# <sup>13</sup>C-LABELLED BREATH TESTS IN MALABSORPTION

# STUDIES OF PATIENTS WITH COELIAC DISEASE

# **Kari Tveito**





**Faculty of Medicine** 

University of Oslo

2011

# © Kari Tveito, 2011

Series of dissertations submitted to the Faculty of Medicine, University of Oslo No. 1169

ISBN 978-82-8264-157-9

All rights reserved. No part of this publication may be reproduced or transmitted, in any form or by any means, without permission.

Cover: Inger Sandved Anfinsen. Printed in Norway: AIT Oslo AS.

Produced in co-operation with Unipub.

The thesis is produced by Unipub merely in connection with the thesis defence. Kindly direct all inquiries regarding the thesis to the copyright holder or the unit which grants the doctorate.

#### ACKNOWLEDGEMENTS

The work presented in this thesis was carried out at the section for Gastroenterology,
Lovisenberg Diakonale Hospital, in collaboration with the Institute of Internal Medicine,
Rikshospitalet University Hospital, the Department of Pathology, Oslo University
Hospital, Ullevål, and the section for Gastroenterology, Department of Medicine,
Stavanger University Hospital. Financial support from the South-Eastern Norway
Regional Health Authority and Lovisenberg Diakonale Hospital made it possible to
perform the studies. I am most grateful for this funding.

First of all I want to express my deepest gratitude to my supervisor Viggo Skar for brilliant supervision and continuous support and enthusiasm during the work of this thesis. Thank you for believing in me and giving me the opportunity to become a research fellow.

I also want to thank statistician Cathrine Brunborg and co-supervisor professor Leiv Sandvik for weekly meetings, invaluable discussions and guidance through these years. I could not have carried out the mathematical analyses without your excellent help.

I am very grateful for support from co-supervisors Knut E.A. Lundin and professor Ludvig Sollid who shared of their cutting-edge knowledge about coeliac disease and provided important advice for the projects.

A special thanks goes to co-author Else Marit Løberg who went through the duodenal biopsies and provided expert judgments on the histological pathology. This part of the work turned out to be very important for interpretations of our breath analysis results. I am very grateful for the work that Jorunn Bratlie carried out in analysing xylose concentrations in blood and urine samples from our patients. Without her excellent contribution, we would not have been able to compare the breath test technology with traditional xylose malabsorption tests.

We were indeed very lucky to be given the opportunity to collaborate with the section of Gastroenterology at Stavanger University Hospital who enrolled patients in the sorbitol studies. A special thanks goes to co-author Anne Kristine Hetta who meticulously recruited patients and gathered clinical, histological and biochemical data.

This study would not have been carried out without the financial, moral and administrative support from the Lovisenberg Diakonale Hospital. I am very grateful for the generous support from clinical chiefs at the Department of Medicine, Anne Marit Tangen and Per Gerlyng. A special thanks goes to chief doctor Bjørn Holm who provided the most important support and encouragement for the PhD candidates in our group throughout these years. We are very grateful that he made it possible to include a research nurse in the group. Mia Askedal turned out to be perfect for this job. She became my most important collaborator and friend during my PhD period, and assisted in all sorts of work from organizing patient controls, follow-up, breath test analysis, administration and help in reading through the papers in progress. Thank you, Mia, you were the best work-

partner I could have wished for. I also want to thank other members of the group, especially Håvar Hope, for a great working environment, instructive and fun meetings.

A warm thanks goes to the staff at the section of Gastroenterology who never said no to helping with project-related tasks. I am especially grateful to have worked with such wonderfully competent nurses and secretaries in the outpatient clinic who showed outstanding interest in the project and assisted greatly in many ways. I also want to thank all the colleagues at the Department of Medicine at Lovisenberg Diakonale Hospital.

A special thanks goes also to Oslo University Hospital, Aker, chief of the section of Gastroenterology at the Department of Medicine, Jørgen Jahnsen, and professor in Gastroenterology Bjørn Moum who generously supported me, gave me the opportunity to finish my last papers in the thesis, and also continue my research in the Department of Medicine, Aker, under their supervision.

Finally, I would like to express my gratitude to family and friends for always being there. A special thanks goes to Sidharth Mishra and Sebastian Straube who also provided invaluable research advice. My deepest gratitude goes to my parents for their continuous love and support. A very special thanks goes to my sisters, Marit and Anne, my dearest and best friends of all.

Oslo, February 2011

Kari Tveito

# TABLE OF CONTENTS

Ackno	cknowledgements			
Table of contents				
1 Abstract				
2 Abbreviations				
3 Introduction				
3.1 Coeliac disease			11	
	3.1.1	Genes and environment	11	
	3.1.2	Pathophysiology	12	
	3.1.3	Epidemiology	12	
	3.1.4	Clinical aspects	13	
3.2. Breath analysis in gastroenterology			14	
	3.2.1	<sup>4</sup> C- and <sup>13</sup> C-breath tests	14	
	3.2.2 I	Hydrogen breath tests	15	
	3.3 D-xylose testing in malabsorption			
	3.4 Summary of introduction			
4 Aim of the study				
5 List of Papers			21	
	5.1 Paper I		22	
	5.2 Paper II		23	
	5.3 Paper III			
	5.4 Paper IV		25	

5.5 Other publications during the PhD programme	26	
6 Methods		
6.1 Study subjects	27	
6.1.1 Patients	27	
6.1.2 Healthy control subjects	27	
6.2 Breath test protocols	28	
6.2.1 <sup>14</sup> C-D-xylose breath test	28	
6.2.2 <sup>13</sup> C-D-xylose breath test	29	
6.2.3 <sup>13</sup> C-sorbitol breath test	29	
6.2.4 H <sub>2</sub> -sorbitol breath test	30	
6.3 D-xylose in plasma and urine	30	
6.4 Statistics	31	
6.5 Ethics	31	
7 Discussion		
7.1 Strength and limitations of the studies	32	
7.2 The <sup>13</sup> C-D-xylose breath test and the <sup>13</sup> C-sorbitol breath test	33	
7.3 The H <sub>2</sub> -sorbitol breath test	35	
7.4 Pathophysiological considerations	36	
7.5 Clinical considerations	38	
7.6 Conclusion	39	
8 References		
9 Errata		
10 Original papers		

#### 1 ABSTRACT

Breath tests have been widely used in gastroenterology for several decades. The most well known test nowadays is perhaps the <sup>13</sup>C-urea breath test for Helicobacter Pylori. Isotope-labelled breath test can, however, be applied in a wide range of conditions. The main aim of this thesis has been to determine the usefulness of <sup>13</sup>C-labelled substances to diagnose small intestinal malabsorption. To this end, we have focused on coeliac patients, since coeliac disease is one of the most common causes of small intestinal malabsorption in the industrialised part of the world today.

The main objective of the first study of this thesis was to determine the diagnostic accuracy of the 13C-D-xylose breath test in diagnosis and follow-up of coeliac disease. Two cohorts of coeliac patients were reviewed retrospectively. In one cohort patients had been tested with a 14C-D-xylose breath test before and after gluten-free treatment (GFD). In the other patients had been tested with a 13C-D-xylose breath test before and after GFD. Results were similar in the two cohorts, and confirmed that the non-radioactive isotope <sup>13</sup>C-D-xylose can replace the radioactive isotope <sup>14</sup>C-D-xylose without loss of diagnostic accuracy. Furthermore, we developed a diagnostic index for the D-xylose breath tests in which <sup>13</sup>CO<sub>2</sub> or <sup>14</sup>CO<sub>2</sub> levels in breath at 30 minutes were combined with levels in breath at 210 minutes in a gas fraction index: 30 minutes value/210 minutes value. Using this diagnostic index, we found sensitivities of 84-95% and specificities of 87-94% with the two breath tests. After treatment with a gluten-free diet, most patients improved their gas fraction index, but did not reach the levels of healthy control subjects.

In the second study we compared the <sup>13</sup>C-D-xylose breath test to measurements of D-xylose in plasma and urine in untreated coeliac patients and treated coeliac patients.

Test sensitivity and specificity for coeliac disease versus healthy controls were 88% and 84% with the <sup>13</sup>C-D-xylose breath test, 65% and 71% with a 1-h plasma D-xylose test, and 55% and 74% with a 4-h urine D-xylose excretion test. Breath test results improved significantly in the treated coeliac group compared to untreated patients, but were not normalized compared to healthy control subjects.

The aim of the two last studies was to develop a novel <sup>13</sup>C-sorbitol breath test and determine the diagnostic accuracy of the test for coeliac disease. We performed a prospective study of coeliac patients who were tested at diagnosis before commencing on a gluten-free diet and after approximately one year on GFD. Coeliac patients were compared to a group of patients with similar gastrointestinal symptoms without coeliac disease and to healthy control subjects. All study subjects performed the H<sub>2</sub>-sorbitol breath test concomitant with the <sup>13</sup>C-sorbitol breath test. We found that the <sup>13</sup>C-sorbitol breath test duration could be shortened to one-hour without loss of diagnostic precision versus a four-hour test. We proposed a diagnostic algorithm to divide patients into high, moderate and low-risk groups for coeliac disease. <sup>13</sup>C-sorbitol breath test results increased in treated coeliac patients, but did not reach levels of healthy control subjects. The H<sub>2</sub>-sorbitol breath test had unsatisfactory specificity versus control groups.

We concluded that <sup>13</sup>C-labelled breath tests are sensitive tools for small intestinal malabsorption. Further studies are needed to determine how <sup>13</sup>C-labelled breath tests can be of value in follow-up of coeliac disease.

# **2 ABBREVIATIONS**

<sup>13</sup>C-SBT <sup>13</sup>C-sorbitol breath test

GC gas chromatography

GFD gluten-free diet

GF-index gas fraction index

HLA human leukocyte antigen

HPLC high performance liquid chromatography

H<sub>2</sub>-SBT H<sub>2</sub>-sorbitol breath test

IEL intraepithelial lymphocyte

Ig immunoglobulin

IRMS isotope ratio mass spectrometry

tTGA tissue-transglutaminase antibody

SIBO small intestinal bacterial overgrowth

#### **3 INTRODUCTION**

#### 3.1 Coeliac disease

Coeliac disease is a chronic inflammatory disease of the small intestine induced by ingestion of proline-rich and glutamine-rich gluten proteins in wheat, rye and barley in genetically susceptible people [1]. The first description of the disease dates from the 1<sup>st</sup> and 2<sup>nd</sup> centuries CE [2]. The disorder is characterized by a diverse clinical heterogeneity that ranges from asymptomatic to severely symptomatic. Patients with coeliac disease have increased morbidity due to the frequent association with autoimmune disorders [3-9], and increased mortality resulting from the emergence of T-cell clonal proliferation that predispose the patient to enteropathy-type T-cell lymphoma [10-12]. A strict glutenfree diet is the mainstay of safe and effective treatment.

#### 3.1.1 Genes and environment

Coeliac disease is strongly linked with HLA-DQ genes. Most patients carry a variant of DQ2 and others carry a variant of DQ8 [13]. The association between HLA genes and coeliac disease is very strong compared with other HLA-linked diseases [14]. However, other non-HLA regions must be involved as well, since the concordance rate between HLA identical siblings is much lower than between monozygotic twins who show a concordance rate of 85% [15, 16].

Coeliac disease is induced in genetically susceptible people by gluten and possibly other environmental cofactors. Intestinal infections may be a trigger for coeliac disease, perhaps because an infection causes a transient rise in small-bowel permeability and

release of tissue transglutaminase that enhances gluten immunogenicity [17, 18]. Some drugs can enhance a person's susceptibility to gluten - such as a course of interferon alpha [19]. Furthermore, infant-feeding practices and whether infants are introduced to dietary gluten while still being breastfed might affect the risk of developing coeliac disease [20, 21].

#### 3.1.2 Pathophysiology

After crossing the epithelium into the lamina propria, gluten peptides are deamidated by tissue transglutaminase and presented by DQ2+ or DQ8+ antigen-presenting cells to pathogenic CD4+ T cells. Activated CD4+ T cells drive a T-helper type 1 response that leads to the development of the coeliac lesion. The severity of the lesion - which can range from intraepithelial and lamina propria infiltration of inflammatory cells with normal architecture to crypt hyperplasia and varying degrees of villous atrophy to completely flat mucosa – can be graded by the Marsh classification, as modified by Oberhufer and colleagues [22].

#### 3.1.3 Epidemiology

The accuracy of estimates of the true prevalence of coeliac disease has been substantially improved by the increased reliability of serological tests. The IgA antigliadin antibodies used initially had low sensitivity and specificity [23-25], and the antiendomysial antibody test significantly improved the diagnostic accuracy. An important break-through came with the discovery that antiendomysial antibodies detect tissue transglutaminase [26].

The IgA tissue transglutaminase antibody test used for screening of coeliac disease today has very high sensitivity and specificity for coeliac disease [27, 28]. By screening with serological tests, the prevalence of biopsy-proven coeliac disease in Finnish and Italian schoolchildren was reported to be 1:99 and 1:106, respectively [29, 30]. Similar rates of seroprevalence have been reported in adult populations in UK (1:87) and USA (1:105) [31, 32]. The highest rate of antiendomysial antibody positivity (5.6%) has been reported in Saharawi children [33].

## 3.1.4 Clinical aspects

Until the beginning of the 1980s, the use of intestinal biopsy was reserved for patients with overt malabsorption. Consequently, the prevalence of malabsorption among patients with coeliac disease was very high. As awareness of the disease improved and the threshold for investigation was lowered, more subtle forms of the disease and the variable clinical expression of the condition was acknowledged [1]. At the end of the 1980s- after the advent of serological tests - the number of patients with minor symptoms was twice the number of patients with overt malabsorption. There was a significant rise in the rate of diagnosis, and a reduction of the male to female ratio [34]. Coeliac disease is two to three times more common in women than in men, but this predominance falls after the age of 65 years.

The factors that determine the severity of symptoms at presentation are not known. Neither the degree of duodenal villous atrophy nor the extent of visible enteropathy assessed by capsule endoscopy correlates with clinical presentation [35, 36].

## 3.2 Breath analysis in gastroenterology

The first gastroenterological applications of breath analysis were carried out by the medical student Charles Dodds in 1920 [37, 38]. He reported that the pressure of carbon dioxide (PC0<sub>2</sub>) in breath rose after meals, and that the rise in PCO<sub>2</sub> differed in patients with pernicious anaemia and pancreatic disease from healthy controls. Following these pioneering efforts, the area remained dormant until, in the 1960s, new techniques were developed simplifying the assessment of <sup>14</sup>CO<sub>2</sub> after administration of <sup>14</sup>C-labeled compounds and of hydrogen (H<sub>2</sub>) after administration of carbohydrate [39].

# 3.2.1 14C-breath tests and 13C-breath tests

Recovery of <sup>14</sup>CO<sub>2</sub> in breath after administration of <sup>14</sup>C-labeled compounds indicates that metabolism of the radioactively labelled compound has occurred. When the rate-limiting step in the metabolism of the compound is intestinal absorption, measurement of breath <sup>14</sup>CO<sub>2</sub> output after oral administration of the compound may provide information about it's rate of absorption. If the compound is metabolized solely by intestinal bacteria measurement of breath <sup>14</sup>CO<sub>2</sub> output after oral administration of the compound may provide information about the exposure of the compound to intestinal bacteria.

The original radiospirometric studies applied to gastroenterological studies were performed by Schwabe et al., who administered <sup>14</sup>C-labeled trioctanoate and monitored breath <sup>14</sup>CO<sub>2</sub> [40]. Cozetto el al., in 1964, applied the same technique to study intestinal absorption of fatty acids, glucose and lactose in children [41].

Tests in which breath <sup>14</sup>CO<sub>2</sub> is measured require the administration of small amounts of

<sup>14</sup>C-labeled compounds. The calculated radiation dose is extremely small. Nevertheless,

the tests should not be performed in children or pregnant women. When <sup>13</sup>C-compounds became widely available at a reasonable cost and methods to measure <sup>13</sup>C abundance in samples of CO<sub>2</sub> were developed, <sup>14</sup>C-labeled substrates were gradually replaced by non-radioactive <sup>13</sup>C-labeled substrates [42-45].

The <sup>13</sup>C-urea breath test for the diagnosis of Helicobacter Pylori infection is the most used stable isotope breath test in gastroenterology today [46, 47]. However, a range of substrates and functions can be monitored by means of <sup>13</sup>C-breath tests [48, 49]. The underlying concept of the <sup>13</sup>C-breath test is simple and similar to the <sup>14</sup>C-breath tests: <sup>13</sup>C is introduced into one or more functional groups in a substrate. The functional groups are linked to the rest of the molecule through bonds that are cleaved by specific enzymes. Once cleavage occurs, the functional group is further oxidized until CO<sub>2</sub> is produced and excreted in breath.

In 1978, Toskes et al. developed a new <sup>14</sup>C-D-xylose breath test [50]. In a series of studies they showed that the <sup>14</sup>C-D-xylose breath test was well suited to detect small intestinal bacterial overgrowth (SIBO) [51-53]. The test was later replaced by the <sup>13</sup>C-D-xylose breath test [54, 55]. However, the initial optimism regarding the utility of the <sup>14</sup>C-D-xylose and <sup>13</sup>C-D-xylose breath tests for SIBO has been challenged by several studies showing less convincing results [56-58].

## 3.2.2 Hydrogen breath tests

In 1975, Newcomer et al., demonstrated the superiority of H<sub>2</sub> measurements to <sup>14</sup>CO<sub>2</sub>-labelled lactose and blood sugar changes in detecting lactose malabsorption [59]. The H<sub>2</sub>-lactose breath test has been shown to have high sensitivity for lactose malabsorption in

many repeated studies since then, and is accepted as the method of choice for lactose malabsorption by many gastroenterologists [60-63].

Soon after the introduction of the H<sub>2</sub>-lactose breath test, the H<sub>2</sub>-breath test was also applied to other complex sugars like fructose [64, 65], maltose [66], sucrose [67], sorbitol [68], and lactulose [69, 70]. In 1988, Corazza et al., presented a sorbitol H<sub>2</sub>-breath test for coeliac disease [71]. The initial study as well as follow-up studies demonstrated high sensitivity of the H<sub>2</sub>-sorbitol breath test for untreated coeliac disease [72, 73].

All cells produce CO<sub>2</sub> during metabolism, but only bacteria can produce H<sub>2</sub> and methane (CH<sub>4</sub>) as metabolic by-products. An increase in H<sub>2</sub> concentration in breath after ingestion of a carbohydrate shows that bacterial fermentation of the carbohydrate has taken place in the intestine [74]. It has been shown that a relationship exists between H<sub>2</sub> and CH<sub>4</sub> production, in which methanogenic bacteria are able to convert H<sub>2</sub> to CH<sub>4</sub>, and this exchange occurs in the colon [75, 76]. A significant proportion of humans have a methanogenic gut bacterial flora [77-80]. Therefore, when intestinal bacteria metabolize carbohydrates, sometimes only H<sub>2</sub> is produced, sometimes both H<sub>2</sub> and CH<sub>4</sub> appears in the exhaled air, and sometimes only CH<sub>4</sub> will be increased. Methane should also be measured in breath tests for carbohydrate malabsorption in order to allow correct interpretation of the test. The response to a challenge-dose of sugar is measured from the pre-dose baseline or the lowest level reached prior to an increase.

#### 3.3 D-xylose testing in malabsorption

Absorption of D-xylose has been a gold standard test in the evaluation of small intestinal malabsorption since the 1960s [81, 82]. However, the diagnostic accuracy of the test has been subject to much controversy in later years [83-87]. D-xylose is a five carbon monosaccharide found naturally in plants [88]. The sugar is incompletely absorbed, which allows it to be used as an absorptive test [89]. The mechanism of D-xylose absorption in the small intestine is disputed. Data from intestinal perfusion studies in normal humans support predominantly passive absorption, transcellular or paracellular, with little or no carrier-mediated transport [90]. D-xylose is absorbed unchanged from the small intestine [91, 92]. Approximately 30% is metabolized by the liver to CO<sub>2</sub> and threitol [93, 94]. Five percent is excreted unchanged in the bile and undergoes enterohepatic cycling [95], and the remainder is excreted in the urine.

Normal subjects absorb approximately 70% of a 25g oral dose of D-xylose [96]. The absorption constant for D-xylose correlates closely with the 1-h serum concentration of D-xylose, and the 5-h urine content of D-xylose correlates closely with it's absolute bioavailability [97]. Essentially all of the absorption, metabolism, and excretion occur within the first five hours after administration.

From the 1960s and onwards a range of studies was performed which showed that a 5-h urine excretion test with 25g D-xylose distinguished clearly between normal individuals and patients with proximal intestinal malabsorption [98-110]. To improve the sensitivity, specificity and ease of D-xylose testing, various modifications of dosing and the inclusion of serum analyses have been suggested. Most studies have used the 25g or 5g D-xylose absorption test with either 5-h urine collection and/or 1-h serum levels. In

paediatrics, the 1-h serum level after 5g of D-xylose has been the preferred test due to the difficulty of collecting urine accurately in children [111-116].

Other factors than malabsorption may affect the result of the D-xylose test and must be considered in clinical practice. The excretion of D-xylose in urine is dependent on its renal clearance, and renal insufficiency complicates the interpretation of the test [117, 118]. Interpretation of D-xylose tests in geriatric patients must therefore take status of renal function into account [119-121]. Delayed gastric emptying may alter the absorption kinetics and give low 1-h serum levels [122].

The 25g D-xylose H<sub>2</sub>-breath test was developed as an alternative to urine and serum analysis of D-xylose. This is an indirect test of D-xylose malabsorption, in which a rise in H<sub>2</sub> signals that unabsorbed D-xylose has been delivered to the colon and metabolized by enteric bacteria. The H<sub>2</sub>-breath test circumvents some of the problems with the traditional D-xylose tests such as dependence on renal clearance and accurate urine collection. H<sub>2</sub> production after a 25g D-xylose dose was first shown to be well correlated with 5-h D-xylose excretion and 90-min D-xylose levels in serum in tropical sprue [123]. Similar results were found when H<sub>2</sub>-breath testing was compared with 5-h urinary D-xylose for patients with various malabsorptive disorders [124]. One study showed that the 25g D-xylose H<sub>2</sub>-breath test was more reliable in diagnosing coeliac disease than the 5-h urine D-xylose content [125]. Isotope-labelled breath tests with D-xylose have been evaluated for small intestinal bacterial overgrowth, but not for coeliac disease or other malabsorptive states.

# 3.4 Summary of introduction

Coeliac disease is a small intestinal enteropathy that can develop in genetically disposed individuals upon exposure to gluten. Before the development of serological tests for coeliac disease and easy access to endoscopic procedures, malabsorption testing played a pivotal role in the diagnosis of coeliac disease. D-xylose testing was the most commonly used malabsorptive test. Initially D-xylose measurements were made in serum and urine, but in later years the D-xylose H<sub>2</sub>-breath test was developed as an alternative. Isotope-labelled breath tests have been much used in gastroenterology, but not as primary tests of coeliac disease or other malabsorptive states.

#### 4 AIM OF THE STUDY

The aim of this thesis has been to evaluate isotope-labelled breath tests as diagnostic tools in small intestinal malabsorption. We chose to focus on coeliac disease; the most frequent cause of small intestinal malabsorption in the Western world today.

The specific aims of the present study were:

- 1. To determine the diagnostic accuracy of the <sup>13</sup>C-xylose breath test in diagnosis and follow-up of coeliac disease.
- 2. To compare the <sup>13</sup>C-xylose breath test with xylose absorption tests based on D-xylose measurements in blood and urine.
- **3.** To develop a novel <sup>13</sup>C-sorbitol breath test and determine the diagnostic accuracy of this test in diagnosis and follow-up of coeliac disease.
- **4.** To compare the <sup>13</sup>C-sorbitol breath test with the conventional H<sub>2</sub>-sorbitol breath test.

#### **5 LIST OF PAPERS**

Paper I

Tveito K, Brunborg C, Sandvik L, Løberg EM, Skar V. 13C-xylose and 14C-xylose breath tests for the diagnosis of coeliac disease. Scand J Gastroenterol. 2008;43(2):166-173.

Paper II

Tveito K, Brunborg C, Bratlie J, Askedal M, Sandvik L, Lundin KE., Skar V. Intestinal malabsorption of D-xylose: comparison of test modalities in patients with coeliac disease. Scand J Gastroenterol. 2010 Nov;45(11):1289-94

Paper III

Tveito K, Hetta AK, Askedal M, Brunborg C, Sandvik L, Løberg EM, Skar V. A novel one-hour 13C-sorbitol breath test versus the H2-sorbitol breath test for assessment of coeliac disease. Scand J Gastroenterol. 2009;44(7):813-19.

Paper IV

Tveito K, Hetta AK, Askedal M, Løberg EM, Skar V. Follow-up of coeliac disease with the novel one-hour 13C-sorbitol breath test versus the H2-sorbitol breath test.

(Submitted)

## 5.1 Paper I

The aim of the study was to evaluate the diagnostic properties of the <sup>13</sup>C-D-xylose breath test and the <sup>14</sup>C-D-xylose breath test in coeliac disease and develop a diagnostic breath test index.

Data from 41 patients with untreated coeliac disease who had performed the <sup>14</sup>C-D-xylose breath test, and 60 patients with untreated coeliac disease who had performed the <sup>13</sup>C-D-xylose breath test were reviewed retrospectively. All patients in the <sup>14</sup>C-D-xylose breath test cohort repeated the breath test on a gluten-free diet (GFD), and 37 patients in the <sup>13</sup>C-D-xylose breath test cohort repeated the breath test on GFD. Patients were compared to a healthy control group.

For both breath tests, combining breath test values at 30 minutes and 210 minutes (the gas fraction (GF) index: 30 min/210 min) resulted in the highest diagnostic accuracy. The breath tests had very similar diagnostic properties for untreated coeliac disease, with sensitivities of 84-95% and specificities of 87-94%.

After treatment with a gluten-free diet, most patients improved their GF-index, but did not reach the levels of healthy controls.

# 5.2 Paper II

The aim of this study was to compare the  $^{13}$ C-D-xylose breath test with D-xylose measurements in plasma and urine in a retrospective cohort of patients with coeliac disease. Data from 91 untreated coeliac patients, and 98 treated coeliac patients, who had performed the  $^{13}$ C-D-xylose breath test were evaluated. 1-h plasma D-xylose had been measured in 48 untreated and 41 treated coeliac patients. 4-h urine D-xylose excretion in urine had been measured in 47 untreated and 51 treated patients. Patients were compared to a healthy control group (n = 43).

Test sensitivity and specificity for coeliac disease versus healthy controls were 88% and 84% with the <sup>13</sup>C-D-xylose breath test, 65% and 71% with the 1-h plasma D-xylose test, and 55% and 74% with the 4-h urine D-xylose excretion test. Breath test results improved significantly in the treated coeliac group compared to untreated patients, but were not normalized compared to healthy control subjects. No difference was found between D-xylose levels in plasma and urine between treated coeliac patients and healthy control subjects.

## 5.3 Paper III

The aim of this study was to examine the diagnostic properties of a novel <sup>13</sup>C-sorbitol breath test in coeliac disease. We compared the <sup>13</sup>C-sorbitol breath test to the H<sub>2</sub>-sorbitol breath test in 39 untreated coeliac patients, 40 patients with symptoms compatible with coeliac disease referred for evaluation who turned out not to have coeliac disease, and 26 healthy control subjects.

The <sup>13</sup>C- sorbitol breath test and the H<sub>2</sub>-sorbitol breath test were performed simultaneously. Breath samples were gathered every 30 minutes for four hours. Single time point variables and combinations of single time point variables were analyzed statistically to chose the best test variable(s) for the <sup>13</sup>C-sorbitol breath test. Optimal combined sensitivity and specificity turned out to be obtained with single time point measurements at 60 minutes or 90 minutes. The 60 minutes variable was chosen to shorten test duration. A diagnostic algorithm dividing patients into high, moderate and low risk groups for coeliac disease was proposed. Following the algorithm, sixty-two per cent of coeliac patients were detected with 100% specificity. The H<sub>2</sub>-sorbitol breath test had a sensitivity of 71%, a specificity of 46% versus healthy controls, and a specificity of 25% versus patient controls.

## 5.4 Paper IV

The aim of this study was to evaluate the novel <sup>13</sup>C-sorbitol breath test for follow-up of coeliac disease compared to the H<sub>2</sub>-sorbitol breath test. Patients with coeliac disease from the first study of the <sup>13</sup>C-sorbitol breath study were invited to participate after approximately one year on a gluten-free diet. 27/39 patients repeated the breath tests, and 25 patients had a new duodenal biopsy. Patients were compared to 40 patients without coeliac disease and 26 healthy subjects from the previous study.

As in the previous study, the <sup>13</sup>C- sorbitol breath test and the H<sub>2</sub>-sorbitol breath test were performed simultaneously. Breath samples were gathered every 30 minutes for four hours.

The rate of <sup>13</sup>CO<sub>2</sub> excretion increased significantly at all time points before 210 minutes in treated versus untreated coeliac patients, but remained lower than in healthy control subjects and patient controls at all time points except 30 minutes. This also applied to treated patients who had completely normal histological duodenal mucosa after treatment with GFD. 20/26 patients had increased <sup>13</sup>CO<sub>2</sub> breath levels after 60 minutes compared to their test before treatment. 17/27 patients had a negative H<sub>2</sub>-sorbitol breath test after treatment with GFD. Peak H<sub>2</sub> concentrations did not correlate with <sup>13</sup>C-sorbitol breath test results. Mean time to reach peak H<sub>2</sub> concentration and mean peak H<sub>2</sub> concentration did not change after treatment.

# 5.5 Other publications during the PhD programme

Hope HB, Tveito K, Aase S, Messelt E, Utzon P, Skar V. Small intestinal malabsorption in chronic alcoholism determined by 13C-D-xylose breath test and microscopic examination of the duodenal mucosa. Scand J Gastroenterol. 2010;45(1):39-45. Erratum in: Scand J Gastroenterol. 2010 Dec;45(12):1519.

#### 6 METHODS

## 6.1 Study subjects

#### 6.1.1 Patients

The <sup>13</sup>C-D-xylose breath test studies had a retrospective design. Patient data were collected from the breath test database of <sup>13</sup>C-D-xylose breath tests and <sup>14</sup>C-D-xylose breath tests at Lovisenberg Diakonale Hospital and Ullevaal University Hospital. Clinical files and duodenal biopsies were re-assessed to confirm that the diagnosis of coeliac disease was correct.

In the <sup>13</sup>C-sorbitol breath test studies patients were included and followed up prospectively between 2006 and 2008 in the Department of Gastroenterology at Stavanger University Hospital and Lovisenberg Diakonale Hospital. Consecutive patients with suspected coeliac disease were included after giving written consent to participate in the study. Patients underwent gastroscopy with duodenal biopsies, routine laboratory testing and were questioned about gastrointestinal symptoms at inclusion and follow-up. Patients who did not have biopsy-confirmed coeliac disease served as patient controls. Patients with verified coeliac disease were followed-up after approximately one year.

## 6.1.2 Healthy control subjects

Healthy control subjects were recruited from the medical staff and medical students at Lovisenberg Diakonale Hospital. Some of the test subjects had performed the breath test as controls when the breath test was introduced at Lovisenberg Diakonale Hospital. They

had not been tested with serologic markers for coeliac disease. Healthy control subjects included after 2004 underwent serological testing for IgA and IgG tissue-transglutaminase antibodies, and levels within reference values were required to participate. All healthy control subjects had normal bowel habits, and had not been investigated for gastrointestinal disease in the past. In the <sup>13</sup>C-sorbitol breath test studies, serious illness in the past, having a 1<sup>st</sup> degree relative with coeliac disease and having used antibiotics in the previous month served as additional exclusion criteria.

## 6.2 Breath test protocols

All breath tests were performed after an overnight fast. Subjects were quietly seated and requested not to eat, drink or smoke during the test period. In the <sup>13</sup>C-sorbitol breath test where hydrogen and methane were also measured, subjects were instructed to ingest a meal consisting of fish or chicken and rice and avoid high fibre food the evening before the test. Breath was sampled in duplicates at every time point, and mean values of the duplicates were used.

# 6.2.1 <sup>14</sup>C-D-xylose breath test

1 g D-xylose and 10  $\mu$ Ci <sup>14</sup>C-D-xylose, 99% (Amersham, U.K.), were given orally in 500 ml water as described by Skar et al. [126]. The subjects exhaled through a disposable device directly into counting vials. Specific radioactivity of the breath samples was determined by liquid scintillation. The results were expressed as percentage of orally administered dose <sup>14</sup>C-D-xylose expired as <sup>14</sup>CO<sub>2</sub> per mmol CO<sub>2</sub>, corrected for the endogenous production of CO<sub>2</sub> (9 mmol CO<sub>2</sub>/kg body weight/h).

# 6.2.2 13 C-D-xylose breath test

100 mg  $^{13}$ C-1, 2-D-xylose, 99% (Cambridge Isotope Laboratories, Massachusetts, USA), and 5 g D-xylose were dissolved in 250 ml tap water. Two end-expiratory breath samples were collected via a straw in 12 ml tubes (Labco Limited, High Wycombe, UK) at each time point. The  $^{13}$ CO<sub>2</sub>/ $^{12}$ CO<sub>2</sub> ratio in the breath samples was determined by gas chromatography (GC) and continuous flow isotope ratio mass spectrometry (IRMS, Europe Scientific 20/20, Crewe, UK). The delta-value ( $\delta$ ) between the  $^{13}$ CO<sub>2</sub>/ $^{12}$ CO<sub>2</sub> ratio of the samples and the PDB limestone standard was calculated [127], using the following formula:

 $\delta = [(R_{sa}/R_{st})-1] \times 1000$ , in which  $R_{sa} = ^{13}C/^{12}C$  in the  $CO_2$  of the sample and  $R_{st} = ^{13}C/^{12}C$  in the  $CO_2$  of the standard. The results were expressed as the percentage of  $^{13}C$ -recovery per hour (% dose/h). For this calculation the formula of Schoeller et al. was used [128], and the  $CO_2$  production was assumed to be 300 mmol/m<sup>2</sup> body surface area per hour. Body surface area was calculated by the weight-height formula of Haycock et al [129].

# 6.2.3 <sup>13</sup>C-sorbitol breath test

5 g D-sorbitol and 100 mg <sup>13</sup>C-D-sorbitol, 99% (Aldrich, Milwaukee, USA) dissolved in 250 ml tap water was given orally. Two parallel end-expiratory breath samples were collected via a straw in 12 ml tubes (Exetainers, Labco Limited, High Wycombe, UK) at every time point for analysis of <sup>13</sup>CO<sub>2</sub>. The <sup>13</sup>CO<sub>2</sub>/<sup>12</sup>CO<sub>2</sub> -ratio in the breath samples was determined by gas chromatography (GC) and continuous flow isotope ratio mass

spectrometry (IRMS, Europe Scientific 20/20, Crewe, UK), and expressed as the percentage of <sup>13</sup>C-recovery (% of dose <sup>13</sup>C/h) for every time point with a similar formula as in the in the <sup>13</sup>C-D-xylose breath test studies.[128] The CO<sub>2</sub> production was assumed to be 300 mmol/m<sup>2</sup> body surface area per hour. Body surface area was calculated by the weight-height formula of Haycock et al.[129].

#### 6.2.4 H<sub>2</sub>-sorbitol breath test

Duplicate end-expiratory breath samples were collected in 30 ml plastic syringes fitted with a T-piece and immediately analyzed for  $H_2$  and methane (CH<sub>4</sub>) by a QuinTron Model SC Microlyzer (QuinTron Instrument Company, Milwaukee, USA).

Concentrations used for calculation were means of duplicate samples. Hydrogen increase  $\geq 20$  ppm from the lowest previous level reached prior to an increase counted as a positive test.

## 6.3 D-xylose in plasma and urine

Blood samples were drawn after one hour, and urine was collected for four hours after starting the test with an empty bladder. Duplicate samples of blood and urine were frozen at -20 degrees  $C^{\circ}$  before analysis.

D-xylose was measured in urine by high-performance liquid chromatography (HPLC) modified after Larew and Johnson,[130] using an electrochemical detector equipped with a gold electrode (ESA coulochem, model 5040, Chelmsford, Massachusetts, USA), a Rheodyn injector (Rheodyn inc., California, USA), 20 mikroliter loop and a Supelco C-611 column with Supelcogel C-611 guarding column (Supelco, Bellefonte, Pennsylvania,

USA). Mobil phase was freshly prepared 1mmol/l NaOH (Sigma, St.Louis, Missouri, USA) delivered at 1.0 ml/minute isocratic run at 65 degrees C°. Lactose (Sigma) was used as internal standard and D-xylose (Sigma) as external standard. Peak heights were measured and corrected according to the internal standard. A standard curve was used to determine D-xylose concentration. The coefficient of variation was 6%.

1-h plasma D-xylose levels were measured in duplicate samples by the method of Roe and Rice [131]. The coefficient of variation was 10%.

#### 6.4 Statistics

Statistical analyses were performed using SPSS 14.0, 15.0 or 18.0. Comparison between independent groups was made using either independent sample Student's t-test or one-way ANOVA for normally distributed variables, and Mann Whitney U test or Kruskal-Wallis test for continuous variables with skewed distribution. Linear regression was used when comparing two continuous variables. Pearson's chi-square test was used for categorical outcome variables. Pearson's correlation coefficient or Spearman's correlation coefficient was used to estimate correlation between variables. Single time point test variables were analyzed by logistic regression. Receiver operating characteristic (ROC) curves were used to determine diagnostic accuracy. Findings with p values < 0.05 were considered statistically significant.

#### 6.5 Ethics

The regional ethical committee approved all studies. Informed consent was given by all control subjects and by patients included after 2004.

#### 7 Discussion

In this first series of studies of <sup>13</sup>C-labelled breath tests in coeliac disease we found high sensitivity for untreated coeliac disease compared to healthy control subjects, and in the <sup>13</sup>C-sorbitol breath test study, also compared to a patient group with comparable clinical symptoms. The latter finding is especially important as it demonstrates the specificity of <sup>13</sup>C-breath testing in clinical practice.

In follow-up studies of coeliac patients on a gluten-free diet, we found improved breath test results, but treated coeliac patients did not reach healthy control levels. Furthermore, we found that the <sup>13</sup>C-D-xylose breath test has comparable diagnostic properties to the <sup>14</sup>C-D-xylose breath test, and is superior to D-xylose measurements in plasma and urine. Finally, when comparing the <sup>13</sup>C-sorbitol breath test to the H<sub>2</sub>-sorbitol breath test, we showed superiority of the <sup>13</sup>C-labelled breath test versus H<sub>2</sub>-testing.

## 7.1 Strengths and limitations of the studies

The validity of our findings is strengthened by the fact that we carried out studies with three different isotope-labelled breath test protocols with very similar test results.

Furthermore, many patients were tested before and after treatment with a gluten-free diet, which gave us the opportunity to investigate individual changes in breath test results.

Comparison with other test modalities was performed simultaneously. Thus we could exclude intraindividual variance over time as a confounding factor. The prospective design of the <sup>13</sup>C-sorbitol breath test study is considered a major strength of this study, especially since we were able to collect both clinical, biochemical and histological data.

Other factors than malabsorption such as dysmotility and small intestinal bacterial overgrowth can interfere with breath test results. A limitation of our study is that we did not carry out additional studies of this. A limitation of the study of D-xylose modalities is that we carried out a 4-h xylose excretion test in urine instead of the traditional 5-h D-xylose excretion test and measured 1-h D-xylose levels in plasma instead of serum.

Further studies are needed to assess the relationship between mucosal changes in patients on a gluten-free diet and breath test results as this was only done in a relatively small number of patients in the <sup>13</sup>C-sorbitol breath test study.

# 7.2 The <sup>13</sup>C-D-xylose breath test and the <sup>13</sup>C-sorbitol breath test

One of the aims in the studies with both isotopes was to decide what is the optimal test duration and determine a diagnostic algorithm. We reached different conclusions for the two breath tests. In the study of the <sup>13</sup>C-D-xylose breath test (and the <sup>14</sup>C-D-xylose breath test), we concluded that the highest diagnostic accuracy was achieved combining excretion of breath <sup>13</sup>CO<sub>2</sub> in the early and late test phase. The 30min/210 min index was proposed. With the <sup>13</sup>C- sorbitol breath test, we found that a single measurement at 60 minutes was equally good as a test index combining several variables. How can we explain this discrepancy?

When looking at the breath test curves from the <sup>13</sup>C-D-xylose breath test and the <sup>13</sup>C-sorbitol breath test, a striking difference is apparent in the late test phase. The <sup>13</sup>CO<sub>2</sub> overshoot in the late test phase in coeliac patients with the <sup>13</sup>C-D-xylose breath test has no equivalent in the <sup>13</sup>C-sorbitol breath test. This probably explains why adding a late test

variable did not increase diagnostic accuracy in the <sup>13</sup>C-sorbitol breath test study. We do not know the reason for the different patterns of <sup>13</sup>CO<sub>2</sub> excretion with the two different <sup>13</sup>C-substrates. We predict that it has to do with the amount of substrate reaching colon and colonic fermentation of the substrates.

As for the question of which breath test should be preferred in clinical practice, several aspects should be considered. The best combined sensitivity and specificity for coeliac disease versus healthy controls was found with the <sup>13</sup>C-D-xylose breath test. The classic sprue syndrome of steatorrhoea and malnutrion has become rare, and there has been a trend over time for a greater proportion of coeliac patients presenting with subclinical/silent coeliac disease and having partial villous atrophy [35, 132]. In line with the reports on the changing clinical spectre of coeliac disease, we found that most of the patients in our studies had mild if any signs of malabsorption. This change in clinical presentation might well explain the reduced sensitivity and specificity we found for D-xylose testing in plasma and urine compared to early studies of D-xylose tests. Our results indicate that the <sup>13</sup>C-D-xylose breath test is a sensitive test for subtle small intestinal malabsorption. This conclusion is supported by the fact that D-xylose levels in plasma and urine did not distinguish between treated coeliac disease and healthy control subjects, whereas the <sup>13</sup>C-D-xylose and the <sup>14</sup>C-D-xylose breath test did.

The <sup>13</sup>C-sorbitol breath test had lower optimal combined sensitivity and specificity than the <sup>13</sup>C-D-xylose breath test, but offers a breath test alternative that is short (60 minutes), and can be used to stratify patients into high, medium and low risk for coeliac disease. Furthermore, the <sup>13</sup>C-sorbitol substrate is less expensive than <sup>13</sup>C-D-

xylose. Finally, the changes in <sup>13</sup>C-sorbitol breath test results were similar to the changes in the <sup>13</sup>C-D-xylose breath test in coeliac patients after treatment with a gluten-free diet.

#### 7.3 The H<sub>2</sub>-sorbitol breath test

Both studies of the H<sub>2</sub>-breath test showed unsatisfactory diagnostic accuracy with the H<sub>2</sub>sorbitol breath test. The main weakness of the H2-sorbitol breath test was the high prevalence of positive breath tests in control groups and in treated coeliac patients. The lack of specificity may be specific to the H<sub>2</sub>-sorbitol breath test, but could also reflect a more general problem with the H<sub>2</sub>-breath tests. Several factors point to the latter conclusion. The general use of H<sub>2</sub>-breath tests might be thwarted by the variability of individual gut flora to produce H<sub>2</sub> [133-136]. As many as 10% of some populations may be non-H<sub>2</sub> producers [137-139]. False positive results may also result due to the oral bacterial flora and if the subject has failed to adhere to a low fibre diet the day before the test [140]. In line with previous studies, we showed that H<sub>2</sub>-excretion is dependent on the presence of methanogenic colonic flora [75, 76, 80]. Methane should also be measured and considered when interpreting the H<sub>2</sub>-breath test. Previous studies of the H<sub>2</sub>-sorbitol breath test have not measured CH<sub>4</sub>, and we find this a major weakness of these studies. The specificity of the H<sub>2</sub>-sorbitol breath test depends critically on the sorbitol absorption capacity in healthy subjects. Further studies may elucidate if a changing the amount and concentration of sorbitol in the test solution could improve test specificity.

## 7.4 Pathophysiological considerations

We have concluded that the <sup>13</sup>C-breath tests are sensitive markers of small intestinal malabsorption. However, other factors than small intestinal damage such as small intestinal bacterial overgrowth and dysmotility can also give rise to pathological breath test results. Fordtran et al. first demonstrated a relationship between small intestinal transit time and extent of D-xylose absorption [92, 97]. Subsequently, Goldstein et al. showed that bacteria in the small intestinal lumen can metabolize substantial amounts of D-xylose [141].

Studies of small bacterial overgrowth in coeliac disease are scarce, but the few studies that have been done conclude that small intestinal bacterial overgrowth occurs in a small percentage of untreated coeliac patients, and should be suspected in coeliac patients with persistent clinical symptoms on a gluten-free diet [142, 143]. We did not carry out any experiments to solve this issue. However, the maximum rise in H<sub>2</sub> occurred at a late time point in the <sup>13</sup>C-sorbitol breath test and did not differ from the time to reach peak H<sub>2</sub> concentration in controls. If small intestinal bacterial overgrowth had been present in coeliac patients, we would expect bacterial metabolism of sorbitol and H<sub>2</sub>-rise in the early test phase before colonic fermentation occurred [123]. The time to reach peak H<sub>2</sub> concentration did not change in coeliac patients after gluten-free diet. If small bacterial overgrowth had been present in untreated patients and resolved after treatment, we would expect the time to reach H<sub>2</sub> peak concentration to change. Furthermore, the shape of the isotope-labelled breath test curves did not suggest small intestinal bacterial overgrowth. In SIBO, we would expect high rates of <sup>13</sup>CO<sub>2</sub> excretion in the early test phase (first two hours) due to bacterial metabolism of the substrate in the small intestine.

Several studies have found evidence for gastrointestinal dysmotility in coeliac disease [144-146]. We would expect changes in gastric emptying and small intestinal transit time to reduce and/or delay small intestinal absorption of the substrate. We cannot exclude the possibility that our breath tests reflect dysmotility rather than small intestinal malabsorption per se. However, time to reach peak H<sub>2</sub> concentration was similar in all groups, indicating that there were no significant differences in intestinal transit time between groups. Furthermore, Bjarnarson et al. carried out a series of permeability studies in coeliac disease with <sup>51</sup>Cr EDTA absorption test and found very similar results to our <sup>13</sup>C-breath test studies [147]. Since the <sup>51</sup>Cr-EDTA absorption test consists of a 24 hours urine excretion test, we can exclude the influence of dysmotility on their results.

al. They reported that although permeability was significantly reduced in treated coeliac patients, permeability remained increased compared to healthy control subjects.

Furthermore, they found that 11/14 patients on gluten-free diet with a completely normal mucosa, still had a pathological <sup>51</sup>Cr EDTA absorption test. Based on their findings they raised the intriguing question if patients with coeliac disease have an inherent mucosal defect that gives rise to increased permeability even when histological findings have resolved. In the <sup>13</sup>C-sorbitol breath test study we reached the same conclusion. Breath test results remained abnormal compared to healthy control subjects even in patients with normal duodenal biopsies according to the Marsh classification.

Interestingly, we found similar changes in treated coeliac disease as Bjarnarson et

Studies of patients with HIV and studies of patients with alcoholism have showed abnormal <sup>13</sup>C-D-xylose breath tests in the absence of histological small mucosal changes [148, 149]. However, when electron microscopy was performed, subtle mucosal changes

were detected. We propose that similar findings might be affecting small intestinal absorption capacity in coeliac patients on a gluten-free diet. Further studies are needed to investigate this issue, and to determine the clinical importance of the findings.

Bjarnarson et al. found no association between clinical variables in coeliac patients and permeability test results [150]. Likewise, there was no correlation between clinical symptoms and biochemical signs of malabsorption and breath test results in untreated coeliac patients in the <sup>13</sup>C-sorbitol breath test studies. However, we did find a significant association between IgA TGA levels and breath test results in the untreated coeliac group. Our findings are in line with a previous study showing lack of correlation between clinical presentation and histological changes in coeliac disease [35, 36].

#### 7.5 Clinical considerations

Malabsorption testing in coeliac disease was developed and studied for many years before the development of excellent serological tests for coeliac disease and easy access to endoscopic procedures. Although breath testing is probably not useful as a routine test in coeliac disease, it might be a valuable additional test in complicated cases, and in patients where duodenal biopsies cannot be obtained. The non-invasiveness of the test and lack of serious side effects, makes it useful in children and pregnant women. We suggest that the <sup>13</sup>C-breath test protocols should be evaluated also in children with coeliac disease. Further studies are needed to decide how breath testing can be of value in follow-up of coeliac disease. Several researchers have emphasized the importance of early diagnosis of coeliac disease. Further studies are needed to determine if isotope labelled

breath tests may be of value in the investigation of patients with subtle duodenal histological pathology and so called borderline coeliac disease.

Recent studies show that despite a good clinical response, abnormal endoscopic and histopathological appearances persist in the majority of adult patients with coeliac disease on a gluten-free diet [151-153]. The clinical significance of these findings is uncertain. One study from Finland showed that patients without histological recovery on a gluten-free diet were at greater risk of developing complications of coeliac disease such as malignancy [154]. The normalization of tissue transglutaminase antibodies does not in itself guarantee that the small intestinal villous structure has improved [155]. We do not know if the <sup>13</sup>C-labelled breath tests can be used to determine which patients should be followed up with repeated duodenal biopsies, but this question could be addressed in further studies.

### 7.6 Conclusion

Patients with coeliac disease are excellent experimental models for small intestinal malabsorption since they can be examined before and after treatment with a gluten-free diet. Based on our studies of patients with coeliac disease, we conclude that <sup>13</sup>C-labelled breath tests are sensitive tests for small intestinal malabsorption.

#### 8 References

- [1] Di Sabatino A, Corazza GR. Coeliac disease. Lancet. 2009 Apr 25;373(9673):1480-93.
- [2] Losowsky MS. A history of coeliac disease. Digestive diseases (Basel, Switzerland). 2008;26(2):112-20.
- [3] Hemminki K, Li X, Sundquist J, Sundquist K. Familial association between type 1 diabetes and other autoimmune and related diseases. Diabetologia. 2009 Sep;52(9):1820-8
- [4] Guliter S, Yakaryilmaz F, Ozkurt Z, Ersoy R, Ucardag D, Caglayan O, et al. Prevalence of coeliac disease in patients with autoimmune thyroiditis in a Turkish population. World J Gastroenterol. 2007 Mar 14;13(10):1599-601.
- [5] Mirzaagha F, Azali SH, Islami F, Zamani F, Khalilipour E, Khatibian M, et al. Coeliac disease in autoimmune liver disease: A cross-sectional study and a systematic review. Dig Liver Dis. Mar 15.
- [6] Troncone R, Auricchio R, Paparo F, Maglio M, Borrelli M, Esposito C. Coeliac disease and extraintestinal autoimmunity. J Pediatr Gastroenterol Nutr. 2004 Jun;39 Suppl 3:S740-1.
- [7] Viljamaa M, Kaukinen K, Huhtala H, Kyronpalo S, Rasmussen M, Collin P. Coeliac disease, autoimmune diseases and gluten exposure. Scand J Gastroenterol. 2005 Apr;40(4):437-43.
- [8] Biagi F, Campanella J, Soriani A, Vailati A, Corazza GR. Prevalence of coeliac disease in Italian patients affected by Addison's disease. Scand J Gastroenterol. 2006 Mar;41(3):302-5.
- [9] Guariso G, Conte S, Presotto F, Basso D, Brotto F, Visona Dalla Pozza L, et al. Clinical, subclinical and potential autoimmune diseases in an Italian population of children with coeliac disease. Alimentary pharmacology & therapeutics. 2007 Nov 15:26(10):1409-17.
- [10] West J, Logan RF, Smith CJ, Hubbard RB, Card TR. Malignancy and mortality in people with coeliac disease: population based cohort study. Bmj. 2004 Sep 25;329(7468):716-9.
- [11] Peters U, Askling J, Gridley G, Ekbom A, Linet M. Causes of death in patients with coeliac disease in a population-based Swedish cohort. Archives of internal medicine. 2003 Jul 14;163(13):1566-72.
- [12] Corrao G, Corazza GR, Bagnardi V, Brusco G, Ciacci C, Cottone M, et al. Mortality in patients with coeliac disease and their relatives: a cohort study. Lancet. 2001 Aug 4;358(9279):356-61.
- [13] Sollid LM, Markussen G, Ek J, Gjerde H, Vartdal F, Thorsby E. Evidence for a primary association of coeliac disease to a particular HLA-DQ alpha/beta heterodimer. The Journal of experimental medicine. 1989 Jan 1;169(1):345-50.
- [14] Sollid LM, Lie BA. Coeliac disease genetics: current concepts and practical applications. Clin Gastroenterol Hepatol. 2005 Sep;3(9):843-51.
- [15] Wolters VM, Wijmenga C. Genetic background of coeliac disease and its clinical implications. Am J Gastroenterol. 2008 Jan;103(1):190-5.

- [16] Nistico L, Fagnani C, Coto I, Percopo S, Cotichini R, Limongelli MG, et al. Concordance, disease progression, and heritability of coeliac disease in Italian twins. Gut. 2006 Jun;55(6):803-8.
- [17] Stene LC, Honeyman MC, Hoffenberg EJ, Haas JE, Sokol RJ, Emery L, et al. Rotavirus infection frequency and risk of coeliac disease autoimmunity in early childhood: a longitudinal study. Am J Gastroenterol. 2006 Oct;101(10):2333-40.
- [18] Forsberg G, Fahlgren A, Horstedt P, Hammarstrom S, Hernell O, Hammarstrom ML. Presence of bacteria and innate immunity of intestinal epithelium in childhood coeliac disease. Am J Gastroenterol. 2004 May;99(5):894-904.
- [19] Cammarota G, Cuoco L, Cianci R, Pandolfi F, Gasbarrini G. Onset of coeliac disease during treatment with interferon for chronic hepatitis C. Lancet. 2000 Oct 28:356(9240):1494-5.
- [20] Norris JM, Barriga K, Hoffenberg EJ, Taki I, Miao D, Haas JE, et al. Risk of coeliac disease autoimmunity and timing of gluten introduction in the diet of infants at increased risk of disease. Jama. 2005 May 18;293(19):2343-51.
- [21] Ivarsson A, Hernell O, Stenlund H, Persson LA. Breast-feeding protects against coeliac disease. Am J Clin Nutr. 2002 May;75(5):914-21.
- [22] Oberhuber G, Granditsch G, Vogelsang H. The histopathology of coeliac disease: time for a standardized report scheme for pathologists. Eur J Gastroenterol Hepatol. 1999 Oct;11(10):1185-94.
- [23] Rostami K, Kerckhaert J, Tiemessen R, von Blomberg BM, Meijer JW, Mulder CJ. Sensitivity of antiendomysium and antigliadin antibodies in untreated coeliac disease: disappointing in clinical practice. Am J Gastroenterol. 1999 Apr;94(4):888-94.
- [24] Kull K, Uibo O, Salupere R, Metskula K, Uibo R. High frequency of antigliadin antibodies and absence of antireticulin and antiendomysium antibodies in patients with ulcerative colitis. Journal of gastroenterology. 1999 Feb;34(1):61-5.
- [25] Catassi C, Fanciulli G, D'Appello AR, El Asmar R, Rondina C, Fabiani E, et al. Antiendomysium versus antigliadin antibodies in screening the general population for coeliac disease. Scand J Gastroenterol. 2000 Jul;35(7):732-6.
- [26] Dieterich W, Ehnis T, Bauer M, Donner P, Volta U, Riecken EO, et al. Identification of tissue transglutaminase as the autoantigen of coeliac disease. Nat Med. 1997;3(7):797-801.
- [27] Rostom A, Dube C, Cranney A, Saloojee N, Sy R, Garritty C, et al. The diagnostic accuracy of serologic tests for coeliac disease: a systematic review. Gastroenterology. 2005 Apr;128(4 Suppl 1):S38-46.
- [28] Hill ID. What are the sensitivity and specificity of serologic tests for coeliac disease? Do sensitivity and specificity vary in different populations? Gastroenterology. 2005 Apr;128(4 Suppl 1):S25-32.
- [29] Tommasini A, Not T, Kiren V, Baldas V, Santon D, Trevisiol C, et al. Mass screening for coeliac disease using antihuman transglutaminase antibody assay. Arch Dis Child. 2004 Jun;89(6):512-5.
- [30] Maki M, Mustalahti K, Kokkonen J, Kulmala P, Haapalahti M, Karttunen T, et al. Prevalence of Coeliac disease among children in Finland. N Engl J Med. 2003 Jun 19;348(25):2517-24.

- [31] Fasano A, Berti I, Gerarduzzi T, Not T, Colletti RB, Drago S, et al. Prevalence of coeliac disease in at-risk and not-at-risk groups in the United States: a large multicenter study. Archives of internal medicine. 2003 Feb 10;163(3):286-92.
- [32] West J, Logan RF, Hill PG, Lloyd A, Lewis S, Hubbard R, et al. Seroprevalence, correlates, and characteristics of undetected coeliac disease in England. Gut. 2003 Jul;52(7):960-5.
- [33] Catassi C, Ratsch IM, Gandolfi L, Pratesi R, Fabiani E, El Asmar R, et al. Why is coeliac disease endemic in the people of the Sahara? Lancet. 1999 Aug 21:354(9179):647-8.
- [34] Corazza GR, Frisoni M, Treggiari EA, Valentini RA, Filipponi C, Volta U, et al. Subclinical coeliac sprue. Increasing occurrence and clues to its diagnosis. J Clin Gastroenterol. 1993 Jan;16(1):16-21.
- [35] Brar P, Kwon GY, Egbuna, II, Holleran S, Ramakrishnan R, Bhagat G, et al. Lack of correlation of degree of villous atrophy with severity of clinical presentation of coeliac disease. Dig Liver Dis. 2007 Jan;39(1):26-9; discussion 30-2.
- [36] Murray JA, Rubio-Tapia A, Van Dyke CT, Brogan DL, Knipschield MA, Lahr B, et al. Mucosal atrophy in coeliac disease: extent of involvement, correlation with clinical presentation, and response to treatment. Clin Gastroenterol Hepatol. 2008 Feb;6(2):186-93; quiz 25.
- [37] Dodds EC. Variations in alveolar carbon dioxide pressure in relation to meals. J Physiol. 1920;54.
- [38] Dodds EC, Bennett TI. Variations in alveolar carbon dioxide pressure in relation to meals. A further study. J Physiol. 1920;54.
- [39] Hepner GW. Breath tests in gastroenterology. Advances in internal medicine. 1978;23:25-45.
- [40] Schwabe AD, Cozzetto FJ, Bennettlr, Mellinkoff SM. Estimation of fat absorption by monitoring of expired radio-active carbon dioxide after feeding a radioactive fat. Gastroenterology. 1962 Mar;42:285-91.
- [41] Cozzetto FJ. Radiocarbon Estimates of Intestinal Absorption. Studies of Breath Excretion of C14o2 after Ingestion of Labeled Fatty Acids, Glucose, and Lactose. American journal of diseases of children (1960). 1964 Jun;107:605-11.
- [42] Watkins JB, Schoeller DA, Klein PD, Ott DG, Newcomer AD, Hofmann AF. 13C-trioctanoin: a nonradioactive breath test to detect fat malabsorption. J Lab Clin Med. 1977 Sep;90(3):422-30.
- [43] Solomons NW, Schoeller DA, Wagonfeld JB, Ott D, Rosenberg IH, Klein PD. Application of a stable isotope (13C)-labeled glycocholate breath test to diagnosis of bacterial overgrowth and ileal dysfunction. J Lab Clin Med. 1977 Sep;90(3):431-9.
- [44] Schneider JF, Schoeller DA, Nemchausky B, Boyer JL, Klein P. Validation of 13CO2 breath analysis as a measurement of demethylation of stable isotope labeled aminopyrine in man. Clin Chim Acta. 1978 Mar 1;84(1-2):153-62.
- [45] Schoeller DA, Schneider JF, Solomons NW, Watkins JB, Klein PD. Clinical diagnosis with the stable isotope 13C in CO2 breath tests: methodology and fundamental considerations. J Lab Clin Med. 1977 Sep;90(3):412-21.
- [46] Savarino V, Vigneri S, Celle G. The 13C urea breath test in the diagnosis of Helicobacter pylori infection. Gut. 1999 Jul;45 Suppl 1:I18-22.

- [47] Klein PD, Malaty HM, Martin RF, Graham KS, Genta RM, Graham DY. Noninvasive detection of Helicobacter pylori infection in clinical practice: the 13C urea breath test. Am J Gastroenterol. 1996 Apr;91(4):690-4.
- [48] de Meer K, Roef MJ, Kulik W, Jakobs C. In vivo research with stable isotopes in biochemistry, nutrition and clinical medicine: an overview. Isotopes in environmental and health studies. 1999;35(1-2):19-37.
- [49] Klein PD. 13C breath tests: visions and realities. The Journal of nutrition. 2001 May;131(5):1637S-42S.
- [50] Toskes PP, King CE, Spivey JC, Lorenz E. Xylose catabolism in the experimental rat blind loop syndrome: studies, including use of a newly developed d-[14C]xylose breath test. Gastroenterology. 1978 Apr;74(4):691-7.
- [51] King CE, Toskes PP. Comparison of the 1-Gram (14C)Xylose, 10-Gram Lactulose-H2, and 80-Gram Glucose-H2 breath tests in patients with small intestine bacterial overgrowth. Gastroenterology. 1986;91:1447-51.
- [52] King CE, Toskes PP, Guilarte TR, Lorenz E, Welkos SL. Comparison of the one gram d-[<sup>14</sup>C]xylose breath test to the [<sup>14</sup>C]bile acid breath test in patients with small-intestine bacterial overgrowth. Digestive diseases and sciences. 1980;25:53-8.
- [53] King CE, Toskes PP, Spivey JC, Lorenz E, Welkos S. Detection of small intestinal bacterial overgrowth by means of a <sup>14</sup>C-D-xylose breath test. Gastroenterology. 1979;77:75-82.
- [54] King CE, Toskes PP. Breath tests in the diagnosis of small intestine bacterial overgrowth. Critical reviews in clinical laboratory sciences. 1984;21(3):269-81.
- [55] Dellert SF, Nowicki MJ, Farrell MK, Delente J, Heubi JE. The 13C-xylose breath test for the diagnosis of small bowel bacterial overgrowth in children. J Pediatr Gastroenterol Nutr. 1997 Aug;25(2):153-8.
- [56] Stotzer PO, Kilander AF. Comparison of the 1-gram (14)C-D-xylose breath test and the 50-gram hydrogen glucose breath test for diagnosis of small intestinal bacterial overgrowth. Digestion. 2000;61(3):165-71.
- [57] Riordan SM, McIver CJ, Duncombe VM, Bolin TD, Thomas MC. Factors influencing the 1-g 14C-D-xylose breath test for bacterial overgrowth. Am J Gastroenterol. 1995 Sep;90(9):1455-60.
- [58] Rumessen JJ, Gudmand-Hoyer E, Bachmann E, Justesen T. Diagnosis of bacterial overgrowth of the small intestine. Comparison of the 14C-D-xylose breath test and jejunal cultures in 60 patients. Scand J Gastroenterol. 1985 Dec;20(10):1267-75.
- [59] Newcomer AD, McGill DB, Thomas PJ, Hofmann AF. Prospective comparison of indirect methods for detecting lactase deficiency. N Engl J Med. 1975 Dec 11:293(24):1232-6.
- [60] Casellas F, Malagelada JR. Applicability of short hydrogen breath test for screening of lactose malabsorption. Dig Dis Sci. 2003 Jul;48(7):1333-8.
- [61] Pohl D, Savarino E, Hersberger M, Behlis Z, Stutz B, Goetze O, et al. Excellent agreement between genetic and hydrogen breath tests for lactase deficiency and the role of extended symptom assessment. The British journal of nutrition. Apr 19:1-8.
- [62] Newcomer AD. Screening tests for carbohydrate malabsorption. J Pediatr Gastroenterol Nutr. 1984;3(1):6-8.

- [63] Douwes AC, Fernandes J, Degenhart HJ. Improved accuracy of lactose tolerance test in children, using expired H2 measurement. Arch Dis Child. 1978 Dec;53(12):939-42.
- [64] Kneepkens CM, Vonk RJ, Fernandes J. Incomplete intestinal absorption of fructose. Arch Dis Child. 1984 Aug;59(8):735-8.
- [65] Ravich WJ, Bayless TM, Thomas M. Fructose: incomplete intestinal absorption in humans. Gastroenterology. 1983 Jan;84(1):26-9.
- [66] Gudmand-Hoyer E, Krasilnikoff PA, Skovbjerg H. Sucrose-isomaltose malabsorption. Advances in nutritional research. 1984;6:233-69.
- [67] Davidson GP, Robb TA. Detection of primary and secondary sucrose malabsorption in children by means of the breath hydrogen technique. The Medical journal of Australia. 1983 Jul 9;2(1):29-32.
- [68] Hyams JS. Sorbitol intolerance: an unappreciated cause of functional gastrointestinal complaints. Gastroenterology. 1983 Jan;84(1):30-3.
- [69] Rhodes JM, Middleton P, Jewell DP. The lactulose hydrogen breath test as a diagnostic test for small-bowel bacterial overgrowth. Scand J Gastroenterol. 1979;14(3):333-6.
- [70] Diggory RT, Cuschieri A. The effect of dose and osmolality of lactulose on the oral-caecal transit time determined by the hydrogen breath test and the reproducibility of the test in normal subjects. Annals of clinical research. 1985;17(6):331-3.
- [71] Corazza GR, Strocchi A, Rossi R, Sirola D, Gasbarrini G. Sorbitol malabsorption in normal volunteers and in patients with coelaic disease. Gut. 1988;29:44-8.
- [72] Tursi A, Brandimarte G, Giorgetti GM. Sorbitol H<sub>2</sub>-breath test versus antiendomysium antibodies for the diagnosis of subclinical/silent coeliac disease. Scand J Gastroenterol. 2001;36:1170-2.
- [73] Tursi A, Brandimarte G, Giorgetti GM. H<sub>2</sub> breath test for the diagnosis of coeliac disease. American Journal of gastroenterology. 2002;97(10):2679-80.
- [74] Bond JH, Jr., Levitt MD. Use of pulmonary hydrogen (H 2) measurements to quantitate carbohydrate absorption. Study of partially gastrectomized patients. J Clin Invest. 1972 May;51(5):1219-25.
- [75] Cloarec D, Bornet F, Gouilloud S, Barry JL, Salim B, Galmiche JP. Breath hydrogen response to lactulose in healthy subjects: relationship to methane producing status. Gut. 1990 Mar;31(3):300-4.
- [76] Fritz M, Siebert G, Kasper H. Dose dependence of breath hydrogen and methane in healthy volunteers after ingestion of a commercial disaccharide mixture, Palatinit. The British journal of nutrition. 1985 Sep;54(2):389-400.
- [77] Calloway DH, Murphy EL. The use of expired air to measure intestinal gas formation. Annals of the New York Academy of Sciences. 1968 Feb 26;150(1):82-95.
- [78] Pitt P, de Bruijn KM, Beeching MF, Goldberg E, Blendis LM. Studies on breath methane: the effect of ethnic origins and lactulose. Gut. 1980 Nov;21(11):951-4.
- [79] Bond JH, Jr., Engel RR, Levitt MD. Factors influencing pulmonary methane excretion in man. An indirect method of studying the in situ metabolism of the methane-producing colonic bacteria. The Journal of experimental medicine. 1971 Mar 1;133(3):572-88.
- [80] Bjorneklett A, Jenssen E. Relationships between hydrogen (H2) and methane (CH4) production in man. Scand J Gastroenterol. 1982 Nov;17(8):985-92.

- [81] Craig RM, Atkinson AJ, Jr. D-xylose testing: a review. Gastroenterology. 1988 Jul;95(1):223-31.
- [82] Craig RM, Ehrenpreis ED. D-xylose testing. J Clin Gastroenterol. 1999 Sep;29(2):143-50.
- [83] Krawitt EL, Beeken WL. Limitations of the usefulness of the d-xylose absorption test. American journal of clinical pathology. 1975 Feb;63(2):261-3.
- [84] Gillberg R, Dotevall G, Kastrup W, Lindstedt G, Mobacken H, Swolin B.
- Conventional malabsorption tests: do they detect the adult patient with villous atrophy? Scandinavian journal of clinical and laboratory investigation. 1984 Feb;44(1):91-8.
- [85] Sladen GE, Kumar PJ. Is the xylose test still a worth-while investigation? Br Med J. 1973 Jul 28;3(5873):223-6.
- [86] Bode S, Gudmand-Hoyer E. The diagnostic value of the D-xylose absorption test in adult coeliac disease. Scand J Gastroenterol. 1987 Dec;22(10):1217-22.
- [87] Uil JJ, van Elburg RM, Mulder CJ, Heymans HS. The value of the D-xylose test compared with the differential sugar absorption test in recognizing coeliac disease. Neth J Med. 1996 Aug;49(2):68-72.
- [88] Johnstone TW. Pentosuria, chronic and alimentary. Edinburgh Med J. 1906;20:138-48.
- [89] Fordtran JS, Clodi PH, Soergel KH, Ingelfinger FJ. Sugar absorption tests, with special reference to 3-0-methyl-d-glucose and d-xylose. Ann Intern Med. 1962 Dec;57:883-91.
- [90] Rolston DDK, Mathan VI. Xylose transport in the human jejunum. Dig Dis Sci. 1989 1989;34(4):553-8.
- [91] McChance RA, Madders K. The comparative rates of absorption of sugars from the human intestine. Biochem J. 1930;24(3):795-804.
- [92] Fordtran JS, Soergel KH, Ingelfinger FJ. Intestinal absorption of D-xylose in man. N Engl J Med. 1962 Aug 9;267:274-9.
- [93] Pitkanen E. The conversion of D-xylose into D-threitol in patients without liver disease and in patients with portal liver cirrhosis. Clin Chim Acta. 1977 Oct 1;80(1):49-54
- [94] Segal S, Foley JB. The metabolic fate of C14 labeled pentoses in man. J Clin Invest. 1959;38:407-13.
- [95] Huguenin P, Cochet B, Balant L, Loizeau E. Test d'absorption du D-xylose. Schweiz Med Wochenschr. 1978;108(6):206-14.
- [96] Craig RM, Murphy P, Gibson TP, Quintanilla A, Chao GC, Cochrane C, et al. Kinetic analysis of D-xylose absorption in normal subjects and in patients with chronic renal failure. J Lab Clin Med. 1983;101(3):496-506.
- [97] Breiter HC, Craig RM, Levee G, Atkinson AJ, Jr. Use of kinetic methods to evaluate D-xylose malabsorption in patients. J Lab Clin Med. 1988 Nov;112(5):533-43.
- [98] Shamma'A MH, Ghazanfar SA. D-xylose test in enteric fever, cirrhosis, and malabsorptive states. Br Med J. 1960 Sep 17;2(5202):836-8.
- [99] Gardner FH. A malabsorption syndrome in military personnel in Puerto Rico. AMA. 1956 Jul;98(1):44-60.
- [100] Fourman LP. The absorption of xylose in steatorrhoea. Clin Sci (Lond). 1948 Feb 16;6(4):289-94.

- [101] Rinaldo JA, Jr. Interrelation of tests of intestinal absorption. Gastroenterology. 1961 Jan;40:86-93.
- [102] Butterworth CE, Jr., Perez-Santiago E, Martinez De Jesus J, Santini R. Studies on the oral and parenteral administration of D (+) xylose. N Engl J Med. 1959 Jul 23;261(4):157-64.
- [103] Finlay JM, Wightman KJ. The xylose tolerance test as a measure of the intestinal absorption of carbohydrate in sprue. Ann Intern Med. 1958 Dec;49(6):1332-47.
- [104] Gardner FH, Santiago EP. Oral absorption tolerance tests in tropical sprue. AMA. 1956 Oct;98(4):467-74.
- [105] Fowler D, Cooke WT. Diagnostic significance of d-xylose excretion test. Gut. 1960 Mar;1:67-70.
- [106] Brien FS, Turner DA, Watson EM, Geddes JH. A study of carbohydrate and fat absorption from the normal and diseased intestine in man. II. Changes in the serum lipids in man after the ingestion of butterfat with and without Tween 80 (sorlate). Gastroenterology. 1952 Feb;20(2):294-7.
- [107] Benson JA, Jr., Culver PJ, Ragland S, Jones CM, Drummey GD, Bougas E. The d-xylose absorption test in malabsorption syndromes. N Engl J Med. 1957 Feb 21;256(8):335-9.
- [108] Finlay JM, Hogarth J, Wightman KJ. A Clinical Evaluation of the D-Xylose Tolerance Test. Ann Intern Med. 1964 Sep;61:411-22.
- [109] Chanarin I, Bennett MC. Absorption of folic acid and D-xylose as tests of small-intestinal function. Br Med J. 1962 Apr 7;1(5283):985-9.
- [110] Christiansen PA, Kirsner JB, Ablaza J. D-Xylose and its use in the diagnosis of malabsorptive states. Am J Med. 1959 Sep;27:443-53.
- [111] Lanzkowsky P, Lloyd EA, Lahey ME. The Oral D-Xylose Test in Healthy Infants and Children. a Micro-Method of Blood Determination. Jama. 1963 Nov 2;186:517-9.
- [112] Hawkins KI. Pediatric xylose absorption test: measurements in blood preferable to measurements in urine. Clinical chemistry. 1970 Sep;16(9):749-52.
- [113] Kraut JR, Lloyd-Still JD. The 1-hr blood xylose test in the evaluation of malabsorption in infants and children. Am J Clin Nutr. 1980 Nov;33(11):2328-33.
- [114] Schaad U, Gaze H, Hadorn B. Value of 1-hour blood-xylose test in diagnosis of childhood coeliac disease. Arch Dis Child. 1978 May;53(5):420-2.
- [115] Rolles CJ, Kendall MJ. One hour blood D-xylose as a screening test for malabsorption in infants and young children. Arch Dis Child. 1972 Aug;47(254):673.
- [116] Buts JP, Morin CL, Roy CC, Weber A, Bonin A. One-hour blood xylose test: a reliable index of small bowel function. J Pediatr. 1978 May;92(5):729-33.
- [117] Kendall MJ, Nutter S. The influence of sex, body weight, and renal function on the xylose test. Gut. 1970 Dec;11(12):1020-3.
- [118] Worwag EM, Craig RM, Jansyn EM, Kirby D, Hubler GL, Atkinson AJ, Jr. D-xylose absorption and disposition in patients with moderately impaired renal function. Clinical pharmacology and therapeutics. 1987 Mar;41(3):351-7.
- [119] Mayersohn M. The "Xylose test" to assess gastrointestinal absorption in the elderly: a pharmacokinetic evaluation of the literature. Journal of gerontology. 1982 May;37(3):300-5.
- [120] Johnson SL, Mayersohn M, Conrad KA. Xylose disposition in humans as a function of age. Clinical pharmacology and therapeutics. 1986 Jun;39(6):697-702.

- [121] Kendall MJ. The influence of age on the xylose absorption test. Gut. 1970 Jun;11(6):498-501.
- [122] Beck I, Rona S, Callegarini U. The effect of gastric emptying on D(+)-xylose blood level time curve. Am J Dig Dis. 1962;7:928-35.
- [123] Cook GC. Breath hydrogen after oral xylose in tropical malabsorption. Am J Clin Nutr. 1980 Mar;33(3):555-60.
- [124] Casellas F, Chicharro L, Malagelada JR. Potential usefulness of hydrogen breath test with D-xylose in clinical management of intestinal malabsorption. Dig Dis Sci. 1993 Feb;38(2):321-7.
- [125] Casellas F, De Torres I, Malagelada JR. Follow-up of coeliac disease with D-xylose breath test. Dig Dis Sci. 1996 Oct;41(10):2106-11.
- [126] Skar V, Larsen S, Osnes M. The 1-g 14C-D-xylose breath test in gallstone patients with and without duodenal diverticula. Scand J Gastroenterol. 1985 May;20(4):447-51.
- [127] Nier AO. A redetermination of the relative abundances of the isotopes of carbon, nitrogen, and potassium. Physiol Rev. 1950;77:789.
- [128] Schoeller DA, Klein PD, Maclean WC, Watkins JB, Van Santen E. Fecal <sup>13</sup>C analysis for the detection and quantitation of intestinal malabsorption. J Lab Clin Med 1981;97:439-48.
- [129] Haycock GB, Schwartz GJ, Wisotsky DH. Geometric method for measuring body surface area: a height-weight formula validated in infants, children and adults. J Pediatr. 1978;93(1):62-6.
- [130] Larew LA, Johnson DC. Concentration dependence of the mechanism of glucose oxidation at gold electrodes in alkaline media. J Electroanal Chem. 1989;262(1-2):167-82.
- [131] Roe JH, Rice EW. Photometric method for determination of free pentose in animal tissues. JBiolChem. 1948;347:177.
- [132] Murray JA. The widening spectrum of coeliac disease. Am J Clin Nutr. 1999 Mar;69(3):354-65.
- [133] Koetse HA, Vonk RJ, Pasterkamp S, Pal J, de Bruijn S, Stellaard F. Variations in colonic H2 and CO2 production as a cause of inadequate diagnosis of carbohydrate maldigestion in breath tests. Scand J Gastroenterol. 2000 Jun;35(6):607-11.
- [134] Perman JA, Modler S, Olson AC. Role of pH in production of hydrogen from carbohydrates by colonic bacterial flora. Studies in vivo and in vitro. J Clin Invest. 1981 Mar;67(3):643-50.
- [135] Minocha A, Rashid S. Reliability and reproducibility of breath hydrogen and methane in male diabetic subjects. Dig Dis Sci. 1997 Mar;42(3):672-6.
- [136] King CE, Toskes PP, Spivey JC, Lorenz E, Welkos S. Detection of small intestine bacterial overgrowth by means of a 14C-D-xylose breath test. Gastroenterology. 1979 Jul;77(1):75-82.
- [137] Corazza G, Strocchi A, Sorge M, Bentai G, Gasbarrini G. Prevalence and consistency of low breath H2 excretion following lactulose ingestion. Possible implications for the clinical use of the H2 breath test. Dig Dis Sci. 1993 Nov;38(11):2010-6.
- [138] Vogelsang H, Ferenci P, Frotz S, Meryn S, Gangl A. Acidic colonic microclimate-possible reason for false negative hydrogen breath tests. Gut. 1988 Jan;29(1):21-6.

- [139] King CE, Toskes PP. Comparison of the 1-gram [14C]xylose, 10-gram lactulose-H2, and 80-gram glucose-H2 breath tests in patients with small intestine bacterial overgrowth. Gastroenterology. 1986 Dec;91(6):1447-51.
- [140] Simren M, Stotzer PO. Use and abuse of hydrogen breath tests. Gut. 2006 Mar;55(3):297-303.
- [141] Goldstein F, Karacadag S, Wirts CW, Kowlessar OD. Intraluminal small-intestinal utilization of d-xylose by bacteria. A limitation of the d-xylose absorption test. Gastroenterology. 1970 Sep;59(3):380-6.
- [142] Tursi A, Brandimarte G, Giorgetti G. High prevalence of small intestinal bacterial overgrowth in coeliac patients with persistence of gastrointestinal symptoms after gluten withdrawal. Am J Gastroenterol. 2003 Apr;98(4):839-43.
- [143] Rubio-Tapia A, Barton SH, Rosenblatt JE, Murray JA. Prevalence of small intestine bacterial overgrowth diagnosed by quantitative culture of intestinal aspirate in coeliac disease. J Clin Gastroenterol. 2009 Feb;43(2):157-61.
- [144] Usai P, Usai Satta P, Lai M, Corda MG, Piras E, Calcara C, et al. Autonomic dysfunction and upper digestive functional disorders in untreated adult coeliac disease. European journal of clinical investigation. 1997 Dec;27(12):1009-15.
- [145] Tursi A. Gastrointestinal motility disturbances in coeliac disease. J Clin Gastroenterol. 2004;38(8):642-5.
- [146] Benini L, Sembenini C, Salandini L, Dall OE, Bonfante F, Vantini I. Gastric emptying of realistic meals with and without gluten in patients with coeliac disease. Effect of jejunal mucosal recovery. Scand J Gastroenterol. 2001 Oct;36(10):1044-8.
- [147] Bjarnason I, Marsh MN, Price A, Levi AJ, Peters TJ. Intestinal permeability in patients with coeliac disease and dermatitis herpetiformis. Gut. 1985 Nov;26(11):1214-9.
- [148] Oktedalen O, Skar V, Dahl E, Serck-Hanssen A. Changes in small intestinal structure and function in HIV-infected patients with chronic diarrhoea. Scand J Infect Dis. 1998;30(5):459-63.
- [149] Hope HB, Tveito K, Aase S, Messelt E, Utzon P, Skar V. Small intestinal malabsorption in chronic alcoholism determined by 13C-D-xylose breath test and microscopic examination of the duodenal mucosa. Scand J Gastroenterol.45(1):39-45.
- [150] Bjarnason I, Marsh MN, Price A, Levi AJ, Peters TJ. Intestinal permeability in patients with coeliac disease and dermatitis herpetisformis. Gut. 1985;26:1214-9.
- [151] Tursi A, Brandimarte G, Giorgetti GM, Elisei W, Inchingolo CD, Monardo E, et al. Endoscopic and histological findings in the duodenum of adults with coeliac disease before and after changing to a gluten-free diet: a 2-year prospective study. Endoscopy. 2006 Jul;38(7):702-7.
- [152] Lee SK, Lo W, Memeo L, Rotterdam H, Green PH. Duodenal histology in patients with coeliac disease after treatment with a gluten-free diet. Gastrointestinal endoscopy. 2003 Feb;57(2):187-91.
- [153] Lanzini A, Lanzarotto F, Villanacci V, Mora A, Bertolazzi S, Turini D, et al. Complete recovery of intestinal mucosa occurs very rarely in adult coeliac patients despite adherence to gluten-free diet. Alimentary pharmacology & therapeutics. 2009 Jun 15;29(12):1299-308.
- [154] Kaukinen K, Peraaho M, Lindfors K, Partanen J, Woolley N, Pikkarainen P, et al. Persistent small bowel mucosal villous atrophy without symptoms in coeliac disease. Alimentary pharmacology & therapeutics. 2007 May 15;25(10):1237-45.

[155] Kaukinen K, Sulkanen S, Maki M, Collin P. IgA-class transglutaminase antibodies in evaluating the efficacy of gluten-free diet in coeliac disease. Eur J Gastroenterol Hepatol. 2002 Mar;14(3):311-5.

## 9 Errata

# Paper II:

Page 1292, Figure 2A

The correct number of healthy controls should be n = 43.

# 10 Original papers

Follow-up of coeliac disease with the novel one-hour <sup>13</sup>C-sorbitol breath

test versus the H2-sorbitol breath test

KARI TVEITO<sup>1</sup>, ANNE KRISTINE HETTA<sup>2</sup>, MIA ASKEDAL<sup>1</sup>, CATHRINE BRUNBORG<sup>3</sup>, LEIV

SANDVIK<sup>3</sup>, ELSE MARIT LØBERG<sup>4</sup>, VIGGO SKAR<sup>1</sup>

<sup>1</sup>Department of Medicine, Lovisenberg Diakonale Hospital, Oslo, Norway, <sup>2</sup>Department of Medicine,

Stavanger University Hospital, Stavanger, Norway, <sup>3</sup>Unit for epidemiology and biostatistics, Oslo

University Hospital - Ulleval, Oslo, Norway, <sup>4</sup>Department of Pathology, Oslo University Hospital-Ullevaal,

Oslo, Norway

Corresponding author:

Kari Tveito, M.D., Department of Medicine, Oslo University Hospital, Aker,

Trondheimsveien 235, N-0514 Oslo, Norway

Email: kari.tveito@oslo-universitetssykehus.no

Running head: <sup>13</sup>C-sorbitol breath test in follow-up of coeliac disease

**Abbreviations**: H<sub>2</sub>-SBT, hydrogen-sorbitol breath test; <sup>13</sup>C-SBT, <sup>13</sup>C-sorbitol breath test;

GFD, gluten-free diet; tTGA, tissue transglutaminase antibody

Key words: breath test, carbon isotopes, coeliac disease, malabsorption, sorbitol

1

#### **Abstract**

**Background:** We recently developed a <sup>13</sup>C-sorbitol breath test (<sup>13</sup>C-SBT) as an alternative to the H<sub>2</sub>-sorbitol breath test (H<sub>2</sub>-SBT) for coeliac disease. In this study we compared the diagnostic properties of the H<sub>2</sub>-SBT and the <sup>13</sup>C-SBT in follow-up of coeliac disease.

**Material and methods:** Twenty-seven coeliac patients on a gluten-free diet performed the breath tests. All had been tested before treatment in the intial study of the <sup>13</sup>C-SBT, in which 39 untreated coeliac patients, 40 patient controls, and 26 healthy volunteers participated.

5 g sorbitol and 100 mg  $^{13}$ C-sorbitol were dissolved in 250 ml tap water and given orally.  $H_2$ ,  $CH_4$  and  $^{13}CO_2$  were measured in end-expiratory breath samples every 30 minutes for four hours. Increased  $H_2$  concentration  $\geq$  20 ppm from basal values was used as cut-off for the  $H_2$ -SBT. 60 minutes values were used as diagnostic index in the  $^{13}$ C-SBT.

**Results:** <sup>13</sup>CO<sub>2</sub> levels at 60 minutes increased in 20/26 treated coeliac patients (77%) after GFD, but were significantly lower than in control groups. Out of 20 patients who had a positive H<sub>2</sub>-SBT before GFD, 12 had a negative H<sub>2</sub>-SBT after GFD. Peak H<sub>2</sub> concentrations were not correlated with <sup>13</sup>C-SBT results.

**Conclusions:** The study confirms the sensitivity of a one-hour <sup>13</sup>C-SBT for small intestinal malabsorption. The <sup>13</sup>C-SBT has superior diagnostic properties compared to the H<sub>2</sub>-SBT in follow-up of coeliac disease.

#### Introduction

Sorbitol is only partially absorbed in the small intestine, and its malabsorption is doseand concentration-dependent.[1-3] Corazza et al. were the first to show that administration of sorbitol at a low dose and concentration to patients with untreated coeliac disease resulted in increased excretion of hydrogen (H<sub>2</sub>) in breath compared to healthy controls.[4] Since then a series of studies have suggested high sensitivity of the H<sub>2</sub>-sorbitol breath test (H<sub>2</sub>-SBT) when screening for coeliac disease in adults.[5-8]

We recently developed a novel <sup>13</sup>C-sorbitol breath test, and compared it's diagnostic properties to that of the H<sub>2</sub>-SBT. We showed that a one-hour <sup>13</sup>C-sorbitol breath test had better sensitivity and specificity for coeliac disease versus healthy controls and patient controls than the H<sub>2</sub>-SBT. A diagnostic algorithm was proposed for the <sup>13</sup>C-sorbitol in which patients were stratified as being at high, moderate or low risk for coeliac disease. In this study we aimed to study the diagnostic properties of the <sup>13</sup>C-sorbitol breath test in follow up of treated coeliac patients compared to the H<sub>2</sub>-SBT. Coeliac patients who participated in the previous <sup>13</sup>C-sorbitol study were invited to participate.

#### Methods

Study subjects

Coeliac patients who had participated in our previous study of the <sup>13</sup>C-sorbitol breath test were invited to perform another breath test after approximately one year on a gluten-free diet. Twenty-seven out of 39 patients agreed to participate. Patients underwent gastroscopy with duodenal biopsies and routine laboratory testing. The pathologist reported the biopsies according to the 1992 Marsh criteria [9], as modified by Oberhuber et al. [10]. Serological testing for coeliac disease included measurement of IgA and IgG tissue-transglutaminase antibodies (tTGAs, U/ml), IgA gliadin antibodies (Units) and IgA levels (g/l). Patients were asked to range their symptoms of diarrhoea, constipation, bloating, abdominal pain, nausea, and fatigue on a 4-point Likert scale. A mean item score was used to provide a score from 1-4 (no discomfort to severe discomfort) for each dimension.

Patient controls (n = 40) and healthy controls (n = 26) were recruited in the previous study of the <sup>13</sup>C-SBT [11]. Coeliac disease had been excluded by duodenal biopsies in the patient control group. The majority of patients in this group suffered from irritable bowel syndrome (IBS).

The study was conducted in accordance with the Declaration of Helsinki-Tokyo and approved by the regional ethical committee. All subjects gave written consent to participate in the study.

### Breath test protocol

Test subjects were examined after an overnight fast of at least 10 hours. They were instructed to avoid high fibre food, and to consume a meal of rice with fish or meat the evening before the test. During the test period, all subjects continued fasting, were quietly seated and refrained from smoking., An oral load of 5 g sorbitol and 100 mg  $^{13}$ C-D-sorbitol, 99% (Aldrich, Milwaukee, USA), dissolved in 250 ml tap water, was given directly after obtaining four basal breath samples. Breath samples were collected at 30 minutes intervals for four hours. Duplicate end-expiratory breath samples were collected in 30 ml plastic syringes fitted with a T-piece and immediately analyzed for hydrogen (H<sub>2</sub>) and methane (CH<sub>4</sub>) by a QuinTron Model SC Microlyzer (QuinTron Instrument Company, Milwaukee, USA). Concentrations used for calculation were means of duplicate samples. Hydrogen increase  $\geq$  20 ppm from the lowest previous values (basal values) counted as a positive test.

Two parallel end-expiratory breath samples were collected via a straw in 12 ml tubes (Exetainers, Labco Limited, High Wycombe, UK) at every time point for analysis of <sup>13</sup>CO<sub>2</sub>. The <sup>13</sup>CO<sub>2</sub>/<sup>12</sup>CO<sub>2</sub> -ratio in the breath samples was determined by gas chromatography (GC) and continuous flow isotope ratio mass spectrometry (IRMS, Europe Scientific 20/20, Crewe, UK), and expressed as the percentage of <sup>13</sup>C-recovery (% of dose <sup>13</sup>C/h) for every time point. The formula of Schoeller et al. was used for this calculation.[12] The CO<sub>2</sub> production was assumed to be 300 mmol/m<sup>2</sup> body surface area per hour. Body surface area was calculated by the weight-height formula of Haycock et al. [13].

#### Statistics

Data from patient characteristics are outlined in numbers and percentages. Mean with standard deviation (SD) or median with range are presented, as appropriate.

Comparison between independent groups was made using either independent sample Student's *t*-test or one-way ANOVA for normally distributed variables, and Mann Whitney U test or Kruskal-Wallis test for continuous variables with skewed distribution. The paired Student's *t*-test was used for comparison of variables before and after glutenfree treatment. Linear regression was used when comparing two continuous variables. Spearman's correlation coefficient was used to test for correlation between variables. When comparing dichotomised variables, a chi-square test was used. Findings with *p* values < 0.5 were considered significant. All statistical analyses were performed using SPSS software package version 18.0 (SPSS Inc., Chicago, Illinois, USA).

#### Results

Subject characteristics

The 39 coeliac patients enrolled in the study had Marsh IIIa-c villous atrophy in small intestinal biopsies when they were diagnosed with coeliac disease. Twenty-seven patients agreed to participate in the follow-up study after treatment with gluten-free diet (median duration 12 months, range 6-24 months). Twenty patients had normal duodenal biopsies by this time. Four biopsies were classified as Marsh I, one was classified as Marsh IIIa, and two patients refused to have a new biopsy. Patient controls were described in detail in a previous article [11]. Patients and healthy control characteristics are summarised in table I.

#### H<sub>2</sub>-sorbitol breath test results

Positive H<sub>2</sub>-sorbitol breath tests were obtained in 27/38 untreated coeliac patients (71%). After gluten-free diet, 17/27 coeliac patients (63%) had a negative H<sub>2</sub>-sorbitol breath test, while 10/27 coeliac patients (37%) had a positive H<sub>2</sub>-sorbitol breath test (Figure 1). One of the intital H<sub>2</sub>-sorbitol breath tests from the 27 coeliac patients tested twice was discarded due to technical problems. Twenty of the remaining 26 breath tests were positive before GFD. After GFD, twelve of these patients had a positive H<sub>2</sub>-sorbitol breath test (60%), and eight were negative (40%). Out of the six coeliac patients who had negative H<sub>2</sub>-sorbitol breath tests before treatment, five also had a negative test after GFD, but one patient had a positive breath test after GFD. As shown in the previous study,

30/40 patient controls (75%), and 14/26 healthy subjects (54%) had a positive H<sub>2</sub>-sorbitol breath test.

Mean peak  $H_2$  concentration was  $37 \pm 37$  ppm in coeliac patients,  $25 \pm 28$  ppm in treated coeliac patients,  $42 \pm 32$  ppm in patient controls, and  $28 \pm 24$  ppm in healthy controls. Mean time to reach peak  $H_2$  concentration was  $188 \pm 49$  min in coeliac patients,  $190 \pm 48$  min in treated coeliac patients,  $178 \pm 58$  min in patient controls, and  $190 \pm 49$  min in healthy controls. Peak  $H_2$  concentration differed significantly between treated coeliac patients and patient controls (p = 0.01), but not between other groups. Time to reach peak  $H_2$  concentration did not differ significantly between groups.

### <sup>13</sup>C-sorbitol breath test results

Mean breath <sup>13</sup>CO<sub>2</sub> levels were significantly higher in the 26 treated coeliac patients (one test was discarded due to technical problems) than in untreated coeliac patients at all time points before 240 minutes, and significantly lower in treated coeliac patients than in control subjects without coeliac disease at all time points exept 30 minutes (Figure 2). On an individual level, <sup>13</sup>CO<sub>2</sub> levels at 60 minutes increased in 20/26 treated patients (77%).

In our previous study of the  $^{13}$ C-sorbitol breath test, 60 minutes  $^{13}$ CO<sub>2</sub> levels in breath were chosen as the diagnostic test variable, and the following diagnostic algorith proposed: Excreted dose of  $^{13}$ C/h at 60 minutes: < 2% = high probability for coeliac disease, 2-4% = moderate probability for coeliac disease, and > 4% = low probability for coeliac disease. Sixteen of the treated coeliac patients (59%) had a 60 minutes  $^{13}$ CO<sub>2</sub> level < 2% before treatment.  $^{13}$ CO<sub>2</sub> levels increased above the 2% cut-off in 12/16 patients (75%, Figure 3). The patient with the lowest level of exhaled  $^{13}$ CO<sub>2</sub> at 60 minutes

had Marsh 3a villous atrophy in the control biopsy performed after GFD. The other three patients with excreted dose of  $^{13}$ C/h at 60 minutes <2% had normal control duodenal biopsies. Seventeen treated patients (63%) excreted between 2-4% of ingested dose of  $^{13}$ C after 60 minutes, and five patients (19%) had  $^{13}$ CO<sub>2</sub> levels > 4% of ingested dose.

Comparison of breath tests and correlation with clinical variables

No significant association was found between peak  $H_2$  concentrations and  $^{13}CO_2$  values at 60 minutes in any of the groups. In the coeliac group there was an inverse association between  $^{13}CO_2$  values at 60 minutes and age (p = 0.037), which was not found in the other groups. Age was inversely associated with peak  $H_2$  concentration in the treated coeliac group (p = 0.007), but not in the other groups. Duration of gluten-free diet was not significantly associated with  $^{13}CO_2$  values at 60 minutes or to peak  $H_2$  concentrations.

IgA tTGA levels were inversely associated to 60 minutes  $^{13}$ CO<sub>2</sub> values (p = 0.025) in the untreated coeliac group, but there was no association in the treated coeliac group. Peak H<sub>2</sub> concentrations were not associated to IgA tTGA levels in any of the groups. In the coeliac group, peak H<sub>2</sub> concentrations increased significantly with increasing severity of duodenal lesions, classified from Marsh IIIa to Marsh IIIc (p < 0.001). IgA tTGA levels increased and  $^{13}$ CO<sub>2</sub> excretion rates at 60 minutes decreased from Marsh IIIa to Marsh IIIc, but these associations did not reach significance.

Symptom scores decreased significantly for all symptom variables except bloating after GFD (Table 2). The 60 minutes variable after GFD was inversely associated to fatigue after GFD (p = 0.039), but not to other symptom scores.

S-folate and s-vitamin  $B_{12}$  increased significantly after GFD (s-folate: mean difference 4,44 nmol/L, p=0.001, and vitamin  $B_{12}$ : mean difference 55,3 pmol/L, p<0.001). S-ferritin, but not b-haemoglobin (b-Hb), increased significantly in both gender after GFD (women: mean difference 20,1 mikrogram/L, p=0.038, men: mean difference 71,2 mikrogram/L, p=0.001). In the treated coeliac group,  $^{13}CO_2$  excretion rates at 60 minutes were not associated with levels of s-folat, s-vitamin  $B_{12}$ , b-Hb and s-ferritin. Peak  $H_2$  concentration in the treated group was positively associated to b-Hb in men (n=11, p=0.018), but not in women. There was no association between peak  $H_2$  concentration and s-folat, s-vitamin  $B_{12}$  or s-ferritin in the treated group.

#### Discussion

This first study of the <sup>13</sup>C-sorbitol breath test in patients with treated coeliac disease, confirms the sensitivity of the <sup>13</sup>C-sorbitol breath test for small intestinal malabsorption. Seventy-seven percent of coeliac patients tested both before and after gluten-free diet increased their <sup>13</sup>C-sorbitol breath test score after GFD. The H<sub>2</sub> breath test with a cut-off level at 20 ppm had a sensitivity of 63% and a specificity of 71% for treated versus untreated coeliac patients.

The improvement in breath test results was accompagnied by normalization of duodenal biopsies in all but four patients, significant decline in symptom scores and increased levels of s-ferritin, s-folate and s-vitamin B<sub>12</sub>. Generally, laboratory results were not associated with breath test results, as in our previous study of the <sup>13</sup>C-sorbitol breath test and the H<sub>2</sub>-sorbitol breath test in untreated coeliac disease. That increasing peak H<sub>2</sub> concentration was associated with increasing b-Hb in treated male coeliac patients was unexpected, and may be due to a type I statistical error.

There was a significant association between peak H<sub>2</sub> concentration and increasing histopathology in the untreated coeliac group. A parallell, but non-significant trend was seen for <sup>13</sup>CO<sub>2</sub> excretion rates at 60 minutes. Furthermore, a non-significant increase in IgA tTGA levels from Marsh IIIa-IIIc was seen. Our results are in line with studies by Tursi et al., who found higher serum values of IgA tTGA with increasing histopathology in untreated coeliac disease, and a strict correlation between H<sub>2</sub>-sorbitol breath test cut-off values (in ppm and minutes) and histological lesions [14, 15]. However, <sup>13</sup>CO<sub>2</sub> excretion rates remained lower in treated coeliac patients with normal duodenal biopsies than healthy controls and patients without coeliac disease. Furthermore, 40% of the

patients who had a positive H<sub>2</sub>-sorbitol breath test before GFD, also had a positive H<sub>2</sub>-sorbitol breath test after GFD. These results are in line with a report from Bjarnason et al. who, using the <sup>51</sup>Cr EDTA absorption test, found persistent small intestine permeability in treated coeliac patients despite small intestinal mucosal healing [16]. Likewise, studies of patients with HIV and diarrhoea [17], and a study of patients with high alcohol consumption and diarrhoea [18], have shown <sup>13</sup>C-xylose breath test results compatible with small intestinal malabsorption despite light microscopically normal duodenal mucosa. Electron microscopic studies of the two latter patient groups revealed subtle architectural damage of the small intestinal mucosa. Similar minor small intestinal histopathology may persist also in treated coeliac patients. We also cannot exclude, as Bjarnason et al. proposed, the possibility of an inherent increase in small intestinal mucosal permeability in coeliac patients. Further studies are needed to investigate this issue and to determine the clinical importance of the findings.

In a kinetic study of D-xylose absorption, Breiter et al. found that total breath H<sub>2</sub> excretion was not correlated with the extent of D-xylose absorption [19]. Likewise, we could not show any correlation between the results of the <sup>13</sup>C-sorbitol breath test and peak H<sub>2</sub> excretion. H<sub>2</sub> production in the large intestine due to small intestinal malabsorption of carbohydrate has been showed to vary considerably between individuals, and also interindividually over time [20-22]. This may explain the lower sensitivity of the H<sub>2</sub>-sorbitol breath test compared to the <sup>13</sup>C-sorbitol breath test, and also the lack of correlation between the two breath tests. Our previous study of the H<sub>2</sub>-sorbitol breath test showed very low specificity of the H<sub>2</sub>-sorbitol breath test for untreated coeliac disease versus patients with gastrointestinal symptoms without coeliac disease

and healthy control subjects [11]. The present study shows that the H<sub>2</sub>-sorbitol breath test is also not ideally suited for follow-up of coeliac disease.

Some studies have showed changes in intestinal motility after treatment of coeliac disease with GFD [23-25]. Time to reach peak H<sub>2</sub> concentration was unchanged after GFD in our study. If there had been a significant change in motility with a gluten-free diet, we would expect a difference in rate of delivery of sorbitol to the large intestine and a change in time to reach peak H<sub>2</sub> concentration. Our study did not provide such evidence, and we believe that the changes in <sup>13</sup>CO<sub>2</sub> excretion rate after GFD is mainly due to increased small intestinal sorbitol absorption capacity.

As expected, coeliac patients reported reduced gastrointestinal symptoms after GFD. Interestingly, the fatigue score after GFD was inversely associated to <sup>13</sup>C-sorbitol breath test results. Studies of patients with untreated coeliac disease have not showed any correlation between symptom presentation and severity of mucosal damage [26]. Further studies are needed to examine the possibility of a correlation of fatigue in patients with treated coeliac disease and persistently reduced small intestinal absorption capacity.

In conclusion, we recommend that the H<sub>2</sub>-sorbitol breath test be abandoned due to low specificity versus patients with other gastrointestinal disorders such as IBS, and versus healthy controls. The novel <sup>13</sup>C-sorbitol breath test shows superior diagnostic properties to the H<sub>2</sub>-sorbitol breath test for diagnosis and follow-up of coeliac disease.

**Acknowledgements** Kari Tveito was supported by a grant from Eastern Norway Regional Health Authority.

#### References

- [1] Hyams JS. Sorbitol intolerance: an unappreciated cause of functional gastrointestinal complaints. Gastroenterology. 1983 Jan;84(1):30-3.
- [2] Jain NK, Patel VP, Pitchumoni CS. Sorbitol intolerance in adults. Prevalence and pathogenesis on two continents. J Clin Gastroenterol. 1987 Jun;9(3):317-9.
- [3] Rumessen JJ, Gudmand-Hoyer E. Malabsorption of fructose-sorbitol mixtures. Interactions causing abdominal distress. Scand J Gastroenterol. 1987 May;22(4):431-6.
- [4] Corazza GR, Strocchi A, Rossi R, Sirola D, Gasbarrini G. Sorbitol malabsorption in normal volunteers and in patients with coeliac disease. Gut. 1988 Jan;29(1):44-8.
- [5] Pelli MA, Capodicasa E, De Angelis V, al. e. Sorbitol H2-breath test in celiac disease. Importance of early positivity. Gastroenterology International. 1998;11(2):65-8.
- [6] Tursi A, Brandimarte G, Giorgetti GM. Sorbitol H2-breath test versus antiendomysium antibodies for the diagnosis of subclinical/silent coeliac disease. Scand J Gastroenterol. 2001 Nov;36(11):1170-2.
- [7] Tursi A, Brandimarte G, Giorgetti GM. H<sub>2</sub> breath test for the diagnosis of celiac disease. American Journal of gastroenterology. 2002;97(10):2679-80.
- [8] Tursi A, Brandimarte G, Giorgetti GM, Inchingolo CD. Effectiveness of the sorbitol H2 breath test in detecting histological damage among relatives of coeliacs. Scand J Gastroenterol. 2003;38(7):727-31.
- [9] Marsh MN. Gluten, major histocompatibility complex, and the small intestine. A molecular and immunobiologic approach to the spectrum of gluten sensitivity ('celiac sprue'). Gastroenterology. 1992;102(1):330-54.
- [10] Oberhuber G, Granditsch G, Vogelsang H. The histopathology of coeliac disease: time for a standardized report scheme for pathologists. European Journal of Gastroenterology & Hepatology. 1999;11(10):1185-94.
- [11] Tveito K, Hetta AK, Askedal M, Brunborg C, Sandvik L, Loberg EM, et al. A novel one-hour 13C-sorbitol breath test versus the H2-sorbitol breath test for assessment of coeliac disease. Scand J Gastroenterol. 2009;44(7):813-9.
- [12] Schoeller DA, Klein PD, Maclean WC, Watkins JB, Van Santen E. Fecal <sup>13</sup>C analysis for the detection and quantitation of intestinal malabsorption. J Lab Clin Med 1981;97:439-48.
- [13] Haycock GB, Schwartz GJ, Wisotsky DH. Geometric method for measuring body surface area: a height-weight formula validated in infants, children and adults. J Pediatr. 1978;93(1):62-6.
- [14] Tursi A, Brandimarte G, Giorgetti GM. Prevalence of antitissue transglutaminase antibodies in different degrees of intestinal damage in celiac disease. J Clin Gastroenterol. 2003;36:219-21.
- [15] Tursi A, Brandimarte G, Giorgetti GM. Sorbitol H2-breath test versus antiendomysium antibodies to assess histological recovery after gluten-free diet in coeliac disease. Dig Liver Dis. 2002 Dec;34(12):846-50.

- [16] Bjarnason I, Marsh MN, Price A, Levi AJ, Peters TJ. Intestinal permeability in patients with coeliac disease and dermatitis herpetiformis. Gut. 1985 Nov;26(11):1214-9.
  [17] Oktedalen O, Skar V, Dahl E, Serck-Hanssen A. Changes in small intestinal structure and function in HIV-infected patients with chronic diarrhoea. Scand J Infect Dis. 1998;30(5):459-63.
- [18] Hope HB, Tveito K, Aase S, Messelt E, Utzon P, Skar V. Small intestinal malabsorption in chronic alcoholism determined by 13C-D-xylose breath test and microscopic examination of the duodenal mucosa. Scand J Gastroenterol.45(1):39-45.
- [19] Breiter HC, Craig RM, Levee G, Atkinson AJ, Jr. Use of kinetic methods to evaluate D-xylose malabsorption in patients. J Lab Clin Med. 1988 Nov;112(5):533-43.
- [20] Koetse HA, Vonk RJ, Pasterkamp S, Pal J, de Bruijn S, Stellaard F. Variations in colonic H2 and CO2 production as a cause of inadequate diagnosis of carbohydrate maldigestion in breath tests. Scand J Gastroenterol. 2000 Jun;35(6):607-11.
- [21] Perman JA, Modler S, Olson AC. Role of pH in production of hydrogen from carbohydrates by colonic bacterial flora. Studies in vivo and in vitro. J Clin Invest. 1981 Mar;67(3):643-50.
- [22] Minocha A, Rashid S. Reliability and reproducibility of breath hydrogen and methane in male diabetic subjects. Dig Dis Sci. 1997 Mar;42(3):672-6.
- [23] Usai P, Usai Satta P, Lai M, Corda MG, Piras E, Calcara C, et al. Autonomic dysfunction and upper digestive functional disorders in untreated adult coeliac disease. European journal of clinical investigation. 1997 Dec;27(12):1009-15.
- [24] Tursi A. Gastrointestinal motility disturbances in celiac disease. J Clin Gastroenterol. 2004;38(8):642-5.

disease. Dig Liver Dis. 2007 Jan;39(1):26-9; discussion 30-2.

[25] Benini L, Sembenini C, Salandini L, Dall OE, Bonfante F, Vantini I. Gastric emptying of realistic meals with and without gluten in patients with coeliac disease.
Effect of jejunal mucosal recovery. Scand J Gastroenterol. 2001 Oct;36(10):1044-8.
[26] Brar P, Kwon GY, Egbuna, II, Holleran S, Ramakrishnan R, Bhagat G, et al. Lack of correlation of degree of villous atrophy with severity of clinical presentation of coeliac

Table I. Clinical characteristics of study patients.

	Untreated coeliac patients,	Treated coeliac patients, n = 27	Patient controls, n = 40	Healthy control subjects, n = 26
	n = 39	patients, ii 27	10	subjects, ii 20
Age, median	36 (19-82)	44 (20-83)	29 (17-65)	30 (22-60)
(range)				
Female/Male	24/15	15/12	26/14	18/8
BMI, mean (SD)	22.9 (3.6)	23.7 (3.4)	22.3 (2.8)	22.3 (2.5)
Small bowel				
histology:				
Marsh 0	0	20	36	
Marsh I	0	4	4	
Marsh IIIa	15	1	0	
Marsh IIIb	12	0	0	
Marsh IIIc	12	0	0	
IgA tTGA > 5U/ml	36	2	5	0
IgA tTGA 2-5U/ml	0	6	1	0
IgA tTGA < 2U/ml	3	19	0	26

BMI, body mass index, kg/m<sup>2</sup>; IgA tTGA, IgA tissue transglutaminase antibody

Table II. Symptoms graded 1-4 (none-mild-moderate-severe) in coeliac patients tested twice.

Symptoms	n	Untreated coeliac patients Mean (SD)	Treated coeliac patients Mean (SD)	<i>p</i> -value
D: 1	27	2.7 (1.0)	1.6.(1.0)	0.001
Diarrhoea	27	2.7 (1.0)	1.6 (1.0)	0.001
Bloating	22	2.3 (0.9)	2.1 (1.1)	0.55
Abdominal pain	26	2.9 (1.1)	1.4 (0.7)	0.001
Nausea	26	3.3 (1.1)	1.2 (0.5)	< 0.001
Constipation	27	2.9 (1.3)	1.2 (0.5)	< 0.001
Fatigue	20	2.7 (1.1)	1.8 (1.1)	0.03

n; number of individuals in whom the paired t-test was performed.

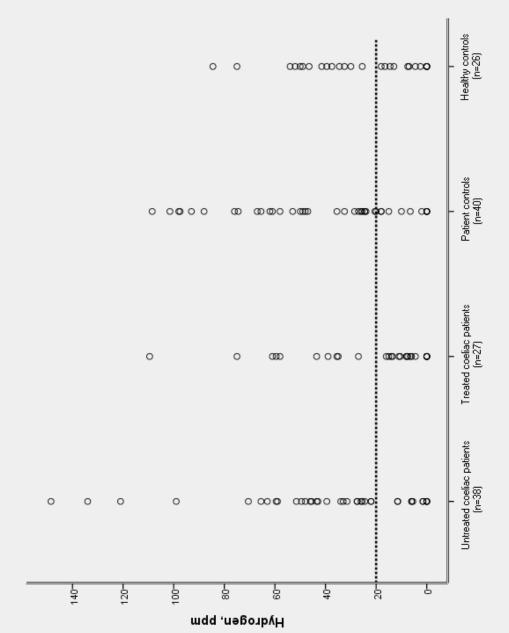


Figure 1. Scatterplot of peak H<sub>2</sub> concentrations in the four study groups. Dottet line represents diagnostic cut-off value.

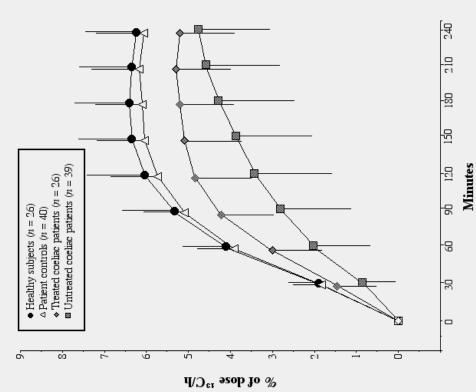


Figure 2.  $^{13}$ C-sorbitol breath test: Mean (SD)  $^{13}$ CO $_2$  excretion in study subjects measured every 30 minutes for 240 minutes.

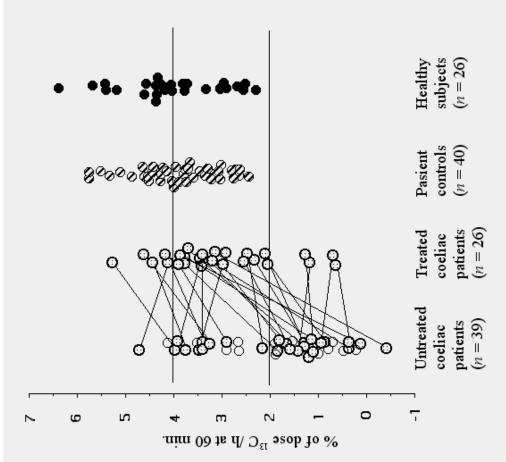


Figure 3.  $^{13}$ C-sorbitol breath test: Individual  $^{13}$ CO<sub>2</sub> levels in breath after 60 minutes in study subjects. Horizontal lines represent cut-off values in the diagnostic algorithm.