. Nadolol (β_{1+2}) and ICI-118551 (β_2), but not atenolol (β_1) or SR59230A [$\beta_{(3)/1L}$] prevented this increase. All β AR antagonists allowed L-659,066 to augment tyramine-induced norepinephrine overflow in SHR and epinephrine secretion in both strains. Inhibition of cAMP-degradation with milrinone and β_3 AR agonist (BRL37344) enhanced the effect of L-659,066 on release of both catecholamines in SHR and epinephrine in WKY. $\beta_{1/2}$ AR antagonists and BRL37344 opposed the L-659,066-dependent elimination of the TPR-response to tyramine in WKY. α_2 AR/ β AR antagonists had little influence on the TPR-response in SHR. Milrinone potentiated the L-659,066-dependent reduction of the TPR-response to tyramine. Conclusions: β_2 AR activity was a required substrate for α_2 AR auto inhibition of norepinephrine release in WKY. β_{1+2} AR opposed α_2 AR inhibition of norepinephrine release in SHR and epinephrine secretion in both strains. β AR- α_2 AR reciprocal control of vascular tension was absent in SHR. Selective agonist provoked β_3 AR- G_i signaling and influenced the tyramine-induced TPR-response in WKY and catecholamine release in SHR.

Keywords: α_2 -adrenoceptors, β -adrenoceptors, hypertension, total peripheral vascular resistance, sympathetic nervous system, catecholamine release, spontaneously hypertensive rats

Introduction

α_2 - and β -adrenoceptors (AR) comprise the $\alpha_{2A,B,C}$ and the $\beta_{1,2,3}$ subtypes. By coupling to inhibitory (G_i) and stimulatory (G_s) G-proteins, respectively, $\alpha_{2A,B,C}$ - and β_{1+2} AR have opposite

effect on adenylyl cyclase, and therefore have reciprocal actions on various functions involved in the control of blood pressure (BP), including the release of catecholamines and vascular tension. The β_3 AR has been shown to couple to G_i and induce a negative inotropic effect in the heart (Gauthier et al., 1998). Transmitter release from peripheral sympathetic nerves is inhibited by presynaptic α_2 AR, primarily the α_{2A} -subtype (Trendelenburg et al., 2003; Berg and Jensen, 2013), and stimulated by presynaptic β_2 AR (Stjarne and Brundin, 1976; Westfall et al., 1979). Also the β_1 AR has recently been demonstrated to enhance norepinephrine release, providing a plausible rationale for the antihypertensive action of β_1 AR antagonists, the most frequently used β -blockers in the treatment of hypertension (Berg, 2014a). α_2 AR auto-receptors in addition inhibited adrenal epinephrine secretion in *in vivo* experiments (Brede et al., 2003; Berg et al., 2012), whereas β_1 - or β_2 AR antagonists had no effect (Berg, 2014a). α_2 AR-mediated auto inhibition of neuronal and adrenal catecholamine release has been shown to be dysfunctional in the spontaneously hypertensive rat (SHR) (Berg and Jensen, 2013). This dysfunction may contribute to the hyperadrenergic and hypertensive state in this model of human hypertensive disease, in agreement with the high plasma norepinephrine concentration and hypertension observed in α_{2A} AR-gene-deleted mice (Makaritsis et al., 1999). The failing α_2 AR auto inhibition in SHR may result from an altered interaction between different presynaptic receptors, as indicated by the restored α_2 AR function in SHR after α_{2C} AR stimulation or angiotensin AT1 receptor inhibition (Berg, 2013) (Figure 1). The β_3 AR has been shown to be less sensitive to catecholamine-induced desensitization than the β_1 - and β_2 AR (Mallem et al., 2004; Rouget et al., 2004), and a β_3 AR up-regulated and β_1 AR down-regulated relaxation was demonstrated in SHR thoracic aortic rings (Mallem et al., 2004). It may therefore be hypothesized that alterations in β AR signaling may alter α_2 AR auto inhibition of catecholamine release in SHR.

α_{2B} AR (Philipp et al., 2002) and β AR are also present in vascular smooth muscle cells (VSMC), where they modulate the α_1 AR-mediated vasoconstrictory response to norepinephrine (Figure 2). VSMC tension is in addition influenced by endothelial α_{2A} AR and β_2 AR, which both stimulate nitric oxide (NO) synthesis (Shafaroudi et al., 2005; Queen et al., 2006). Also vasodilatory and vasoconstrictory α_2 AR-mediated control of total peripheral vascular resistance (TPR) appeared dysfunctional in SHR (Berg and Jensen, 2011, 2013).

Presynaptic receptors modulate norepinephrine release from the nerve terminal vesicles. This control is not reflected by differences in norepinephrine overflow to plasma, due to that the response is terminated by re-uptake through the norepinephrine re-uptake transporter (NET). Presynaptic control of release is therefore not easily studied *in vivo*. However, tyramine stimulates norepinephrine release by reversing the transport through NET (Figure 1), and consequently prevents re-uptake through NET. Presynaptic modulation of concomitant vesicular release can therefore be demonstrated as differences in overflow to plasma (Berg et al., 2012; Berg and Jensen, 2013). The presynaptic receptors will be stimulated by the released norepinephrine and/or by other agonists present in the vicinity. Secreted epinephrine is not subjected to re-uptake through NET, and was therefore not influenced by tyramine, but was stimulated to some extent

by the stress induced by the experiment itself (Berg et al., 2012). Thus, the inhibitory effect of presynaptic α_2 AR on catecholamine release could be demonstrated by the ability of the non-selective, peripherally restricted, α_2 AR antagonist L-659,066 to increase tyramine-induced norepinephrine overflow and epinephrine secretion (Berg and Jensen, 2013). The stimulating effect of presynaptic β AR was demonstrated by pre-treatment with β_{1or2} AR antagonists (Berg, 2014a). Inhibition of release could be stimulated in SHR by the β_3 AR agonist BRL37344, compatible with stimulation of β_3 AR- G_i activation (Berg, 2014b). The reduced release observed after the β_3 AR antagonist SR59230A was likely to result from its inhibitory effect on the G_s -coupled low-affinity state β_1 AR (β_{1L} AR) (Berg, 2014b). The use of tyramine also allowed a concomitant examination of the role of post-synaptic α_2 AR and β AR in the cardiovascular response to the released norepinephrine (Figure 2).

The purpose of the present investigation was therefore to gain a better understanding of the reason underlying the failing α_2 AR auto-inhibition of catecholamine release in SHR, i.e., to test if there was a difference in the interaction between the α_2 AR and the three β AR subtypes in this strain compared to that in their normotensive controls (WKY). A second goal was to test if a difference in β_{1-3} AR activity was responsible for the failing α_2 AR influence on the TPR-response to the tyramine-stimulated norepinephrine release in SHR.

Materials and Methods

Experimental Procedure

All experiments were approved by The Norwegian Animal Research Authority (NARA) and conducted in accordance with the Directive 2010/63/EU of the European Parliament. Male, 12–14 weeks old SHR (Okamoto, SHR/NHsd strain, 273 ± 2 g body weight, $n = 109$) and their normotensive control, i.e., WKY (Wistar Kyoto, 279 ± 9 g body weight, $n = 124$) on conventional rat chow diet (0.7% NaCl) were anesthetized with sodium pentobarbital (65–70 mg/kg, IP) and tracheotomized. A heparinized catheter was inserted into the femoral artery to record systolic (SBP) and diastolic (DBP) BP. The rats were subsequently connected to a positive-pressure respirator and ventilated with air throughout the experiment. Cardiac output (CO, i.e., minus cardiac flow) and heart rate (HR) were recorded by a flow probe on the ascending aorta, connected to a T206 Ultrasonic Transit-Time Flowmeter (Transonic Systems Inc., Ithaca, NY, USA). After surgery was completed, the arterial catheter was flushed with 0.15 ml heparinized (1000 U/ml) phosphate-buffered saline (PBS; 0.01 M Na-phosphate, pH 7.4, 0.14 M NaCl). Mean arterial BP [MBP = (SBP-DBP/3) + DBP] and TPR (=MBP/CO) were calculated. Body temperature was maintained at $37 - 38^\circ\text{C}$ by external heating, guided by of a thermo sensor inserted inguinally into the abdominal cavity. Drugs were dissolved in PBS and administered as bolus injections (0.6–1 ml/kg) through a catheter in the femoral vein, unless otherwise indicated.

Experimental Design

After a control period of about 10 min, control rats were pre-treated with vehicle (PBS), and subsequently infused with tyramine for 15 min ($1.26 \mu\text{mol/kg/min}$) (Berg, 2005; Berg and

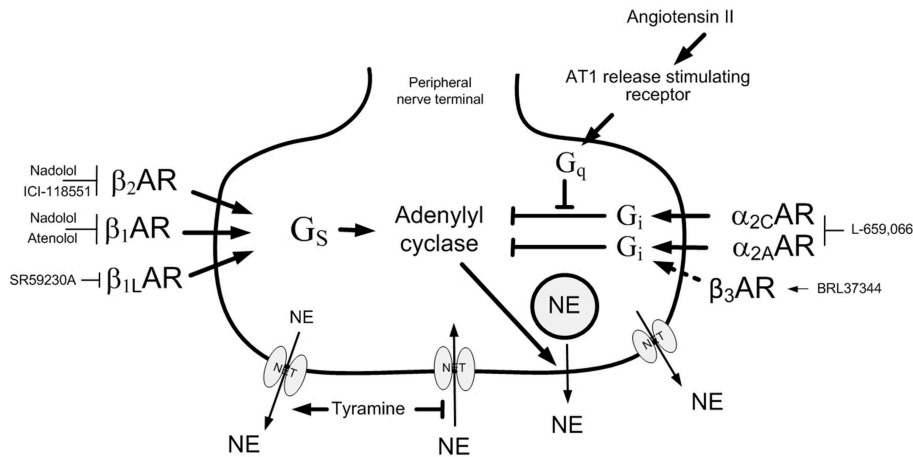


FIGURE 1 | Control of norepinephrine release from peripheral sympathetic nerve endings. Tyramine stimulates norepinephrine release by reverse transport through NET. Consequently, re-uptake through NET is prevented, and presynaptic modulation of vesicular release is reflected as differences in overflow to plasma (Berg et al., 2012; Berg and Jensen, 2013). The release of norepinephrine from secretory granules is activated by adenylyl cyclase, which is stimulated by $\beta_{1,1L,2}AR$ - G_s and inhibited by α_2AR - G_i . In WKY, α_2AR auto

inhibition required β_2AR activity, but was independent of β_1AR signaling. In SHR, blocking $\beta_{1,1L,2}AR$ activity allowed α_2AR inhibition of release. The β_3AR -selective agonist BRL37344 reduced norepinephrine overflow in SHR but not WKY (dotted arrow). The β_3AR antagonist SR59230A reduced overflow apparently due to its ability to inhibit $\beta_{1L}AR$ and not the β_3AR . The action of antagonists and agonist are indicated. NE, norepinephrine; Pointed arrows, positive effects; Blunted arrows, inhibitory actions.

Jensen, 2013). In the experimental groups, the rats were pre-treated with drugs to modify presynaptic and postsynaptic α_2AR or βAR signaling, separately or combined.

Inhibition of βAR -Signaling

Inhibition of βAR -signaling was achieved by βAR -antagonists, the peripherally restricted, i.e., which does not cross the blood-brain barrier, nadolol (β_{1+2} , 8.5 $\mu\text{mol/kg}$), or atenolol (β_1 , 5.6 $\mu\text{mol/kg}$), the not-restricted ICI-118551 (β_2 , 1 $\mu\text{mol/kg}$ initial dose, then 0.3 $\mu\text{mol/kg/min}$ throughout the experiment) or the β_3AR antagonist SR59230A (13.8 $\mu\text{mol/kg}$) (Berg et al., 2010). SR59230A also inhibited the putative β_4AR (Malinowska and Schlicker, 1997), later identified as $\beta_{1L}AR$ (Kaumann et al., 2001).

Amplification of βAR -Signaling

Amplification of βAR -signaling was achieved by pre-treatment with the phosphodiesterase 3 (PDE3) inhibitor milrinone (1.4 μmol) (Berg et al., 2009) which will prevent degradation of cyclic AMP (cAMP) (Figure 2). The β_3AR was stimulated by the agonist BRL37344 ((\pm)-(R*,R*)-[4-[2-[[2-(3-Chlorophenyl)-2-hydroxyethyl]amino]propyl]phenoxy]-acetic acid sodium hydrate) infused at a rate of 1 nmol/kg/min throughout the experiment (Malinowska and Schlicker, 1997; Berg, 2014b).

Inhibition of α_2AR -Signaling

Inhibition of α_2AR -signaling was achieved by pre-treatment with the none-selective, peripherally restricted α_2AR antagonist L-659,066 (Clineschmidt et al., 1988) (4.4 $\mu\text{mol/kg}$) (Berg et al., 2012; Berg and Jensen, 2013).

Inhibition of G_i Signaling

Since G_i represents a main signaling pathway for all α_2AR , G_i -signaling was abolished by the G_i -inhibitor *Bordetella pertussis* toxin (PTX, 15 $\mu\text{g/kg}$, i.p., -48 h) (Anand-Srivastava et al., 1987). The latter rats were pre-treated with PBS during the experiment.

The Interaction between α_2AR and βAR Signaling

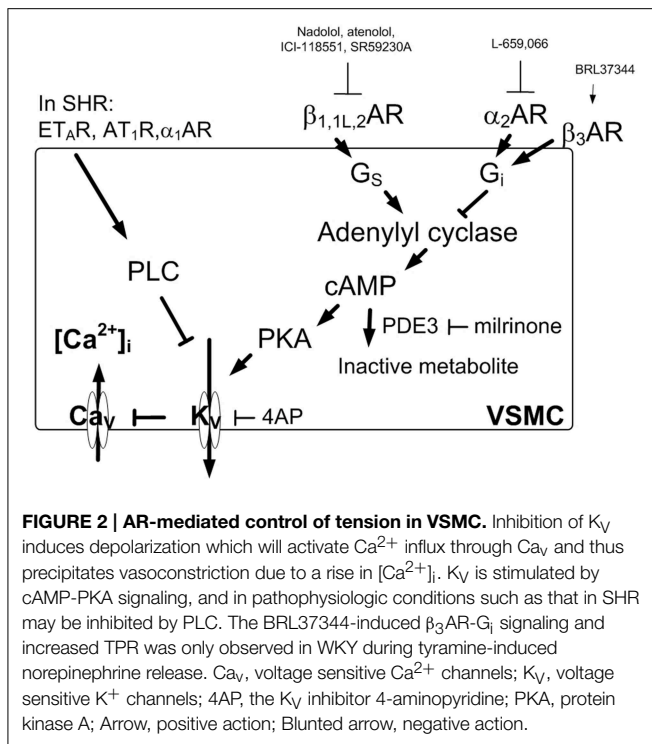
The interaction between α_2AR and βAR signaling was studied by combining βAR antagonists/agonist/milrinone with the α_2AR antagonist L-659,066 as indicated in Table 1. Ten min was allowed between drugs, except for SR59230A which was followed by L-659,066 or tyramine after 5 min.

Measurement of Plasma Catecholamines

1.5 ml blood was collected from the femoral artery into tubes containing 45 μl 0.2 M glutathione and 0.2 M ethylene glycol-bis(2-aminoethylether)-N,N,N',N'-tetraacetic acid (EGTA) (4°C). Plasma was stored at -80°C until catecholamine concentrations were determined using 400 μl plasma and the 5000 Reagent kit for HPLC analysis of catecholamines in plasma from chromsystems GmbH, Munich, Germany, as described by the manufacturer. The samples were run on a Shimadzu monoamines analyzer system, using an isocratic flow rate of 0.8 ml/min, and an electrochemical detector (Decade II) and a SenCell electrochemical flow cell (Antec Leyden, Zoeterwoude, The Netherlands).

Drugs

L-659,066 was a kind gift from Merck, Sharp and Dohme Labs, Rahway, NJ. ICI-118551 was obtained from ICI-Pharma,



Cheshire, UK; SR59230A from Santa Cruz Biotechnology, Heidelberg, Germany; and pentobarbital from The Norwegian National Hospital, Oslo, Norway. The remaining drugs were from Sigma Chemical Co., St. Louis, MO, USA.

Statistical Analyses

Results are presented as mean values \pm s.e.m. Effect of pre-treatment, differences in the cardiovascular baselines, (data averaged every min), and the plasma catecholamine concentrations were evaluated by overall tests (One-Way ANOVA), followed by two-tailed two-sample Student's *t*-tests. The cardiovascular response-curves to tyramine (data averaged every min) were analyzed using Repeated Measures Analyses of Variance and Covariance, first as over-all tests, subsequently for each group separately or between two groups. Significant responses and groups differences were subsequently located at specific times using two-tailed one- and two-sample Student's *t*-tests, respectively. For non-parametric data, two-sample Student's *t*-tests were substituted by Kruskal-Wallis tests. At each step, testing proceeded only when the presence of significant differences and/or interactions was indicated. For the cardiovascular data, the *P*-value was for all tests and each step adjusted according to Bonferroni, whereas $P \leq 0.05$ was considered significant for the catecholamine data.

In those cases, where comparisons were made with rats included in previous publications (Table 1), these experiments were performed, in part or in full, intermittently with the present groups. When this was not the case, some of the previous experiments were substituted with new to ensure that all experiments overlapped in time. Control rats were included randomly throughout the study.

Results

The Influence of β_{1-3} AR Signaling on α_2 AR-mediated Inhibition of Tyramine-induced Norepinephrine Overflow to Plasma The Effect of Modulating β AR Signaling

As previously described (Berg and Jensen, 2013; Berg, 2014a), all β AR antagonists, including SR59230A, reduced the tyramine-induced overflow of norepinephrine to plasma in both strains (Figure 3). Since the effect of SR59230A was likely to involve inhibition of β_{1L} AR rather than β_3 AR (Berg, 2014b), this result demonstrated that the overflow was enhanced by β_{1+1L+2} AR activation. However, the β_3 AR agonist BRL37344 precipitated a minor reduction in overflow in SHR but not WKY (Figure 4), compatible with a G_i -mediated inhibition of release in SHR (Berg, 2014b).

The Effect of inhibiting α_2 AR- G_i Signaling

The G_i inhibitor PTX had no significant effect on the tyramine-induced norepinephrine overflow in either strain (Figure 4). As before (Berg and Jensen, 2013), the α_2 AR antagonist L-659,066 increased norepinephrine overflow in WKY, but the rise was not statistically significant in SHR (Figures 3, 4).

The Interaction between α_2 AR and β AR Signaling

L-659,066 eliminated the reduction in the tyramine-induced overflow induced by the β_1 AR antagonist atenolol in WKY ($P = NS$ compared to the controls or L-659,066 alone, $P = 0.004$ compared to atenolol alone) (Figure 3). L-659,066 also totally reversed the reduction following pre-treatment with the $\beta_{(3)+1L}$ AR antagonist SR59230A ($P = 0.005$ compared to SR59230A alone, $P = NS$ compared to the controls and L-659,066 alone) (Figure 3). However, L-659,066 counter-acted only in part the reduction following the β_{1+2} AR antagonist nadolol ($P \leq 0.004$ compared to the WKY controls and L-659,066 alone, $P = 0.049$ compared to nadolol alone), and did not at all counter-act the reduction induced by the β_2 -selective antagonist ICI-118551 ($P = 0.05$ and 0.005 compared to the controls and L-659,066 alone, respectively, $P = NS$ compared to ICI-118551 alone) (Figure 3). These results demonstrated that α_2 AR antagonist increased tyramine-stimulated norepinephrine release in WKY also in the presence of β_1 AR and $\beta_{(3)+1L}$ AR antagonist, but not when the β_2 AR subtype was blocked.

The plasma norepinephrine concentration in L-659,066 + milrinone-treated WKY was higher than that in the controls ($P = 0.05$) but not different from that after L-659,066 alone (Figure 4). This result showed that enhanced cAMP signaling, i.e., down-stream of adenylyl cyclase (Figure 2), did not influence the response to L-659,066 in this strain. BRL37344 did not alter the effect of L-659,066 on tyramine-induced norepinephrine release (Figure 4), demonstrating that β_3 AR- G_i signaling did not contribute to the response in WKY.

In SHR, L-659,066 eliminated the reduction in tyramine-induced norepinephrine overflow induced by all four β AR antagonists ($P \leq 0.003$ compared to β AR-antagonist alone), and was higher than that in the controls in all groups ($P \leq 0.018$), except in the ICI-118551 + L-659,066-treated group ($P = NS$) (Figure 3). These results demonstrated that α_2 AR-mediated

TABLE 1 | Cardiovascular baselines after pre-treatment, i.e., prior to tyramine and, in parenthesis, the response to pre-treatment.

Pre-treatment	WKY				SHR			
	MBP mm Hg	HR beats/min	CO ml/min	TPR mm Hg/ml/min	MBP mm Hg	HR beats/min	CO ml/min	TPR mm Hg/ml/min
PBS (control) ^b	66 ± 3 (-3 ± 2)	341 ± 8 (-6 ± 4)	32 ± 2 (2 ± 0)	2.1 ± 0.1 (-0.3 ± 0.1)	92 ± 5* (-5 ± 5)	391 ± 5* (-16 ± 5)	19 ± 1* (0 ± 1)	4.6 ± 0.2* (-0.3 ± 0.2)
INHIBITION OF βAR SIGNALING								
Nadolol (β_{1+2} ANT) ^b	65 ± 2 (-3 ± 1)	338 ± 5 (-21 ± 6)	31 ± 2 (1 ± 1)	2.1 ± 0.1 (-0.2 ± 0.0)	64 ± 5 [†] (-25 ± 2) [†]	345 ± 10 [†] (-80 ± 7) [†]	13 ± 2 [†] (-1 ± 1)	4.6 ± 0.5 (-2.0 ± 0.8) [†]
Atenolol (β_1 ANT) ^b	61 ± 2 (-12 ± 1)	338 ± 9 (-32 ± 7) [†]	28 ± 2 (1 ± 1)	2.4 ± 0.1 (-0.5 ± 0.1)	63 ± 3 [†] (-32 ± 5) [†]	338 ± 9 (-69 ± 10) [†]	13 ± 1 [†] (-4 ± 1)	5.1 ± 0.4 (-0.4 ± 0.5)
ICI-118551 (β_2 ANT) ^b	50 ± 4 (-6 ± 2)	311 ± 12 (-31 ± 4) [†]	28 ± 4 (2 ± 1)	1.8 ± 0.1 (-0.4 ± 0.1)	70 ± 4 (-23 ± 4)	358 ± 7 [†] (-59 ± 8) [†]	18 ± 1 (1 ± 1)	4.3 ± 0.2 (-1.2 ± 0.2)
SR59230A ($\beta_3 + \beta_{1L}$ ANT) ^b	84 ± 4 [†] (10 ± 3) [†]	390 ± 8 [†] (30 ± 7) [†]	27 ± 2 (4 ± 1)	3.3 ± 0.3 [†] (-0.2 ± 0.2)	90 ± 8 (-6 ± 5)	424 ± 13 (-21 ± 5)	18 ± 2 (0 ± 1)	5.1 ± 0.4 (-0.5 ± 0.3)
STIMULATION OF THE βAR SIGNALING PATHWAY								
Milrinone (PDE3 INH)	40 ± 2 [†] (-35 ± 1) [†]	343 ± 7 (2 ± 8)	30 ± 2 (-2 ± 1)	1.4 ± 0.1 [†] (-1.0 ± 0.1) [†]	52 ± 3 [†] (-59 ± 7) [†]	413 ± 14 (-1 ± 9)	18 ± 1 (-4 ± 1)	2.9 ± 0.2 [†] (-2.4 ± 0.5) [†]
BRL44408 (β_3 AGON) ^b	51 ± 2 [†] (3 ± 3)	312 ± 9 (-8 ± 9)	44 ± 4 (9 ± 2) [†]	1.2 ± 0.1 [†] (-0.6 ± 0.1) [†]	109 ± 7 (-9 ± 7)	409 ± 15 (0 ± 9)	23 ± 1 [†] (4 ± 1) [†]	4.8 ± 0.3 (-0.4 ± 0.3)
INHIBITION OF α_2AR-G_i SIGNALING								
PBS after PTX (G _i INH) ^a	52 ± 3 (-2 ± 2)	338 ± 9 (-21 ± 4)	39 ± 4 (3 ± 1)	1.3 ± 0.2 [†] (-0.1 ± 0.1)	65 ± 5 (-8 ± 6)	389 ± 12 (-24 ± 11)	22 ± 3 (-1 ± 2)	3.3 ± 0.5 (-0.5 ± 0.2)
L-659,066 (α_2 ANT) ^b	59 ± 7 (-17 ± 2) [†]	345 ± 11 (-10 ± 5)	35 ± 2 (2 ± 0)	1.7 ± 0.1 (-0.6 ± 0.0) [†]	69 ± 6 [†] (-21 ± 4) [†]	398 ± 12 (-16 ± 10)	17 ± 1 (-1 ± 0)	4.3 ± 0.4 (-1.1 ± 0.2)
INTERACTION BETWEEN βAR AND α_2AR SIGNALING								
Nadolol + L-659,066	47 ± 6 [†] (-6 ± 2) [†]	323 ± 8 (-30 ± 10)	23 ± 3 (3 ± 2)	2.0 ± 0.1 (-0.7 ± 0.2)	61 ± 4 [†] (-18 ± 7)	356 ± 8 [†] (-69 ± 9) [†]	15 ± 2 (-1 ± 1)	4.4 ± 0.4 (-0.8 ± 0.6)
Atenolol + L-659,066	56 ± 5 (-18 ± 5) [†]	340 ± 10 (-52 ± 18) [†]	27 ± 4 (5 ± 2)	2.2 ± 0.2 (-1.1 ± 0.3) [†]	67 ± 5 (-35 ± 4) [†]	363 ± 8 (-90 ± 7) [†]	14 ± 1 (0 ± 1)	4.7 ± 0.3 (-2.3 ± 0.2) [†]
ICI-118551 + L-659,066	49 ± 4 (-26 ± 3) [†]	336 ± 7 (-39 ± 7) [†]	26 ± 1 (1 ± 1)	1.9 ± 0.2 (-1.2 ± 0.1) [†]	77 ± 7 (-16 ± 9)	356 ± 4 [†] (-68 ± 1) [†]	19 ± 1 (0 ± 1)	4.0 ± 0.1 [†] (-1.1 ± 0.2)
SR59230A + L-659,066	55 ± 6 [†] (-3 ± 4) [†]	364 ± 25 (23 ± 9) [†]	28 ± 3 (2 ± 3)	2.0 ± 0.2 [†] (-0.3 ± 0.2)	88 ± 9 (-3 ± 9)	408 ± 11 (2 ± 9)	19 ± 2 (-1 ± 1)	4.7 ± 0.4 (0.2 ± 0.4)
Milrinone + L-659,066	37 ± 1 [†] (-42 ± 6) [†]	366 ± 12 (-5 ± 5)	28 ± 2 (1 ± 2)	1.3 ± 0.1 [†] (-1.5 ± 0.2) [†]	39 ± 2 [†] (-62 ± 6) [†]	431 ± 9 [†] (0 ± 15)	19 ± 1 (-2 ± 1)	2.2 ± 0.2 [†] (-3.0 ± 0.4) [†]
BRL44408 + L-659,066	41 ± 4 [†] (-28 ± 6) [†]	379 ± 11 [†] (20 ± 9)	45 ± 8 (10 ± 7)	1.0 ± 0.1 [†] (-1.0 ± 0.1) [†]	77 ± 8 [†] (-33 ± 9) [†]	407 ± 5 (2 ± 6)	25 ± 1 [†] (4 ± 2)	3.1 ± 0.2 [†] (-2.4 ± 0.5) [†]

^aSince PTX was given 4 h prior to the experiment, its effect was reflected as differences in the baselines themselves. Comparisons were made between WKY and SHR controls (* after SHR-values) and between the PBS-controls and the experimental groups, using the response to PBS to evaluate the effect of pre-treatment ([†]). Comparisons were also made between groups pre-treated with β AR-antagonist + α_2 AR-antagonist and corresponding groups pre-treated with β AR-antagonist alone ([†]) or α_2 AR-antagonist alone ([†]).

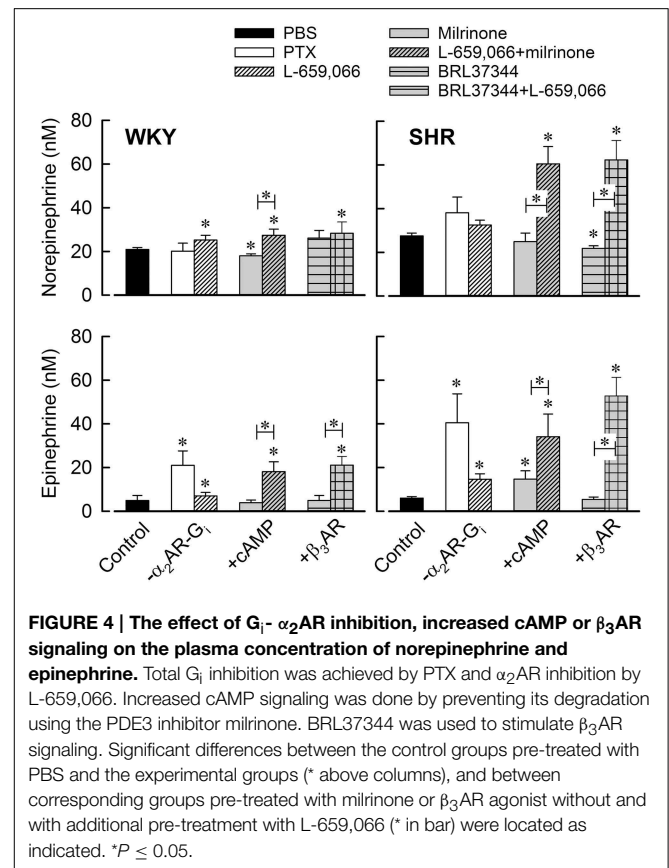
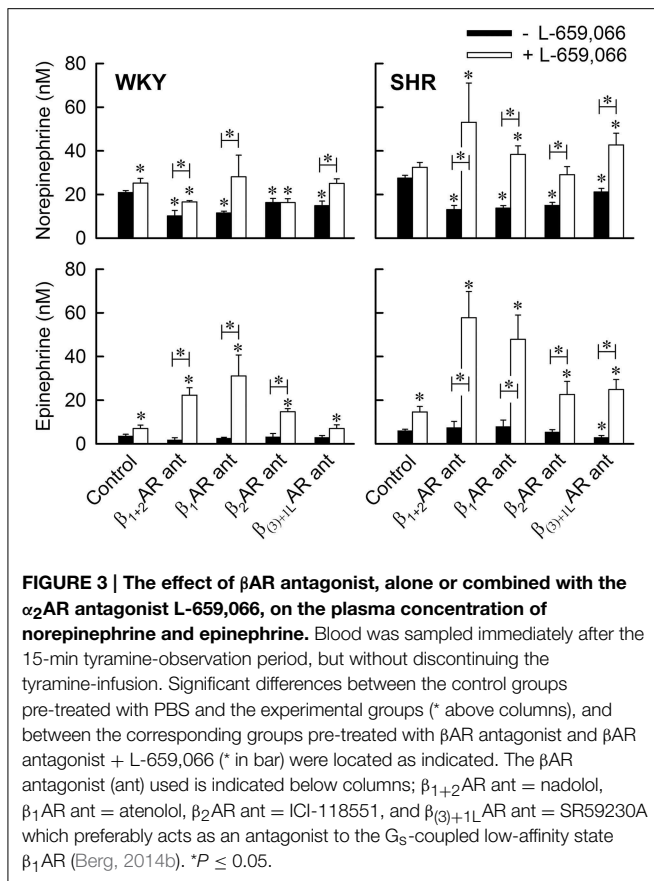
^bThe rats in these groups were in part or in full the same as in (Berg and Jensen, 2013; Berg, 2014a,b). Six to ten rats were included in each group, except for the WKY and SHR controls and the SHR ICI-118551 and L-659,066 groups which comprised 17, 18, 13, and 13 rats, respectively. *, $P \leq 0.05$; [†], $P \leq 0.0036$; [†], [†], $P \leq 0.0083$.

auto inhibition of norepinephrine release was opposed by both β_{1+1L} AR and β_2 AR activity in SHR, with the β_1 AR having the greatest impact. Norepinephrine overflow was also increased after pre-treatment with L-659,066 + milrinone ($P \leq 0.001$ compared to the control, milrinone- or L-659,066-only groups), showing that α_2 AR auto inhibition was enhanced by inhibition of cAMP degradation. In addition, the effect of L-659,066 on overflow was higher when combined with the β_3 AR agonist BRL37344 ($P \leq 0.006$ compared to the controls and after BRL37344 or L-659,066 alone) (Figure 4).

The Effect of PTX, β_{1-3} AR Antagonists, Milrinone, and β_3 AR Agonist on α_2 AR-mediated Inhibition of Epinephrine Secretion

The Effect of Modulating β AR Signaling

The β AR antagonists (Figure 3), milrinone and β_3 AR agonist (Figure 4) had by themselves no effect on the concentration of epinephrine in plasma collected at the end of the experiment in either strain, except for a minor reduction after SR59230A (Figure 3) and an increase after milrinone (Figure 4) in SHR ($P \leq 0.04$).



The Effect of Inhibiting α_2 AR- G_i Signaling (Figure 4)

PTX clearly increased the secretion of epinephrine in both strains ($P \leq 0.032$), demonstrating a tonic G_i -mediated inhibition of epinephrine release. Some increase in the plasma epinephrine concentration was observed in L-659,066-pre-treated WKY ($P = 0.048$) and, in this collection of animals, also in the L-659,066-pre-treated SHR ($P = 0.004$). This observation demonstrated that part of the tonic inhibition of release was due to α_2 AR activity.

The Interaction between α_2 AR and β AR Signaling

However, when L-659,066 was combined with β AR-antagonist (Figure 3), the plasma epinephrine concentration was for all groups in both strains higher than that in the corresponding controls or after each drug alone ($P \leq 0.05$). These results demonstrated that β AR signaling interfered with α_2 AR auto inhibition of epinephrine secretion. The only exceptions were the WKY SR59230A + L-659,066-pre-treated group where the plasma concentration was not different from that in the controls or after each antagonist alone, and the SHR ICI-118551 + L-659,066 group, which was not different from that after L-659,066 alone (Figure 3). A potentiated effect of L-659,066 was also seen when L-659,066 was combined with milrinone or the β_3 AR-agonist BRL37344 (Figure 4).

The Role of β_{1-3} AR and α_2 AR in the Control of Cardiovascular Baselines (Table 1)

As previously documented (Berg et al., 2010; Berg, 2014a), nadolol, atenolol, and ICI-118551 reduced HR in WKY, and MBP, HR and TPR baselines in SHR, although the difference was not statistically significant for all. SR59230A increased MBP and HR in WKY but had no effect in SHR (Berg, 2014b). Milrinone alone had no significant effect on baseline HR or CO, but reduced MBP and TPR in both strains ($P \leq 0.003$ compared to the controls). BRL37344 itself reduced TPR baseline in WKY and increased CO baseline in both strains (Berg, 2014b). Baselines in rats pre-treated with the G_i inhibitor PTX were not significantly different from that in the controls ($P \geq 0.0036$), except for a reduced TPR in WKY. L-659,066 reduced baseline MBP in both strains and TPR significantly in WKY only.

After nadolol/atenolol/ICI-118551 + L-659,066, changes in the cardiovascular baselines in WKY were largely the same as the combined effect of that observed after the β AR and α_2 AR antagonists alone, except for a greater fall in TPR after ICI-118551 + L-659,066 ($P \leq 0.005$ compared to that after each antagonist alone). In SHR, TPR baseline was reduced after atenolol + L-659,066 ($P \leq 0.008$ compared to that after each antagonist alone), whereas the reduction following nadolol alone was not observed after nadolol + L-659,066. The TPR-response to milrinone + L-659,066 was not different from that after milrinone alone in either strain, but was greater than that after

L-659,066 alone in SHR ($P \leq 0.005$). After BRL37344 + L-659,066, the changes seen after BRL37344 or L-659,066 alone remained, but in WKY there was a further reduction in TPR ($P \leq 0.008$ compared to BRL37344 or L-659,066 alone). L-659,066 had no effect on HR or CO baselines in either strain, and did not alter the HR-response to β AR antagonist, milrinone or β_3 AR agonist.

The Influence of β_{1-3} AR and α_2 AR on the TPR-response to Tyramine-stimulated Norepinephrine Release

In agreement with previous studies (Berg et al., 2010; Berg and Jensen, 2013), the tyramine-induced release of norepinephrine activated a transient rise in TPR (Figures 5, 6) and a sustained increase in HR (Figures 7, 8), MBP and CO (not shown) in both strains. Pre-treatment with PTX or L-659,066 eliminated the TPR-response to tyramine in WKY ($P \leq 0.001$ compared to WKY controls, $P = \text{NS}$ for single curve evaluation), and in SHR reduced the TPR-peak response ($P \leq 0.008$), but had no effect on the later response (Figure 5). Milrinone reduced the TPR-peak response in both strains (Figure 5). After pre-treatment with milrinone + L-659,066, the tyramine-induced vasoconstriction was reversed to a vasodilatory response in WKY ($P \leq 0.025$ compared to the control and milrinone-only groups at 3 and 15 min), and eliminated in SHR ($P = \text{NS}$, single curve evaluation) (Figure 5). When the β_3 AR agonist BRL37344 was given

prior to L-659,066, the TPR-response to tyramine was higher than that after L-659,066 alone in WKY ($P = 0.024$ at 15 min) but not different in SHR (Figure 5).

The L-659,066-dependent elimination of the TPR-response to tyramine in WKY was in part reversed by prior administration of nadolol, atenolol or ICI-118551, with a non-additive effect of β_1 - and β_2 AR blockade (Figure 6). Significant differences were not detected between the L-659,066 and the β AR antagonist + L-659,066 pre-treated groups in SHR (Figure 6). When the β_{3+1L} AR antagonist SR59230A was combined with L-659,066, the TPR-response to tyramine was not altered in WKY, whereas the vasoconstriction developed more slowly than after L-659,066 alone in SHR (Figure 6).

The Influence of β_{1-3} AR and α_2 AR on the Tyramine-induced Tachycardia

The tyramine-induced tachycardia was not influenced in either strain by inhibition of α_2 AR- G_i -signaling by L-659,066 or PTX, or by inhibition of cAMP-degradation by milrinone or stimulation of β_3 AR with BRL37344, either alone or combined with L-659,066 (Figure 7). The only exception was milrinone + L-659,066 which halved the HR-response in SHR ($P \leq 0.028$) (Figure 7). As after nadolol alone (Berg et al., 2010), nadolol + L-659,066 eliminated the tachycardia in both strains (Figure 8). Δ HR was clearly reduced after atenolol + L-659,066 and to some extent also after SR59230A + L-659,066 in both strains

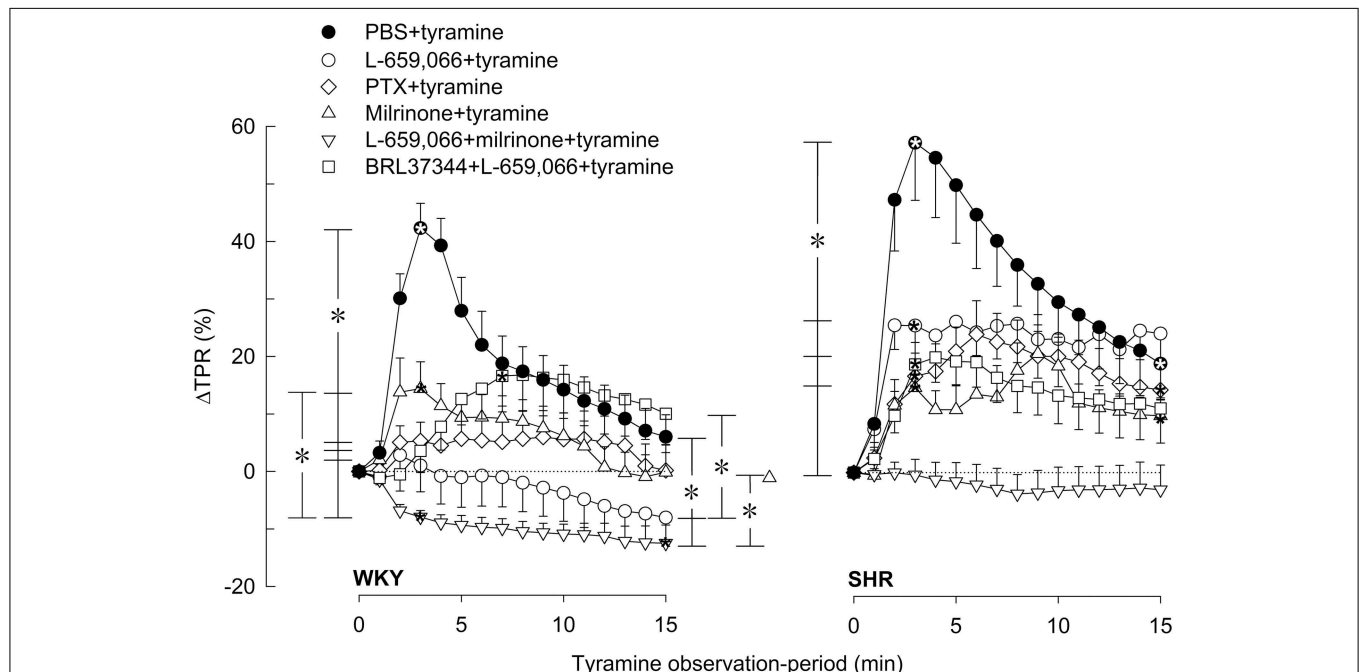
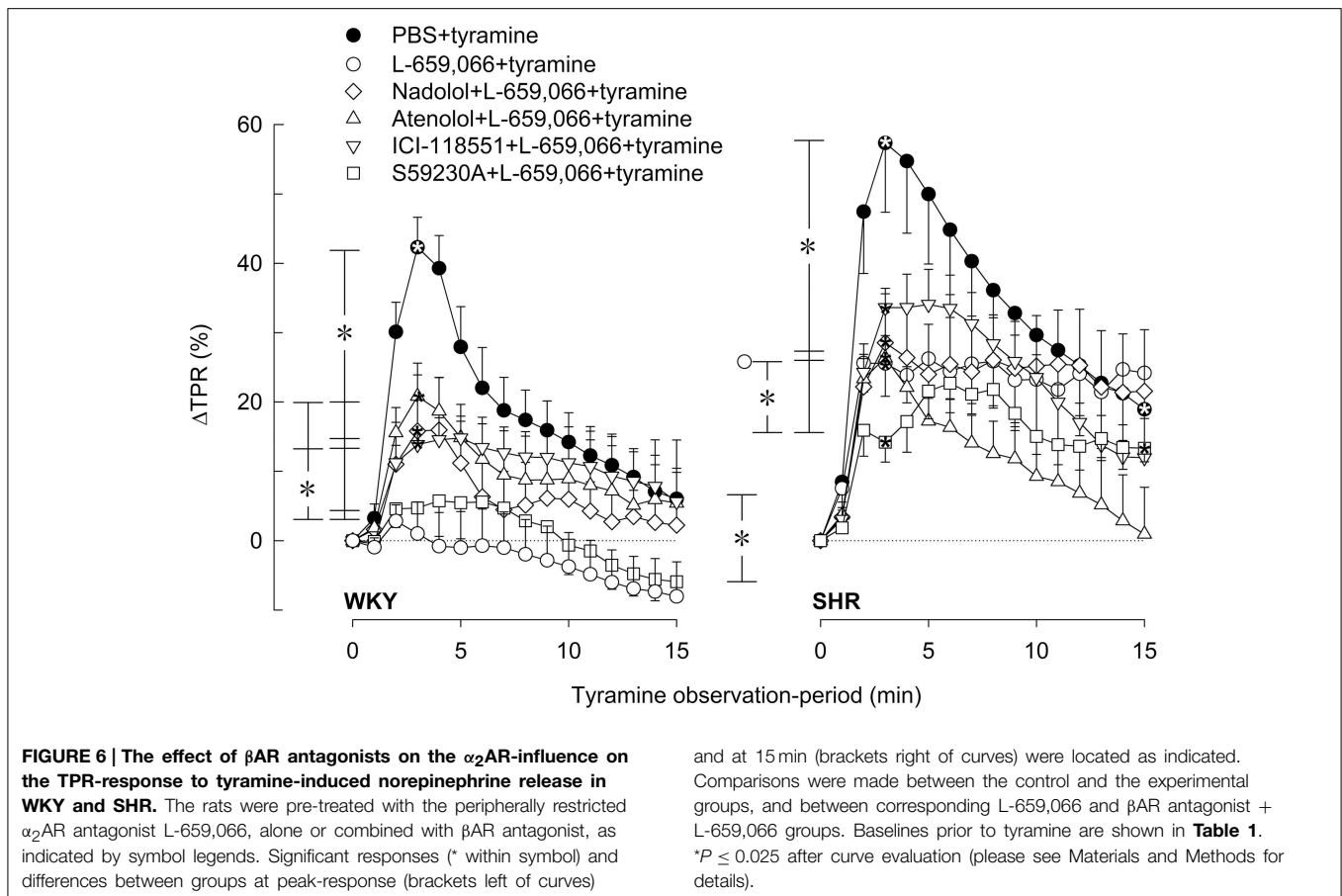


FIGURE 5 | β AR and α_2 AR influence on the TPR-response to tyramine-induced norepinephrine release in WKY and SHR. The rats were pre-treated as indicated by symbol legends. Significant responses (* within symbol) and differences between groups at peak-response (brackets left of curves) and at 15 min (brackets right of curves) were located as

indicated. Comparisons were made between the control and the experimental groups, and between corresponding groups pre-treated with L-659,066 and milrinone + L-659,066. Baselines prior to tyramine are shown in Table 1. * $P \leq 0.025$ after curve evaluation (please see Materials and Methods for details).



(**Figure 8**), not different from that previously observed after atenolol and SR59230A alone (Berg et al., 2010). Different from that previously documented for ICI-118551 alone, i.e., no effect on the HR-response to tyramine in WKY and a slightly reduced response in SHR (Berg et al., 2010), ICI-118551 + L-659,066 reduced the tachycardia in WKY, but slightly increased the response in SHR (**Figure 8**). The effect of these drugs on the HR-response to tyramine was not paralleled by similar changes in the plasma catecholamine concentrations.

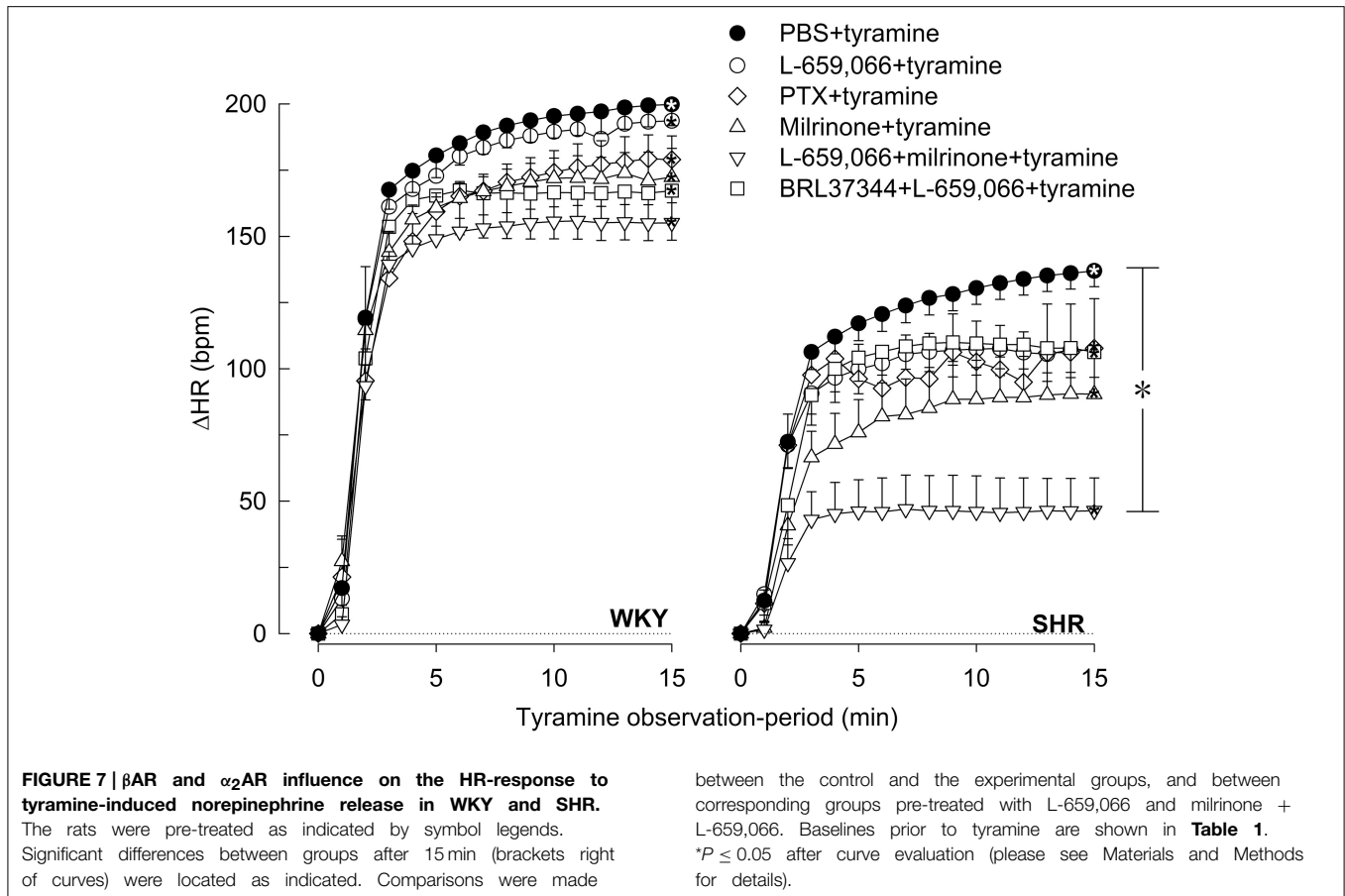
Discussion

The main results in the present study were: (1) The α_2 AR antagonist L-659,066 required the presence of β_2 AR activity to enhance tyramine-induced norepinephrine overflow in WKY but was independent of β_1 AR signaling. (2) α_2 AR-mediated auto inhibition of norepinephrine release in SHR and epinephrine secretion in both strains was opposed primarily by β_{1+1L} AR but also by β_2 AR activity. (3) α_2 AR and β_{1+2} AR reciprocally modulated the TPR-response to the released norepinephrine in WKY but not in SHR. (4) In the presence of L-659,066, β_3 AR agonist stimulated vasoconstriction in WKY and α_2 AR auto inhibition in SHR during tyramine-induced norepinephrine release.

The reduction in tyramine-induced norepinephrine overflow to plasma after the β_1 AR antagonist atenolol was eliminated by

additional pre-treatment with L-659,066 in WKY. The same was observed with the β_{3+1L} AR antagonist SR59230A, most likely due to its ability to inhibit β_{1L} AR (Berg, 2014b) (**Figure 1**). L-659,066 therefore increased release independent of β_{1+1L} AR activity. However, the reduction induced by β_2 AR blocking antagonist, i.e., nadolol or ICI-118551, was somewhat or not at all reversed by L-659,066, respectively. It therefore appeared that β_2 AR-induced stimulation of release was a required substrate for α_2 AR auto inhibition in WKY. Since L-659,066 alone increased the tyramine-induced norepinephrine overflow in WKY, the β_2 AR were evidently active in stimulating release. However, amplifying the response to β_2 AR- G_s signaling by inhibiting PDE3-induced cAMP degradation with milrinone, did not enhance the effect of L-659,066 in WKY. Also the G_i inhibitor PTX did not alter norepinephrine overflow. These results may suggest that α_2 AR- G_i inhibition of norepinephrine release was activated in balance with β_2 AR- G_s stimulation of release. This balance may possibly result from a cAMP-PKA-dependent switch in β_2 AR-signaling from G_s to G_i (Daaka et al., 1997) when cAMP reached a certain level.

In SHR, where L-659,066 did not increase tyramine-induced norepinephrine overflow, additional pre-treatment with β_1 - as well as β_2 AR blockade allowed L-659,066 to increase the plasma norepinephrine concentration. Also SR59230A + L-659,066 increased norepinephrine overflow, most likely due

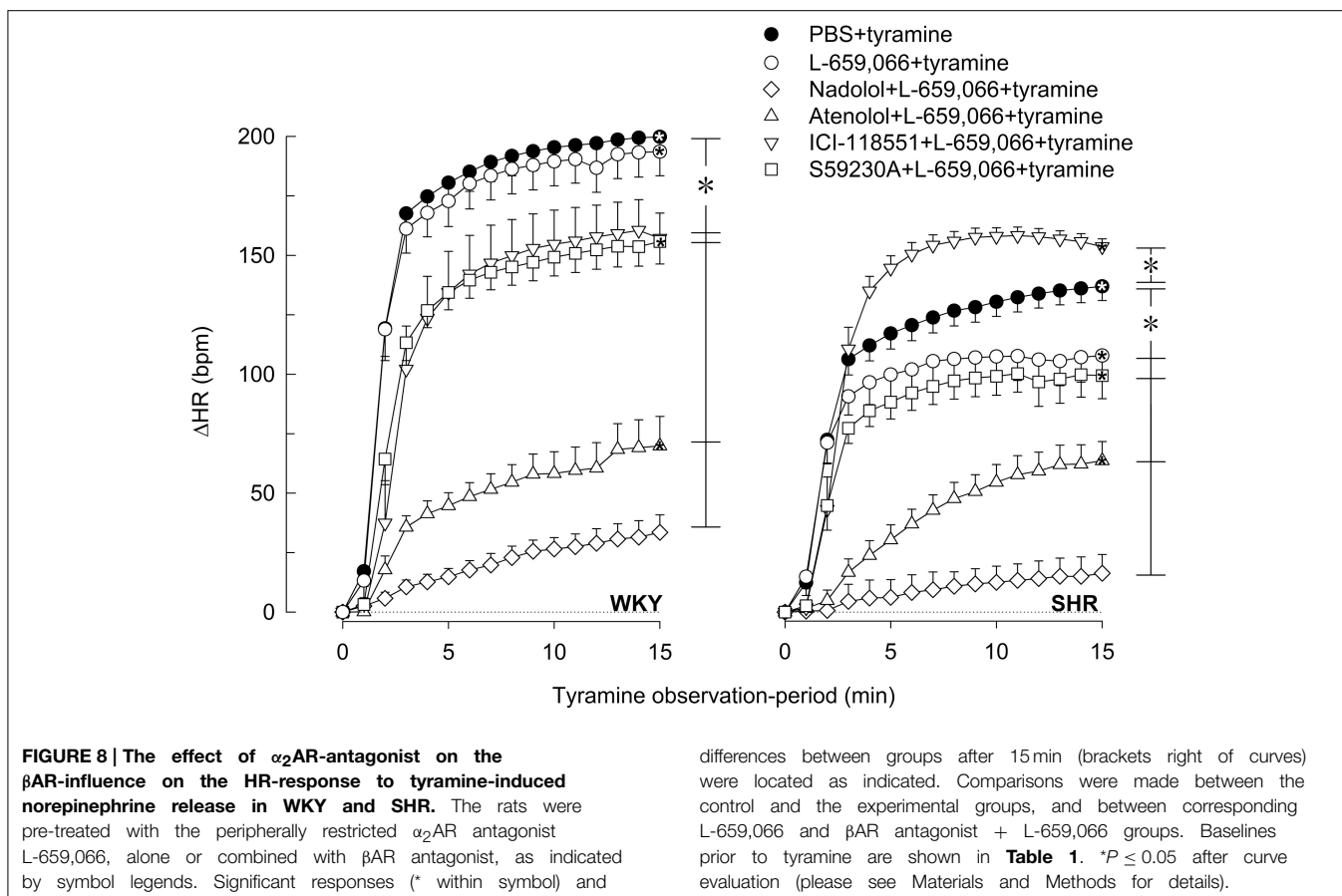


to inhibition of the release-stimulating β_{1L} AR. Thus, both β_1 and β_{1L} AR and β_2 AR counter-acted α_2 AR-mediated inhibition of norepinephrine release in SHR, different from that in WKY. It therefore appeared that excessive β_{1+2} AR activity interfered with the inhibitory effect of α_2 AR- G_i on adenylyl cyclase in this strain.

Surprisingly, L-659,066 combined with the PDE3 inhibitor milrinone doubled norepinephrine release in SHR, even though milrinone alone had no effect. Thus, accumulation of cAMP after preventing its degradation greatly enhanced release only when the inhibitory action of α_2 AR- G_i on adenylyl cyclase was prevented (**Figure 1**). The fact that this was observed in SHR only was compatible with the augmented β AR hampering of α_2 AR auto inhibition in this strain.

Augmented norepinephrine overflow was also observed in SHR when L-659,066 was combined with BRL37344. Since BRL37344 may also activate β_{1+2} AR- G_s signaling (Cernecka et al., 2014), this observation may be explained by a direct stimulating effect on adenylyl cyclase and subsequent norepinephrine release. However, as will be discussed below, BRL37344 in the presence of L-659,066 induced vasoconstriction in WKY, compatible with a stimulating effect on the G_i -coupled β_3 AR. The potentiating effect of BRL37344 on release in SHR may therefore also result from β_3 AR- G_i stimulation, strengthening that activated by α_2 AR (**Figure 1**).

The secretion of epinephrine was tonically down-regulated by G_i in both WKY and SHR, indicated by the 6–7 times increase in the plasma epinephrine concentration in PTX-treated rats. A similar increase was seen in rats without tyramine-stimulated norepinephrine release (Berg et al., 2012). Although the plasma epinephrine concentration after pre-treatment with L-659,066 was less than that after PTX, the difference was not statistically significant, suggesting α_2 AR auto-inhibition of epinephrine secretion to be an important although not the only regulator of G_i in these cells. The secretion of epinephrine was not reduced by milrinone or $\beta_{1/1L/2}$ AR blockade alone. It therefore appeared that α_2 AR tonically inhibited the adrenal secretion of epinephrine in both strains with little β AR influence. In spite of this, $\beta_{1/2}$ AR blockade in both strains and also β_{1L} AR antagonist in SHR potentiated the effect of L-659,066 on epinephrine secretion, apparently with a greater effect of the β_1 - than the β_2 AR in both strains. Thus, different from that observed for norepinephrine release in WKY but similar to that in SHR, β_1 - and β_2 AR in both strains and also β_{1L} AR in SHR opposed α_2 AR-mediated inhibition of the secretion of epinephrine. Like tyramine-stimulated norepinephrine release in SHR but not in WKY, the effect of L-659,066 on the secretion of epinephrine was further enhanced when combined with milrinone or BRL37344 in both strains, most likely through the same mechanisms as discussed above.



The same receptors which presynaptically control catecholamine release are also located postsynaptically and modulate vascular tension and heart rate. Tyramine induces a massive release of norepinephrine without the normal physiological termination of the response by synaptic norepinephrine re-uptake through NET. One may therefore expect the concentration of norepinephrine within the synapse to be more than sufficient to maximally stimulate the postsynaptic receptors in all groups even though norepinephrine release differed. Differences in the cardiovascular response were therefore likely to be primarily due to drug influence on the postsynaptic receptors rather than reflect differences in release due to drug effect on the presynaptic receptors. This conclusion was supported by the fact that the tyramine-induced tachycardia was clearly hampered by β_1 - and β_2 AR antagonists also in the presence of L-659,066 which greatly increased the level of circulating catecholamines. The effect of tyramine does not depend on neuronal action potentials and is therefore not directly influenced by differences in neuronal activity. Due to the anesthesia, the cardiovascular response to tyramine was not modified by activation of baroreflexes, demonstrated by that the HR-response to tyramine was not influenced by atropine (Berg and Jensen, 2013). Moreover, large changes in BP induced by bradykinin or phenylephrine had no effect on HR in similarly anesthetized rats of both strains (Bjørnstad-Østensen and Berg, 1994; Berg et al., 2012).

Norepinephrine release was also not much influenced by the ganglion blocker hexamethonium, but being a nicotine receptor antagonist; it clearly reduced epinephrine secretion in both strains (Berg, 2014a). The vasoconstrictory TPR-response to the tyramine-induced norepinephrine release was due to α_1 AR activation since it was totally abolished by prazosin (Berg et al., 2010). However, concomitant vascular α_2 AR- β AR-activation will modulate this response, and this modulation differed in the two strains. In WKY, the norepinephrine-induced vasoconstriction was totally eliminated by PTX and L-659,066, showing that α_2 AR- G_i -signaling was a major preserver of the α_1 AR-mediated vasoconstriction in this strain. This support was due to that α_2 AR- G_i -signaling opposed β_1 - and β_2 AR-mediated vasodilatation, indicated by that β_1 - or β_2 AR antagonist prevented in part the L-659,066-dependent elimination of TPR-response to tyramine in WKY. Furthermore, accumulation of cAMP after pre-treatment with milrinone clearly reduced the peak-response to tyramine, and, in addition, potentiated the effect of L-659,066, thus precipitating a vasodilatory response to tyramine in milrinone + L-659,066-treated WKY. SR59230A did not alter the TPR-response to tyramine in WKY (Berg, 2014b) or in L-659,066-treated WKY, showing that $\beta_{3/1L}$ AR were not active and did not influence this response. Thus, when α_2 AR-mediated inhibition of adenylyl cyclase was prevented by L-659,066, an increased β_{1+2} AR-dependent vasodilatation in response to the

released norepinephrine and/or epinephrine was allowed. When in addition the degradation of cAMP was blocked, this effect was further enhanced (Figure 2). Thus, α_2 AR- β AR-modulation of the α_1 AR-mediated vasoconstriction was clearly functional in WKY.

In SHR, PTX, and L-659,066 reduced the TPR-peak-response but not the later response to tyramine, but, different from that in WKY, the TPR-response was not eliminated, and was not enhanced by additional pre-treatment with $\beta_{1,1L,2}$ AR antagonist. The slight delay observed in the development of the TPR-response in SHR pre-treated with SR59230A + L-659,066, may possibly result from inhibition of β_3 AR- G_i -signaling. Thus, under the present conditions, there was little interaction between α_2 AR and β AR in the control of vascular tension in SHR. In pathophysiological conditions, including hypertension, enhanced activation of the phospholipase C (PLC)-protein kinase C pathway may lead to inhibition of vasodilatory voltage-sensitive K^+ channels (K_V) (Ko et al., 2010) (Figure 2). The presence of such inhibition in SHR was in fact confirmed by that antagonists against α_1 AR or angiotensin AT₁ and ET_A receptors, which all activate PLC, enhanced the acute vasoconstrictory TPR-response to the K_V inhibitor 4-aminopyridine in SHR but not WKY (Berg, 2003). Since cAMP-induced vasodilatation may be mediated through a protein kinase A (PKA)-dependent opening of K_V (Aiello et al., 1998), the PLC-dependent inhibition of K_V in SHR was therefore likely to interfere with the α_2 AR- G_i/β AR- G_s -cAMP control of vascular tension (Figure 2). A PLC-dependent inhibition of K_V in SHR may therefore explain the absence of α_2 AR- and β AR-modulation of the TPR-response to tyramine-stimulated norepinephrine release in this strain. However, when α_2 AR- G_i -signaling was prevented by L-659,066, and, at the same time, cAMP-signaling was amplified by milrinone, cAMP-mediated vasodilatation dominated the vascular tension response also in SHR. Thus, milrinone + L-659,066 eliminated tyramine-induced vasoconstriction in SHR. However, the effect was still less than that in WKY where milrinone + L-659,066 precipitated a tyramine-induced vasodilatation.

When the β_3 AR were stimulated with the agonist BRL37344 in WKY, the inhibitory effect of L-659,066 on the TPR-response to tyramine was reversed, with a stronger effect as catecholamine release progressed. This effect of BRL37344 could not be explained by its weak β_{1+2} AR agonistic effect (Dolan et al., 1994), and BRL37344 did not interact with the putative β_4 AR (Malinowska and Schlicker, 1997), later identified as the β_{1L} AR (Granneman, 2001; Kaumann et al., 2001). Since the β_3 AR is more resistant to catecholamine-induced desensitization than $\beta_{1/2}$ AR in human tissue (Wallukat, 2002; Rouget et al., 2004),

this subtype may play a more prominent role during prolonged, high levels of norepinephrine such as during the late part of the tyramine-infusion period, particularly when combined with selective agonist. This vasoconstrictory component was likely to be mediated through β_3 AR- G_i signaling.

The tyramine-induced tachycardia was reduced after β_{1-} , β_{2-} , and $\beta_{1L(3)}$ AR antagonist in WKY also in the presence of L-659,066, apparently due to inhibition of postsynaptic β AR, independent of changes in norepinephrine release. In SHR, the tachycardia was reduced after L-659,066, halved after milrinone + L-659,066 and slightly increased after ICI-118551 + L-659,066. The reason for these changes was not obvious, but may result from receptor desensitization in the two former groups, and a possible switch from G_s to G_i for the β_2 AR (Daaka et al., 1997) in the latter group.

Conclusions

α_2 AR-mediated inhibition of norepinephrine release required the presence of β_2 AR in WKY, but was independent of β_1 AR activity. The balanced α_2 AR- β_2 AR interaction in WKY may function to prevent excessive norepinephrine release during physiological conditions with increased epinephrine secretion such as hypoglycemia and exercise, since epinephrine is a better agonist for the β_2 AR subtype than norepinephrine. In SHR, α_2 AR inhibition of norepinephrine release was counteracted by β_1 AR and β_2 AR activity, with an apparently stronger effect of the former. Although an α_2 AR- G_i tonic inhibition dominated the control of epinephrine secretion in both strains, their function was counteracted by β_2 AR and even more by β_1 AR in both strains. The more prominent role of β_1 AR in counter-acting α_2 AR auto inhibition of catecholamine release in SHR may explain why β_1 AR blockers are useful as antihypertensive medication and protective in myocardial infarction and heart failure. The α_1 AR-mediated, vasoconstrictory TPR-response during tyramine-stimulated norepinephrine release was modulated by α_2 AR and $\beta_{1/2}$ AR in WKY. The latter interaction was not functional in SHR, most likely due to a PLC-dependent, reduced K_V vasodilatory influence on VSMC tension, a substrate for cAMP-induced vasodilatation.

Funding

The Norwegian Council on Cardiovascular Diseases and Anders Jahres Fond.

References

- Aiello, E. A., Malcolm, A. T., Walsh, M. P., and Cole, W. C. (1998). Beta-adrenoceptor activation and PKA regulate delayed rectifier K^+ channels of vascular smooth muscle cells. *Am. J. Physiol.* 275, H448–H459.
- Anand-Srivastava, M. B., Srivastava, A. K., and Cantin, M. (1987). Pertussis toxin attenuates atrial natriuretic factor-mediated inhibition of adenylate cyclase. Involvement of inhibitory guanine nucleotide regulatory protein. *J. Biol. Chem.* 262, 4931–4934.
- Berg, T. (2003). The vascular response to the K^+ channel inhibitor 4-aminopyridine in hypertensive rats. *Eur. J. Pharmacol.* 466, 301–310. doi: 10.1016/S0014-2999(03)01555-3
- Berg, T. (2005). Increased counteracting effect of eNOS and nNOS on an alpha(1)-adrenergic rise in total peripheral vascular resistance in spontaneous hypertensive rats. *Cardiovasc. Res.* 67, 736–744. doi: 10.1016/j.cardiores.2005.04.006
- Berg, T. (2013). Angiotensin AT₁ – alpha2Adrenoceptor interaction disturbs alpha2A-autoinhibition of catecholamine release in hypertensive rats. *Front. Neurol.* 4:70. doi: 10.3389/fneur.2013.00070

- Berg, T. (2014a). Beta1-blockers lower norepinephrine release by inhibiting presynaptic, facilitating beta1-adrenoceptors in normotensive and hypertensive rats. *Front. Neurol.* 5:51. doi: 10.3389/fneur.2014.00051
- Berg, T. (2014b). Beta3-adrenoceptors inhibit stimulated norepinephrine release in spontaneously hypertensive rats. *Front. Physiol.* 5:499. doi: 10.3389/fphys.2014.00499
- Berg, T., Degerman, E., and Tasken, K. (2009). Increased cAMP signaling can ameliorate the hypertensive condition in spontaneously hypertensive rats. *J. Vasc. Res.* 46, 25–35. doi: 10.1159/000135662
- Berg, T., and Jensen, J. (2011). Simultaneous parasympathetic and sympathetic activation reveals altered autonomic control of heart rate, vascular tension, and epinephrine release in anesthetized hypertensive rats. *Front. Neurol.* 2:71. doi: 10.3389/fneur.2011.00071
- Berg, T., and Jensen, J. (2013). Tyramine reveals failing alpha2-adrenoceptor control of catecholamine release and total peripheral vascular resistance in hypertensive rats. *Front. Neurol.* 4:19. doi: 10.3389/fneur.2013.00019
- Berg, T., Piercey, B. W., and Jensen, J. (2010). Role of beta1-3-adrenoceptors in blood pressure control at rest and during tyramine-induced norepinephrine release in spontaneously hypertensive rats. *Hypertension* 55, 1224–1230. doi: 10.1161/HYPERTENSIONAHA.109.149286
- Berg, T., Walaas, S. I., Roberg, B. A., Huynh, T. T., and Jensen, J. (2012). Plasma norepinephrine in hypertensive rats reflects alpha(2)-adrenoceptor release control only when re-uptake is inhibited. *Front. Neurol.* 3:160. doi: 10.3389/fneur.2012.00160
- Bjørnstad-Østensen, A., and Berg, T. (1994). The role of nitric oxide, adrenergic activation and kinin-degradation in blood pressure homeostasis following an acute kinin-induced hypotension. *Br. J. Pharmacol.* 113, 1567–1573. doi: 10.1111/j.1476-5381.1994.tb17175.x
- Brede, M., Nagy, G., Philipp, M., Sorensen, J. B., Lohse, M. J., and Hein, L. (2003). Differential control of adrenal and sympathetic catecholamine release by alpha 2-adrenoceptor subtypes. *Mol. Endocrinol.* 17, 1640–1646. doi: 10.1210/me.2003-0035
- Cernecka, H., Sand, C., and Michel, M. C. (2014). The odd sibling: features of beta3-adrenoceptor pharmacology. *Mol. Pharmacol.* 80, 479–484. doi: 10.1124/mol.114.092817
- Clineschmidt, B. V., Pettibone, D. J., Lotti, V. J., Hucker, H. B., Sweeney, B. M., Reiss, D. R., et al. (1988). A peripherally acting alpha-2 adrenoceptor antagonist: L-659,066. *J. Pharmacol. Exp. Ther.* 245, 32–40.
- Daaka, Y., Luttrell, L. M., and Lefkowitz, R. J. (1997). Switching of the coupling of the beta2-adrenergic receptor to different G proteins by protein kinase A. *Nature* 390, 88–91. doi: 10.1038/36362
- Dolan, J. A., Muenkel, H. A., Burns, M. G., Pellegrino, S. M., Fraser, C. M., Pietri, F., et al. (1994). Beta-3 adrenoceptor selectivity of the dioxolane dicarboxylate phenethanolamines. *J. Pharmacol. Exp. Ther.* 269, 1000–1006.
- Gauthier, C., Leblais, V., Kobzik, L., Trochu, J. N., Khandoudi, N., Bril, A., et al. (1998). The negative inotropic effect of beta3-adrenoceptor stimulation is mediated by activation of a nitric oxide synthase pathway in human ventricle. *J. Clin. Invest.* 102, 1377–1384. doi: 10.1172/JCI12191
- Granneman, J. G. (2001). The putative beta4-adrenergic receptor is a novel state of the beta1-adrenergic receptor. *Am. J. Physiol. Endocrinol. Metab.* 280, E199–E202.
- Kaumann, A. J., Engelhardt, S., Hein, L., Molenaar, P., and Lohse, M. (2001). Abolition of (-)-CGP 12177-evoked cardiostimulation in double beta1/beta2-adrenoceptor knockout mice. Obligatory role of beta1-adrenoceptors for putative beta4-adrenoceptor pharmacology. *Naunyn Schmiedeberg's Arch. Pharmacol.* 363, 87–93. doi: 10.1007/s002100000336
- Ko, E. A., Park, W. S., Firth, A. L., Kim, N., Yuan, J. X., and Han, J. (2010). Pathophysiology of voltage-gated K⁺ channels in vascular smooth muscle cells: modulation by protein kinases. *Prog. Biophys. Mol. Biol.* 103, 95–101. doi: 10.1016/j.pbiomolbio.2009.10.001
- Makaritsis, K. P., Johns, C., Gavras, I., Altman, J. D., Handy, D. E., Bresnahan, M. R., et al. (1999). Sympathoinhibitory function of the alpha(2A)-adrenergic receptor subtype. *Hypertension* 34, 403–407. doi: 10.1161/01.HYP.34.3.403
- Malinowska, B., and Schlicker, E. (1997). Further evidence for differences between cardiac atypical beta-adrenoceptors and brown adipose tissue beta3-adrenoceptors in the pithed rat. *Br. J. Pharmacol.* 122, 1307–1314. doi: 10.1038/sj.bjp.0701516
- Malle, M. Y., Toumaniantz, G., Serpillon, S., Gautier, F., Gogny, M., Desfontis, J. C., et al. (2004). Impairment of the low-affinity state beta1-adrenoceptor-induced relaxation in spontaneously hypertensive rats. *Br. J. Pharmacol.* 143, 599–605. doi: 10.1038/sj.bjp.0705990
- Philipp, M., Brede, M., and Hein, L. (2002). Physiological significance of alpha(2)-adrenergic receptor subtype diversity: one receptor is not enough. *Am. J. Physiol. Regul. Integr. Comp. Physiol.* 283, R287–R295. doi: 10.1152/ajpregu.00123.2002
- Queen, L. R., Ji, Y., Xu, B., Young, L., Yao, K., Wyatt, A. W., et al. (2006). Mechanisms underlying beta2-adrenoceptor-mediated nitric oxide generation by human umbilical vein endothelial cells. *J. Physiol.* 576, 585–594. doi: 10.1113/jphysiol.2006.115998
- Rouget, C., Breuille-Fouche, M., Mercier, F. J., Leroy, M. J., Loustalot, C., Naline, E., et al. (2004). The human near-term myometrial beta 3-adrenoceptor but not the beta 2-adrenoceptor is resistant to desensitisation after sustained agonist stimulation. *Br. J. Pharmacol.* 141, 831–841. doi: 10.1038/sj.bjp.0705616
- Shafaroudi, M. M., McBride, M., Deighan, C., Wokoma, A., Macmillan, J., Daly, C. J., et al. (2005). Two “knockout” mouse models demonstrate that aortic vasodilatation is mediated via alpha2a-adrenoceptors located on the endothelium. *J. Pharmacol. Exp. Ther.* 314, 804–810. doi: 10.1124/jpet.105.085944
- Stjerne, L., and Brundin, J. (1976). Beta2-adrenoceptors facilitating noradrenaline secretion from human vasoconstrictor nerves. *Acta Physiol. Scand.* 97, 88–93. doi: 10.1111/j.1748-1716.1976.tb10238.x
- Trendelenburg, A. U., Philipp, M., Meyer, A., Klebroff, W., Hein, L., and Starke, K. (2003). All three alpha2-adrenoceptor types serve as autoreceptors in postganglionic sympathetic neurons. *Naunyn Schmiedeberg's Arch. Pharmacol.* 368, 504–512. doi: 10.1007/s00210-003-0829-x
- Wallukat, G. (2002). The beta-adrenergic receptors. *Herz* 27, 683–690. doi: 10.1007/s00059-002-2434-z
- Westfall, T. C., Peach, M. J., and Tittermary, V. (1979). Enhancement of the electrically induced release of norepinephrine from the rat portal vein: mediation by beta 2-adrenoceptors. *Eur. J. Pharmacol.* 58, 67–74. doi: 10.1016/0014-2999(79)90341-8

Conflict of Interest Statement: The author declares that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2015 Berg. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) or licensor are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.