Brain energy rescue: an emerging therapeutic concept for neurodegenerative disorders of ageing

Stephen C. Cunnane^{1†}, Eugenia Trushina², Cecilie Morland³, Alessandro Prigione⁴, Gemma Casadesus⁵, Zane B. Andrews⁶, Flint M. Beal⁷, Linda H. Bergersen⁸, Roberta D. Brinton⁹, Suzanne de la Monte¹⁰, Anne Eckert¹¹, Jenny Z. Harvey¹², Ross Jeggo¹³, Jack H. Jhamandas¹⁴, Oliver Kann¹⁵, Clothide Mannoury la Cour¹³, William F. Martin¹⁶, Gilles Mithieux¹⁷, Paula I. Moreira¹⁸, Michael P. Murphy¹⁹, Klaus-Armin Nave²⁰, Tal Nuriel²¹, Stéphane H.R. Oliet²², Frédéric Saudou²³, Mark P. Mattson²⁴, Russell H. Swerdlow²⁵ and Mark J. MIllan^{13,26†}

¹Dept Medicine, Université de Sherbrooke and Research Center on Aging, Sherbrooke, QC, Canada.

²Dept of Neurology, Mayo Clinic, 200 First Street SW, Rochester, MN USA.

³Oslo, Dept Pharmaceutical Biosciences, Institute of Pharmacy, University of Oslo, Norway.

⁴Dept of General Pediatrics, Neonatology, and Pediatric Cardiology, University of Dusseldorf, Dusseldorf, Germany.

⁵Dept Biological Sciences, Kent State University, Kent, OH USA.

⁶Monash Biomedicine Discovery Institute and Dept of Physiology, Monash University, Clayton, Victoria 3800, Australia.

⁷Dept Neurology, Weill Cornell Medicine, New York, NY USA.

⁸Dept Anatomy, Institute of Basic Medical Sciences, University of Oslo, Oslo, Norway.

⁹College of Medicine, University of Arizona, Tucson, USA.

¹⁰Brown Medical School, Rhode Island Hospital, Providence, RI USA.

¹¹Psychiatric University Clinic, Basel, Switzerland.

¹²Ninewells Hospital and Medical School, University of Dundee, Dundee UK.

¹³Division of Neuropsychiatry, Institut de recherche Servier, Croissy sur Seine, France.

¹⁴Dept of Medicine and Neuroscience and Mental Health Institute, University of Alberta, Edmonton, AB, Canada.

¹⁵Institute of Physiology and Pathophysiology, University of Heidelberg, Heidelberg, Germany.

¹⁶Institute of Molecular Evolution, University of Dusseldorf, Dusseldorf, Germany.

¹⁷Lyon Est University and INSERM U1213, Lyon, France.

¹⁸Center for Neuroscience and Cell Biology and Faculty of Medicine, University of Coimbra, Portugal.

¹⁹MRC Mitochondrial Biology Unit, University of Cambridge, Cambridge UK.

²⁰Dept Biosciences, University of Heidelberg, Heidelberg, Germany.

²¹Columbia University Medical Center, New York, NY USA.

²²Neurocentre Magendie, INSERM U1215 and Université de Bordeaux, Bordeaux, France.

²³ University of Grenoble Alpes, INSERM, U1216, CHU Grenoble Alpes, Grenoble Institute Neurosciences, Grenoble, France.

²⁴Dept of Neuroscience, Johns Hopkins University School of Medicine, Baltimore, MD 21205 USA.
 ²⁵University of Kansas Alzheimer's Disease Center, 4350 Shawnee Mission Parkway, Fairway, KS
 66205 USA.

²⁶These authors contributed equally to this work, and are equal first authors: Cunnane, S.C., Millan, M.J.

te-mails: stephen.cunnane@usherbrooke.ca; mark.millan@servier.com

Abstract | The brain requires a continuous supply of energy in the form of ATP, most of which is produced from glucose by oxidative phosphorylation in mitochondria, complemented by aerobic glycolysis in the cytoplasm. When glucose levels are limited, ketone bodies generated in the liver and lactate derived from exercising skeletal muscle can also become important energy substrates for the brain. In neurodegenerative disorders of ageing, brain glucose metabolism deteriorates in a progressive, region-specific and disease-specific manner — a problem that is best characterized in Alzheimer disease, where it begins pre-symptomatically. This Review discusses the status and prospects of therapeutic strategies for countering neurodegenerative disorders of ageing by rescuing, protecting or normalizing brain energetics. Approaches described include restoring oxidative phosphorylation and glycolysis, improving insulin sensitivity, correcting mitochondrial dysfunction, ketone-based interventions, acting via hormones that modulate cerebral energetics, RNA therapeutics and complementary multi-modal lifestyle changes.

[H1] Introduction

Increased longevity over the past 50 years has contributed to a rising prevalence of neurodegenerative disorders of ageing (NDAs), particularly Alzheimer disease (AD) and Parkinson disease (PD). These disorders are a major socioeconomic and medical challenge with little prospect of a solution so far. Some drugs provide a degree of symptomatic relief, but disease-modifying treatments for NDAs remain elusive despite concerted attempts to counter the pathological processes of neurotoxic protein accumulation, neuroinflammation [G], axonal or synaptic dysfunction and neuronal death^{1,2}.

Since the concept was first reported 40 years ago, evidence has been accumulating that impaired brain energetics is involved in the aetiology and progression of NDAs, especially AD³⁻⁷. Brain energy metabolism declines subtly during ageing and is frequently present before diagnosis of NDA; it both drives and is driven by functional impairment and neurodegeneration in a destructive cycle^{3,6}.

Accordingly, a broad range of 'brain energy rescue' strategies have recently been explored and are the focus of this Review. These strategies aim to impede the onset and progress of NDAs by improving, preserving and/or restoring brain energy status. We first summarize how fuel is supplied and used across various cell types in the brain, including central, peripheral and endocrine mechanisms that modulate brain energy homeostasis as well as cognitive and neuronal function. The core features of disrupted brain energetics in the five main NDAs — AD, PD, Huntington disease (HD), frontotemporal dementia (FTD) and amyotrophic lateral sclerosis (ALS) — are then outlined. This forms the basis for the brain energy rescue strategies reported in preclinical and clinical studies, including promoting mitochondrial function, alternative fuel sources such as ketones [G], hormonal interventions to improve insulin sensitivity and brain glucose metabolism, and complementary lifestyle approaches. Finally, we highlight genetic and other emerging approaches to enhance and restore brain energetics and we consider the challenges of translating promising preclinical results towards the dual goals of symptom relief and disease modification in NDAs.

[H1] Brain energetics

Despite representing just over 2% of adult body weight, the human brain accounts for 20% of the body's total energy requirement⁸. The brain's main competitors for energy are the liver, kidneys and heart, which have as high or higher rates of energy consumption per gram, but their overall

energy consumption is lower than the brain's. The immune system also consumes considerable energy, especially when activated in NDAs⁹. Brain energy metabolism is influenced by the endocrine modulation of appetite and whole-body energetics — processes that are compromised during healthy ageing and in NDAs (Supplementary Table 1)¹⁰. Food intake, glucose-sensing mechanisms and energy homeostasis are themselves regulated by a complex set of neural networks that in turn modulate autonomic function, appetite, reward and executive functioning. These aspects are beyond the scope of this Review, and have been reviewed elsewhere^{11,12}.

[H2] ATP

ATP is the main currency of brain energy metabolism. Most ATP is used by Na⁺/K⁺-ATPase and Ca²⁺-ATPase, the cell membrane pumps that reset ion gradients during neuronal signaling^{8,13,14}. Excitatory (glutamatergic) neurons consume 80–85% of the brain's ATP, with inhibitory neurons and glial cells accounting for the remainder¹⁵⁻¹⁷. Although non-signalling pathways and cellular processes, including axonal transport, maintenance of cytoskeletal architecture, proton leakage, microglial motility, DNA repair, RNA translation and phospholipid remodelling consume less energy than neurotransmission, the energetic requirements of these processes remain hard to define^{13,16}. ATP is also a neurotransmitter released from neurons, astrocytes and microglia **[G]**¹⁸.

[H2] Glucose

Glucose accounts for $\ge 95\%$ of ATP production in the brain⁸. Within the brain, glucose uptake is orchestrated across several cell types collectively known as the neurovascular unit: brain capillary endothelial cells, pericytes, astrocytes, oligodendrocytes [G], microglia and neurons — the final beneficiary of glucose uptake (Figure 1)^{19,20}. The normally tight spatial and temporal association of local blood flow, oxygen consumption and glucose consumption in the brain is termed neurovascular coupling [G], and is the basis for functional magnetic resonance imaging²⁰.

Brain uptake of glucose from the circulation is driven by the energy demand of activated neurons, not by the level of circulating glucose. Indeed, under normal conditions, the capacity to transport glucose into the brain exceeds the brain's energy requirement by 2–3-fold¹⁵. Simply put, glucose is actively 'pulled' into an area of the brain in response to increased local neuronal activity. Glucose transport is achieved by the coordinated activity of glucose transporters on the capillary endothelium (GLUT1) and plasma membrane of astrocytes (GLUT1, GLUT2, GLUT7), oligodendrocytes (GLUT1) and neurons (GLUT3 as well as GLUT4) in the cortex, hippocampus and

cerebellum^{10,17,21}. Only GLUT4 is mobilized as a direct response to sustained synaptic activity; its membrane insertion is stimulated by the metabolic sensor, AMP-activated protein kinase (AMPK)¹⁷. Membrane translocation of GLUT4 is insulin-dependent in muscle and adipose tissue and probably also in neurons, so insulin resistance **[G]**, as occurs in NDAs, is characterised by reduced neuronal glucose uptake^{17,22}.

To reach neurons from capillaries, blood glucose either diffuses directly through the extracellular space or is channeled through astrocytes *via* their end-feet, which surround the capillary walls. It is taken up by astrocytic GLUT1 and exits through GLUT1 on perisynaptic processes adjacent to neurons and oligodendrocytes (Figure 1). Some glucose that enters astrocytes is metabolized to ATP and some is converted to lactate, which can act both as a neurotransmitter (discussed below) and as an alternative energy source¹⁸.

[H2] Energy use by brain cells

The ATP required by neurons is predominantly generated within mitochondria by oxidative phosphorylation [G] of glucose via the tricarboxylic acid cycle [G] (TCA cycle; also known as the citric acid or Krebs cycle; Box 1)¹⁴. Additional ATP is generated by aerobic glycolysis [G] in the cytoplasm, which is required to support the high energy demands of synaptic transmission¹⁷. Glutamate is the neurotransmitter in most excitatory neurons and is recycled by astrocytes and delivered back to neurons as glutamine for reconversion into glutamate or (to a lesser degree) for use in energy generation by the TCA cycle^{13,23}. Compared to astrocytes, neurons favour oxidative phosphorylation over aerobic glycolysis, have more-rapid TCA cycling and contain less phosphorylated pyruvate dehydrogenase. In contrast, the energy requirements of astrocytes are predominantly met by aerobic glycolysis, so some microdomains in astrocytes contain relatively few mitochondria²³.

Neuronal activation transiently triggers aerobic glycolysis in astrocytes, thereby generating lactate. The astrocyte–neuron lactate shuttle **[G]** hypothesis proposes that neuronal release of glutamate during neural transmission stimulates glucose uptake, glycogen catabolism, aerobic glycolysis and lactate production in neighboring astrocytes^{8,24}. The lactate produced by astrocytes is posited to support neuroplasticity, although its precise contribution and the conditions of lactate exploitation remain unclear^{8,10,15,23} (Box 2).

Oligodendrocytes obtain ATP primarily by aerobic glycolysis. They use lactate for their own energy needs and also supply neighbouring axons with lactate, a process modulated locally by

glutamate release from neurons (Figure 1). This metabolic support of neuronal function by oligodendrocytes is important for effective spatial and temporal information processing in neuronal networks²⁵. Oligodendrocytes are responsible for myelination of axons, which speeds up action potential conduction. However, the insulating myelin sheath restricts access of axons to glucose and other metabolites that would otherwise diffuse across the extracellular space²⁵. The intermittent pattern of axonal myelination in cortical grey matter helps maintain access to extracellular nutrients²⁶. Therefore, the supply of glucose and lactate to myelinated axons by oligodendrocytes requires a highly specialized architecture of the myelin sheath, including a continuum of nanometer-wide cytosolic channels that flank compacted, mature myelin²⁵ (Figure 1). These channels connect the oligodendrocyte soma with the periaxonal space. Interestingly, neurons also deliver *N*-acetyl-aspartate to the oligodendrocytes soma via these channels; this *N*-acetyl-aspartate (as part of the aspartate–oxaloaspartate–malate shuttle) stimulates the TCA cycle and mitochondrial ATP production and is used to generate lipids and myelin²⁷.

Axons also transport mitochondria, RNA, proteins, vesicles and other cargo to presynaptic terminals. This transport is an ATP-dependent process regulated by calcium and involves motor proteins and microtubules. Retrograde transport of vesicles to the cell body for lysosomal degradation is ATP-driven². Fast-conducting axons release trace amounts of glutamate, which stimulates local *N*-methyl-D-aspartate (NMDA) receptors in oligodendrocytes, promoting surface expression of GLUT1 on axonal myelin sheaths and thereby increasing glucose uptake and the rate of aerobic glycolysis. This in turn increases local provision of lactate to axons for ATP generation (Figure 1)^{21,28}. In addition, the molecular motors driving fast axonal transport [G] are equipped with glycolytic enzymes allowing them to generate their own energy "onboard"²⁹ (Figure 1).

Unlike astrocytes and oligodendrocytes, microglia do not directly provide energy to neurons, but the high amounts of lactate released by activated microglia may well be retrieved by local neurons²³. Microglia are predominantly fueled by oxidative phosphorylation, but are metabolically reprogrammed by neuroinflammation in NDAs to an aerobic-glycolysis-predominant phenotype associated with upregulation of GLUT1 and GLUT4^{1,2}. In parallel with this energetic shift, microglia transition from a protective to a disease-driving role in NDAs. When brain glucose supply is chronically suboptimal, the high energy demands of activated microglia further limit energy availability to neurons^{1,30}.

[H2] Neuronal networks and energy use

The provision of energy substrates from astrocytes to synapses, and from oligodendrocytes to axons is critical for communication both within and between brain networks²⁵. Brain regions are connected by tracts of myelinated axons, which are adversely affected by NDAs. For example, corticocortical loops are disrupted in AD and FTD, cortico-striatal pathways in HD, corticospinal tracts in ALS and nigrostriatal projections in PD. These tracts consist mainly of long-range excitatory neurons. Inhibitory interneurons, such as fast-spiking interneurons, are mostly present within local networks, such as the CA3 region of the hippocampus and frontal cortex. Local interneuron dysfunction disrupts synchronization between remote neuronal networks and across brain regions^{31,32}.

Gamma-oscillations (30–100 Hz) are fast brain rhythms synchronizing the activity of excitatory principal neurons and neuronal networks³¹. Fast-spiking GABAergic interneurons generating gamma oscillations have a high density of mitochondria in their axons and a specialised kind of myelination that facilitates provision of energy by oligodendrocytes³³. The high metabolic needs of fast-spiking interneurons are supported primarily by oxidative phosphorylation. Parvalbumin-positive, GABAergic interneurons are particularly sensitive to deficits in energy and oxygen supply^{13,34}. The decreased ability of oligodendrocytes to provide lactate to axons probably aggravates the decline in fast-spiking interneuron activity observed in NDAs.

Rhythmically firing, highly branched, nigrostriatal dopaminergic neurons in the substantia nigra pars compacta are especially vulnerable to mitochondrial failure and oxidative stress, features characteristic of PD (Supplementary Table 1)^{13,35}.

[H2] Neuroendocrine mechanisms

As demonstrated in diverse cellular and animal models, insulin has a globally positive influence on cerebral energy balance and function. It reinforces neuronal energy supply by increasing neuronal glucose uptake by GLUT4 in the hippocampus and cortex (Supplementary Table 2)^{17,22}. Insulin and activation of insulin-like growth factor (IGF)-1 receptors also promote synaptic plasticity and cognitive processes³⁶. Nevertheless, normal insulin sensitivity is paramount; insulin resistance is a major risk factor for AD because it disrupts both the modulation of energy availability by insulin and insulin signalling pathways in the brain^{37,38}.

Other hormones, including ghrelin, incretins [G], leptin, amylin and adiponectin, modulate both appetite and energy homoeostasis and influence numerous aspects of brain function that are compromised in NDAs. The neurobiology of these hormones and their synthetic agonists in

relation to food intake and energy homeostasis, brain energy balance, mitochondrial function, cognition, motor function, neurogenesis, synaptic integrity and neuronal integrity in animal models of NDAs are documented in Supplementary Table 2, with their clinical effects reported in Tables 1-3 and in the section below on therapies based on brain energy rescue.

[H2] Brain use of ketones and lactate

Glucose (and glycogen) reserves within the brain can supply its ATP needs for only a few minutes^{24,39}. Much of the brain's resilience in the face of energetic challenge therefore depends on opportunistic use of alternative fuels sourced from outside the brain. Ketone bodies (ketones) and lactate are the main alternative fuels to glucose and are delivered to the brain by monocarboxylic acid transporters [G] on astrocytes and on the capillary endothelium (Figure 1). The two principal ketones (acetoacetate and D- β -hydroxybutyrate (BHB)), are the main alternative brain fuels to glucose in adults under conditions of dietary carbohydrate or energy restriction. In infants, however, ketones are not only an *essential* brain fuel but also the main substrate for brain lipid synthesis⁴⁰.

Acetoacetate and BHB are both in equilibrium in the blood, but only acetoacetate is metabolized to acetyl coenzyme A (acetyl CoA) which then enters the TCA cycle to generate ATP. After an overnight fast, plasma ketones are usually 0.1–0.2 mM and they supply 3–5% of brain energy requirements. However, during a 3-4 day fast, plasma ketone concentrations can reach 5–6 mM and provide \geq 50% of brain energy requirements³. Unlike glucose, which enters the brain in response to brain cell activity, the rate of ketone entry to the brain is directly related to their plasma concentration³, which explains the glucose-sparing effect of increased ketone levels⁴¹. Unlike glucose, ketones do not undergo aerobic glycolysis and cannot be metabolized to lactate, so only contribute to ATP production via oxidative phosphorylation.

[H2] The gut-brain axis

The gut microbiome is involved in bidirectional communication between the gastrointestinal tract and the brain. The intestinal microbiome generates nutrients and modulates overall energy homeostasis⁴². Disruption of the gut microbiome (dysbiosis) is implicated in the pathogenesis of NDAs⁴³. Dietary fibre is an important substrate for generation of short-chain fatty acids [G] by gut microbiota; dietary fibre also slows down glucose absorption, which improves insulin sensitivity. These effects of dietary fibre are partly mediated by short-chain fatty acids, which are ligands for

G-protein-coupled hydroxycarboxylic acid receptors (such as HCAR2) in enterocytes^{42,44}. Shortchain fatty acids produced by gut bacteria, particularly butyrate, are key fuels for intestinal cells. Propionate, butyrate and succinate (a precursor of propionate) generated by microbiota also improve control of peripheral glucose metabolism, adiposity and body weight⁴⁴. In preclinical studies, exogenous butyrate promoted the development of dendritic spines, long-term potentiation, myelination and memory formation⁴⁵. However, butyrate levels produced endogenously are usually low, so butyrate would at best be expected to be a minor energy substrate for the brain.

The beneficial effect of intestinal gluconeogenesis on insulin sensitivity and systemic energy metabolism is mediated in part by glucose-sensing in the portal vein. This information is relayed via portal sensory nerves to the brain, which suppresses appetite and reduces hepatic gluconeogenesis^{44,46}. Short-chain fatty acids also modulate the immune system, stimulate the release of hormones such as glucagon-like peptide-1 (GLP1) from the gut (Supplementary Table 2) and inhibit histone deacetylases.

[H1] Impaired brain energetics

Impaired brain glucose metabolism compromises transmembrane ion transport, vesicle recycling and synaptic signaling^{17,31,34}. Less-effective maintenance of transmembrane ion gradients and transmitter release, especially in fast-spiking interneurons, leads to hyperexcitability, excitatory/inhibitory imbalance and functional impairment of cortical networks, which further compromises the brain's energy efficiency. These changes are exacerbated by disrupted glutamatergic transmission and abnormal astrocyte and oligodendrocyte function^{13,34,47,48}, as well as impaired autophagy which, in turn, decreases nutrient recycling². Furthermore, the neuroinflammation common to diverse classes of NDA is energetically costly^{-2,49}.

The regional pattern of brain energetic disruption depends on the NDA and its pathophysiological phenotype (Supplementary Table 1). Indeed, brain glucose hypometabolism in NDAs has no single cause; reduction in neuronal glucose uptake, impairment in aerobic glycolysis and the TCA cycle, failure of axonal transport, and the loss of glial energetic support to neurons are all implicated. Supplementary Box 1 outlines how induced pluripotent stem cells (iPSCs) and organoids are illuminating the cellular substrates of energy dysregulation, and Supplementary Box 2 summarizes how disruption of the cerebral microvasculature exacerbates the disruption of brain energy supply in NDAs.

[H2] *Alzheimer disease*. AD is the most common NDA. It is associated with weight loss and poor appetite but also type 2 diabetes (T2D) all of which contribute to lower brain energy availability. AD is characterised by lower uptake of glucose, TCA activity, mitochondrial function, astrocyte and oligodendrocyte energetic support of neurons. In addition, microglial consumption of glucose due to neuroinflammation is elevated, siphoning energy away from neurons (see Supplementary Table 1)^{1,4,50-52}.

Even before diagnosis of AD, a characteristic regional disruption of glucose metabolism is linked to neuropathology and reduced cerebral blood flow in the brain. Nevertheless, the brain in AD still has normal or near-normal oxygen, lactate and ketone metabolism^{3,52-54}. Many positron emission tomography (PET) studies confirm that the entorhinal cortex and parietal lobes including the precuneus have a 10–12% deficit in glucose uptake in mild cognitive impairment [G] (MCI), a deficit that becomes anatomically more widespread with the onset of AD and worsens during its progression (Box 4; Supplementary Table 1). The regional pattern of the brain glucose deficit distinguishes AD from FTD, PD, Lewy body disease and other disorders associated with dementia^{3,4,6}.

White matter atrophy in AD impairs neuronal network operation and axonal mitochondrial transport. Especially in women, white matter loss reflects reduced maintenance and synthesis of myelin (energy-intensive processes) and catabolism of myelin to provide energy in the face of glucose scarcity^{37,55,56}. However, as with impaired glucose uptake in grey matter, white matter ketone uptake remains normal in AD⁵⁷.

[H2] *Parkinson disease*. In idiopathic PD, weight loss and low body mass index are common despite increased visceral fat. A decline in glucose metabolism is seen in the striatum (caudate), the frontal cortex and several other cortical regions but not in the cerebellum: this pattern of hypometabolism correlates with specific patterns of motor and cognitive dysfunction and is predictive of disease progression^{4,58,59}. Although PET imaging cannot resolve the substantia nigra pars compacta where dopaminergic cell bodies degenerate in PD, mitochondrial fragmentation and dysfunction including decreased glycolysis and reduced complex 1 activity, is pronounced in this brain region^{50,60}. Energetic deficits have been reproduced in PD-derived iPSCs over-expressing the gene encoding α -synuclein⁶¹. There is evidence that neuroinflammation further compromises neuronal fuel supply in PD¹.

[H2] *Huntington disease*. HD patients characteristically lose weight, even when increasing their calorie intake. In pre-symptomatic HD, brain glucose hypometabolism is seen the striatum, frontal and temporal cortex, and is linked to impaired neurotransmission in cortico-striatal tracts⁶². Glucose uptake, ATP generation by aerobic glycolysis, mitochondrial function and oxidative phosphorylation are all decreased in HD^{35,63,64}. Astrocytes in the striatum may oxidize fatty acids as an alternative source of energy but reactive oxygen species (ROS) contribute to further tissue damage⁶⁵. The cerebellum is less severely affected by impaired glucose metabolism than the basal ganglia, perhaps because it uses amino acids for gluconeogenesis⁶⁵. HD neurons show disrupted glycolysis⁶⁶ and impaired axonal transport of vesicles owing to the interference of mutant huntingtin protein with molecular motors^{35 67}.

[H2] *ALS and FTD.* FTD and ALS have overlapping genetic risk factors and clinical and pathophysiological features^{6,68-70}. Both are characterized by increased energy expenditure, yet only in FTD is there a distinctive carbohydrate/sweetness preference and weight gain. In contrast, ALS patients eventually lose weight due to insufficient nutrient and energy intake (Supplementary Table 1)^{68,71}. Brain energetics also deteriorate differently in ALS and FTD. FTD is associated with declining glucose metabolism and cerebral blood flow, especially in the frontal lobes, striatum and thalamus where mitochondrial function is disrupted with reduced signalling to the endoplasmic reticulum and aberrant mitophagy^{6,68,69,72}. Conversely, ALS is associated with a regionally complex pattern of lower and higher brain glucose metabolism: of particular note are reductions in mitochondrial function and glycolysis in the cortex, spinal cord and motor neurons, and at neuromuscular junctions in muscle^{68,70,73}. In the Superoxidase Dismutase 1 mouse (SOD1) model of ALS, the pentose phosphate pathway is also impaired⁷¹. Further, a loss of mitochondrial energetics and impaired glycolysis in astrocytes is linked to disruption of *C9orf72*, a genetic risk factor for ALS associated with failure of energetic support of neurons by astrocytes and oligodendrocytes^{74,75}.

[H2] Energy deficits and neurotoxic proteins

Brain glucose hypometabolism contributes to synapse loss and neuronal death in AD, with energetic deficits and neurotoxic protein accumulation mutually aggravating one another in a vicious cycle (Figure 2A)^{3,52,76,77}. Insufficient neuronal glucose and mitochondrial energy

generation compromise the clearance of amyloid-β (Aβ) 42 and tau proteins from the brain. Conversely, accumulation of Aβ42 and tau trigger mitochondrial damage, impair energy production and increase oxidative stress^{77,78}. These neurotoxic proteins also inhibit GLUT4⁵¹ and phosphofructokinase, thereby blocking glucose uptake, aerobic glycolysis and ATP synthesis⁷⁹. Mitochondria accumulate in axonal swellings and are no longer replaced in presynaptic terminals⁸⁰. Failure to clear dysfunctional mitochondria by mitophagy further compromises the bioenergetics of vulnerable neuronal circuits in AD, PD and other NDAs². Excitation/inhibition balance is crucial for network operation at optimal energetic efficiency⁴⁸ and, at the circuit level, an early, neurotoxic protein-driven feature of AD is the energetically expensive hyperexcitability of glutamatergic neurons^{34,81}, which is associated with an imbalance between excitation and inhibition in local cortical and hippocampal networks^{32,47}.

Interestingly, Aβ is involved in a healthy neuronal response to damage and/or infection⁸² but this protective function is lost when Aβ aggregates into plaques. As mentioned above, Aβ exacerbates brain glucose hypometabolism, both in foci of Aβ accumulation and in remote regions, possibly due to a pericyte-mediated constriction of capillary blood flow. In turn, this hypometabolism triggers cellular damage and neuroinflammation^{77,83}. Perturbed astrocytic and oligodendrocyte function, together with accumulation of phosphorylated tau (pTau), exacerbates aging³² and Aβ/pTau-induced network hyperexcitability, thereby perpetuating a cycle of neurodegeneration and declining brain glucose metabolism (Figure 2A)^{13,47}. This vicious cycle driven by energy-failure in AD has similarities with the neural circuit disruption seen in schizophrenia⁸⁴ and in epilepsy⁴ and contributes not only to deterioration of memory and cognition but also to abnormal behaviour in affected patients.

[H2] Energetics and endocrine dysregulation

Insulin resistance is a common feature of AD, PD, FTD, ALS and probably also HD with reduced signalling at central insulin and IGF-1 receptors contributing to deficits in neural function, synaptic plasticity and cellular integrity (Table 1, Supplementary Table 1)^{36,37,85}. Even though insulin itself does not globally promote brain glucose uptake, insulin resistance reduces glucose uptake by cortico-hippocampal neurons expressing insulin-sensitive GLUT4^{17,22}.

NDAs are associated with numerous changes in hormones that modulate brain energetics and neuroplasticity (Supplementary Table 2). The following observations may be highlighted: *First*, plasma leptin and hippocampal leptin signaling are reduced in AD, resulting in a state of leptin

resistance mirroring insulin resistance⁸⁶. This decline in leptin is superimposed on a background of declining plasma leptin with ageing and is linked to impaired learning, memory and long-term potentiation⁸⁷. Second, an age-related reduction in ghrelin signalling in the temporal cortex may be related to neuronal damage and cognitive deficits in AD⁸⁸. Circulating ghrelin is reduced in PD and the loss of its neuroprotective properties is linked to dopaminergic neuron degeneration and motor dysfunction^{88,89}. In addition to blunted neuroprotective properties, antineuroinflammatory effects of ghrelin involving astrocytes and microglia may be diminished in AD and PD^{88,90}. Third, amylin oligomers and aggregates are suspected to damage neurons and the microvasculature in AD, though amylin has a Janus-faced role as further discussed below⁹¹⁻⁹³. Fourth, an increase in circulating adiponectin has been reported in AD and ALS: if centrally expressed, this increase might counter cognitive deficits and exert neuroprotective properties, but this awaits confirmation^{86,94} ^{95,96}. In contrast to the above-mentioned hormones, there are very few data on the relationship between GLP1 and glucose-dependent insulinotropic polypeptide (GIP) and NDAs (Supplementary Table 1)⁹⁷. Brain hormone levels are challenging to measure, and cause-effect relationships hard to disentangle, but changes in the secretion and central actions of these hormones are implicated in the energy imbalance, pathophysiology and functional deficits in NDAs (Supplementary Table 2).

[H2] Energetics and disease risk factors

[H3] *Age*. Ageing is the main risk factor for NDAs, but there is an important distinction between the cognitive, structural and neurometabolic changes associated with healthy ageing versus those occurring in NDAs. During healthy ageing, some cognitive domains such as episodic and working memory and processing speed show a modest decline, whereas others (such as semantic memory) change relatively little⁹⁸. Although the decline in brain volume and cortical thickness forms a continuum between cognitively healthy ageing, MCI and AD, regional changes in brain glucose metabolism seen during healthy ageing, brain glucose metabolism mainly decreases in the frontal cortex, whereas in MCI and AD, the parietal lobe and precuneus are the most markedly affected [Box 4]. Decreased aerobic glycolysis¹⁰¹, loss of myelination, network perturbation and attenuation of neurovascular coupling are integral features of the ageing brain that might provide a template for the onset of the more severe brain energetic deficits in NDAs^{32,56,102,103}. Mitochondrial proteins are expressed at lower levels in brains of older people experiencing

accelerated cognitive decline¹⁰⁴.

[H3] *Metabolic dysregulation*. The risk of NDAs is substantially higher in conditions of metabolic dysregulation, including insulin resistance, obesity and T2D (Table 3)¹⁰⁵. Most strikingly, poorly controlled type 1 diabetes (T1D) or T2D is associated with increased risk of cognitive impairment and AD¹⁰⁶. Intriguingly, similarities exist between AD and T2D with respect to the deleterious effects of A β in the AD brain and the disruptive actions of amylin in the pancreas and brain in T2D: both disorders are associated with neurotoxic protein-induced peripheral metabolic and vascular abnormalities¹⁰⁷. In young women with polycystic ovary syndrome, mild insulin resistance is associated with a pattern of glucose-specific brain hypometabolism similar to that seen in elderly people¹⁰⁸, suggesting that the adverse effect of insulin resistance on brain energy metabolism is independent of age.

T2D doubles the risk of developing PD, possibly owing to increased expression of 2-synuclein⁸⁵. Interestingly, the risk of ALS is increased in T1D, but obesity and T2D are associated with decreased risk of ALS^{2,74}. The metabolic syndrome associated with insulin resistance and weight gain is also present in "atypical" major depression, itself often co-morbid with NDAs, especially AD and PD¹⁰⁹. Effective treatment of T2D, metabolic syndrome and depression would be expected to reduce the risk of developing AD and other NDAs¹¹⁰.

Despite the chronic deficit in brain glucose uptake and utilization in NDAs, the normal ketogenic response to low plasma glucose levels is not stimulated because the main drivers of endogenous ketone production – low blood glucose and low insulin – are absent. Chronic mild hyperglycemia and mild insulin resistance commonly develop as people age, so plasma insulin rarely drop for long enough to release the insulin-mediated inhibition of lipolysis in adipose tissue, the source of the endogenous free fatty acids needed for ketogenesis. This metabolic deterioration continues as AD develops, so the brain experiences a chronic, progressive glucose-specific brain energy gap [G]³ (Figure 2A) that is not corrected by ketone production as it would be if insulin sensitivity was normal and plasma glucose was decreased by a period of carbohydrate or caloric restriction [G].

[H3] *Oestrogen*. Menopause is associated with deteriorating systemic and brain glucose metabolism, weight gain, insulin resistance and loss of mitochondrial efficiency¹¹¹. Ovariectomized rodent models of menopause show metabolic responses similar to fasting,

including increased oxidation of long-chain fatty acids and elevated plasma ketones, as well as white matter and myelin degeneration, changes that in part reflect the use of brain lipids as a source of fatty acids for ATP generation^{56,112}. In fact, oestrogen modulates many facets of brain glucose metabolism, including uptake, aerobic glycolysis and oxidative phosphorylation³⁷. Oestrogen also stimulates the catabolism and clearance of A^[2], in part by upregulating insulindegrading enzyme, so the loss of oestrogen after menopause could directly favour pathological processes leading to AD^{56,112}. Accordingly, declining plasma oestrogen is associated with an increased incidence of AD in women, although this relationship remains controversial^{37,112}.

[H3] *Genetic risk factors*. Possession of two *APOE-* ε 4 alleles (encoding the ApoE4 polymorphism of apolipoprotein E) confers the highest genetic risk of sporadic AD. In *APOE-* ε 4 carriers the brain is hyperexcitable¹¹³, has reduced glucose utilization in regions affected by glucose hypometabolism in AD¹¹⁴, and accumulates more aggregated A β . Regardless of age, the following parameters all decline in *APOE-* ε 4 carriers in response to a high fat diet - brain insulin signaling¹¹⁵, expression of glucose-regulating enzymes and glucose transporters¹¹⁴, mitochondrial function in the cortex, and cognitive function ^{104,116}. These effects of ApoE4 on brain energetics are additive to the adverse effects of A β ⁷⁷.

Some of the adverse effects of ApoE4 may result from production of a C-terminal fragment of the ApoE4 protein, which inhibits the electron transport chain [G]^{117,118}, increases generation of ROS and forces neurons to increase their reliance on aerobic glycolysis or alternative energy substrates¹¹⁸. Whether or not ApoE4 affects ketone metabolism in individuals with MCI or AD is controversial. In one AD study, a ketogenic supplement did not raise plasma ketones or improve cognitive outcomes as much in *APOE-* ε 4 carriers as it did in non-carriers¹¹⁹. A clinical trial of medium-chain triglycerides [G] in AD who were specifically selected noncarriers of *APOE-* ε 4 showed beneficial cognitive outcomes after 1 month¹²⁰. However, in transgenic mice expressing human *APOE-* ε 4, the presence of ApoE4 did not significantly affect brain ketone uptake versus wild-type controls¹¹⁴.

Polymorphisms in major risk genes for PD, including *PINK1* (encoding PTEN-induced putative kinase protein 1) and *PRKN* (encoding E3 ubiquitin-protein ligase parkin) are closely linked to impaired brain ATP production^{50,121}. Phosphorylation of the endocytic sorting protein Rab10 by leucine-rich repeat serine/threonine-protein kinase 2 (LRRK2) is essential for GLUT4 translocation to the neuronal plasma membrane and is defective in PD patients possessing the *LRRK2* G2019S

mutation¹²². In HD, axonal transport of mitochondria and glycolytic proteins to the synapse is hindered by mutant huntingtin protein²⁹. In ALS and FTD, the proteins encoded by risk genes such as *TARDBP* (encoding TAR DNA-binding protein [TDP]-43) interfere with mitochondrial function and quality control, thereby compromising ATP production¹²³. Furthermore, the most prominent risk gene for ALS and FTD, *C9orf72*, encodes part of a complex with guanine nucleotide exchange factor activity that is linked to decreased autophagic lysosome-driven nutrient recycling, leading to frontal and thalamic glucose hypometabolism and aberrant lipogenesis^{2,124}. Indeed, many products of risk genes associated with NDAs interfere with autophagic lysosomal clearance, which has a doubly disabling effect because the metabolic end-products of carbohydrates, fats and proteins are then lost to energy generation pathways².

[H1] Therapies based on brain energy rescue

As outlined above, the prevailing notion that impaired brain glucose metabolism in NDAs is simply a *consequence* of neuronal dysfunction is being re-evaluated. Most notably in AD, the progressive decline in brain glucose uptake and metabolism creates a chronic brain energy gap that contributes to brain cell dysfunction and accumulation of neurotoxic proteins even before the onset of cognitive and neuropsychiatric deficits³ (Figure 2B). Once glycolysis is impaired and neuronal function starts to decline, the brain energy deficit cannot be corrected by simply increasing blood glucose; indeed, additional dietary glucose aggravates the insulin resistance already commonly present in older people¹⁰. Furthermore, brain glucose uptake is driven by neuronal activity not by circulating glucose levels³. Conversely, ketones and lactate are an alternative brain energy source⁴¹, brain uptake of which is driven by their availability in the circulation.

Because no single common pathway causes brain energy deficits in NDAs, brain energy rescue strategies may need to target different metabolic pathways and processes depending on the disease in question^{3,4,125} (Figure 3B). Some of these strategies focus on a single enzyme, transporter or metabolite, but others are broader (Tables 1-3). The following discussion first addresses the energetic dimension of mitochondrial dysfunction in NDAs. Strategies that have broader effects such as modulating redox status and ketone-based approaches are described next, then hormone-based approaches to brain energy rescue, followed by a suite of novel strategies currently under exploration. These strategies should all act synergistically with preventive lifestyle changes, such as increased exercise and dietary improvements that help

counter insulin resistance^{126,127} (Table 3, Box 3). For links between mitochondrial dysfunction, oxidative stress and neurodegeneration, see two recent reviews^{60,78}.

[H2] Support of mitochondrial function

Despite continued uncertainty about the extent to which mitochondrial damage is a consequence versus cause of the onset or progression of NDAs⁶⁵, considerable research focuses on improving mitochondrial function by protecting the electron transport chain, promoting mitochondrial biogenesis **[G]** and/or reducing oxidative damage to mitochondria¹²⁵. Assessment of mitochondrial integrity is mostly indirect, but histochemical evidence of decreased cytochrome C activity in post-mortem brain samples from young adult *APOE*- $\varepsilon 4$ carriers¹¹⁸ demonstrates that impaired mitochondrial function can be present in pre-symptomatic individuals at risk of AD.

CP2, a proprietary tricyclic pyrone, improves cognitive and behavioural phenotypes in transgenic AD mice, in part by binding to and partially inhibiting the flavin mononucleotide subunit of complex 1. This improves mitochondrial bioenergetics and overall brain energy status, possibly because of increased mitochondrial biogenesis¹²⁸. CP2 also stimulates AMPK, promotes neuronal resistance to oxidative stress, reduces brain levels of pTau and Aβ, improves axonal trafficking, and increases brain-derived neurotrophic factor (BDNF) and synaptic proteins in vivo^{128,129}. Controlling the activity of complex 1 specifically seems to underpin this beneficial effect¹³⁰ because mutations that inhibit both complexes 1 and 3 or both complexes 1 and 5 are detrimental to brain energetics¹³¹.

The mitochondria-targeted antioxidant, Mito-Q, reduces oxidative stress in mitochondria and is neuroprotective in several NDA models (Table 1). Resveratrol stimulates mitochondrial biogenesis through the sirtuin 1 (SIRT1)/AMPK/peroxisome proliferator-activated receptor (PPAR)γ coactivator 1α (PGC1α) pathway. Resveratrol also recruits AMPK to enhance autophagy, which removes damaged organelles (including mitochondria) and misfolded proteins and recycling their components, thereby promoting ATP generation^{2,132}. Replacement of old and/or damaged mitochondria starts in the neuronal cell body with new mitochondria being transported along axons to presynaptic terminals¹⁵. Both ageing and NDAs increase mitochondrial division in a manner decoupled from the normal fission–fusion cycle, suggesting that mitochondrial fragmentation could be beneficial in NDAs¹³³. Quinazolinone or its derivatives such as mitochondrial division inhibitor 1 (Mdivi-1) were originally described as selective inhibitors of mitochondrial fission, but their neuroprotective effects in both *in vitro* and *in vivo* models of AD,

PD and traumatic brain injury are now thought to reflect improved mitochondrial fusion and biogenesis¹³⁴⁻¹³⁶, and possibly better function of complex 1.

A pilot clinical study showed that S-equol, a selective oestrogen receptor-β agonist, improves cytochrome C oxidase activity in AD¹³⁷, so treatments that improve mitochondrial function by selective partial inhibition of complex 1, target mitochondrial uncoupling proteins or increase mitochondrial biogenesis may result in clinical improvement in NDAs (Table 2). Mitochondrial uncoupling proteins could help cells to resist oxidative and metabolic stress⁹⁸. Low doses of the uncoupling agent, dinitrophenol, had a neuroprotective effect in preclinical models of AD, PD and HD¹³⁸. In a mouse model of HD, mitochondrial respiration was improved by bexarotene, a retinoid X receptor agonist and PPAR^D activator¹³⁹.

[H2] Redox state, glycolysis and the TCA cycle

The redox state **[G]** of a cell is typically measured by the ratio of oxidized to reduced nicotinamide adenine dinucleotide (the NAD⁺:NADH ratio), which is a non-invasive marker of global brain energy status^{5,78,140}. In general, nutrients and metabolites that raise either blood NAD⁺ levels or the blood NAD⁺:NADH ratio improve the energetic status of the brain¹⁴¹. The NAD⁺ precursor, nicotinamide riboside, mitigates cognitive impairment, synaptic degeneration and neuronal death in transgenic mouse models of AD^{78,98,142}. Nicotinamide riboside also improves mitochondrial function in PD neurons and reduces age-related loss of dopaminergic neurons and associated motor deficits in an animal model of PD¹⁴³. Another potential approach to raising the NAD⁺:NADH ratio is dietary supplementation of oxaloacetate¹⁴⁴. In several *in vitro* and animal models of PD, terazocin (a drug approved for benign prostatic hypertrophy) stimulated phosphoglycerate kinase-1 activity, aerobic glycolysis and ATP production¹⁴⁵. Patients taking terazocin to treat other conditions had a decreased risk of developing PD and slower PD progression, so its repurposing to treat PD seems promising.

Supplementation with pyruvate could potentially improve brain energetics by stimulating pyruvate dehydrogenase^{146,147}, a possibility supported by the rescue of defective aerobic glycolysis by pyruvate in HD-derived human iPSCs⁶⁶. Treatments that improve mitochondrial function have had mixed success in preclinical models of ALS and these approaches remain largely untested in humans^{70,74}.

Interventions that raise circulating ketones also increase acetyl CoA, which fuels the TCA cycle independently of aerobic glycolysis. Preclinical studies show that supplementation with BHB,

caprylic acid (an 8-carbon saturated fatty acid), oxaloacetic acid, capric acid (a 10-carbon saturated fatty acid), as well as a ketogenic diet [G] or caloric restriction (Box 3), all contribute to increased TCA cycle activity within the brain^{144,148} (Figure 3B). In humans, plasma medium-chain fatty acids can be transported into and metabolized by the brain¹⁴⁹. TCA cycle intermediates also give rise to bioactive molecules such as the neurotransmitter, acetylcholine, that are decreased in AD. These responses are generally reversible and therefore transformation of glutamate into α -ketoglutarate (which enters the TCA) generates ATP in neurons and glia⁹⁸.

Triheptanoin, a triglyceride of heptanoic acid, delays motor symptoms and is neuroprotective in animal models of ALS⁷⁴, epilepsy and ischemic stroke^{150,151}. Triheptanoin also reduces the effort needed to undertake exercise in HD, a beneficial effect associated with increased creatine phosphate in the brain¹⁵². Triheptanoin appears to substitute for the branched-chain amino acids that are an endogenous substrate of anaplerosis [G] and are decreased in HD^{152,153}.

[H2] Ketone-based strategies

Several clinical trials show that ketogenic interventions result in cognitive and/or functional improvements in MCl¹⁵⁴⁻¹⁵⁶, AD^{119,120,157-159} and PD^{160,161}. These interventions fall into two categories: ketogenic dietary supplements containing medium-chain triglycerides (either caprylic acid (C8) alone or caprylic acid plus capric acid (C8C10)); and the very low carbohydrate ketogenic diet (Table 2; Box 4). In the phase 1¹¹⁹ and phase 2¹⁵⁴ placebo-controlled studies of C8 and C8C10 supplementation, the interventions lasted from 12 weeks¹¹⁹ to 6 months¹⁵⁴, respectively. Two subsequent feasibility studies of ketogenic diets in AD showed improved global cognitive scores in the most compliant patients, but did not have control groups^{157,162}.

A recent 6 month study of C8C10 supplementation in MCI showed a direct and statistically significant dose–response relationship between brain uptake of ketones and/or plasma ketone and executive function, verbal fluency and language, strongly implying that ketones were directly and mechanistically linked to cognitive improvement via brain energy rescue¹⁵⁴. Because of the short half-life of ketones in the body, however, the main challenge with ketogenic interventions is to achieve a sustained therapeutic level of ketosis. In two studies of ketogenic supplements that had a sample size large enough to provide adequate statistical power to assess cognitive outcomes, the 24-h average plasma ketone level was ≤ 0.2 mM for C8¹¹⁹ and ≤ 0.4 mM for C8C10¹⁵⁴; these ketone levels would only have partially corrected the brain energy gap in MCI and less so in AD (Figure 2B, Box 4). In other clinical trials with a ketogenic diet in AD^{157,162}, MCI^{155,156} or PD^{160,161},

higher plasma ketone levels were directly related to improved clinical outcomes, but sample size and patient adherence were inadequate to produce definitive evidence of a cognitive benefit.

In PD, consuming a ketogenic diet for 8 weeks led to a substantial reduction in urinary problems, pain and fatigue scores *versus* those in a control group consuming a low-fat diet^{160,161}. The ketogenic diet group also showed a trend towards improved motor scores versus the control group. Ketogenic interventions are being explored with some success in animal models of PD¹⁶³, ALS¹⁶⁴ and HD¹⁶⁵ (Table 1), but randomized, controlled clinical trials of this approach are yet to be reported in these NDA. A ketogenic diet promotes neurovascular function and metabolic status in mice, along with a healthier intestinal microbiome profile¹⁶⁶.

Studies in which a single dose of a ketogenic supplement transiently improved cognition in AD¹⁶⁷ and in cognitively normal older people (66 years old)¹⁵⁸ suggest that ketones reduce the brain energy gap by bypassing glycolysis and providing acetyl CoA to enter the TCA cycle directly (Figure 3B). This interpretation is supported by reports that mild-to-moderate ketosis lasting <4 h prevents the autonomic, cognitive and behavioural symptoms of acute insulin-induced hypoglycemia in T1D¹⁶⁸. In turn, mild ketosis probably spares some glucose to be used by pathways other than glycolysis and oxidative phosphorylation⁴¹; including the pentose phosphate pathway which generates NADPH but also anaplerosis for the TCA cycle (Figure 3). Whether glucose-sparing is central to the therapeutic effect of ketone supplementation in NDA remains to be determined.

Metabolism of ketones in the brain not only generates fuel, but also provides an important substrate for the synthesis of brain lipids, including myelin⁴⁰. Ketones are also substrates for post-translational protein modification and activate cell signaling¹²⁵. The density and activation of HCAR2 is increased in the substantia nigra in PD¹⁶⁹ and is neuroprotective in an animal model of PD¹⁷⁰. Reducing neuronal hyperexcitability by raising GABAergic tone may contribute to the efficacy of ketone supplementation in individuals with NDAs as it does in epilepsy^{13,148,171}.

Disease modification—that is, retardation or reversal of neuropathology — is a crucial goal in the treatment of NDAs. Studies in transgenic AD mice show that ketones decrease A^{\square} deposition in the brain¹⁷² and reduce the excitatory effect of A β 42 on neurons¹⁴⁶. These preclinical studies have been confirmed in a pilot clinical study in MCl¹⁵⁶, suggesting that in addition to providing an alternative brain energy substrate that bypasses the brain glucose deficit, ketogenic interventions could potentially improve cognitive outcomes in MCI and AD by slowing pathological processes that result in A β accumulation and damage.

High-fat diets are commonly perceived to increase the risk of cardiovascular disease, so it is important to consider their potential risks. Very-high-fat ketogenic diets have been assessed in five clinical trials of durations ranging from 6 to 12 weeks. Three of these trials were conducted in MCI or AD^{155,157,162} and the other two in PD^{160,161}. In none of these trials were common biomarkers of cardiovascular risk increased, including body weight or plasma LDL cholesterol or triglycerides. However, no standard definition of a ketogenic diet exists and some 'high-fat' diets used experimentally (and possibly also clinically) might adversely affect cardiovascular health outcomes (in particular during very long-term utilisation) because they contain an excess of refined carbohydrate. In clinical trials, ketogenic medium-chain triglycerides were not associated with increased cardiovascular or metabolic risk (Table 2); indeed, like the very high fat ketogenic diet, medium-chain triglycerides are commonly used to treat obesity and T2D, which are themselves risk factors for NDAs.

[H2] Increasing insulin sensitivity

Brain energy homeostasis is closely linked to peripheral insulin sensitivity, both of which depend on the balance between global energy intake and use. The two main determinants of peripheral insulin sensitivity are exercise and intake of refined carbohydrate¹⁷³. When lifestyle changes are ineffective or difficult to implement, peripheral injections of insulin are commonly used to treat T2D, but the challenge is to avoid episodes of hypoglycemia and exacerbation of insulin resistance, both of which increase morbidity¹⁶⁸.

[H3] Intranasal insulin and insulin sensitizers. Intranasal insulin and intranasal insulin sensitizers could potentially mitigate the deleterious effects of insulin resistance and a glucose deficit (Figure 3, Tables 1-3). Intranasal insulin enters the brain directly via olfactory neurons, which enables treatment of CNS insulin resistance while minimizing systemic hypoglycemia. Short-term studies show that intranasal insulin enhances cognitive function in healthy young adults, MCI and mild– moderate AD, in part by stimulating brain glucose metabolism^{38,174}. Little or no intranasally administered insulin enters the peripheral circulation, but intranasal insulin delivery still needs to be optimized to achieve a more consistent increase in brain insulin before its efficacy for cognitive improvement can fully be assessed³⁸.

Metformin decreases hepatic glucose production, which improves insulin sensitivity in T2D, so it is being investigated for therapeutic use in NDAs¹⁷⁵. Metformin reduces neuropathology and

corrects memory deficit in AD mice¹⁷⁶ and normalizes cortical network disruption and anxious behaviour in HD mice¹⁷⁷. Within the brain, metformin also stimulates autophagy, improves synaptic function and reduces neuroinflammation — effects that mimic those of caloric restriction and exercise^{2,178}. Metformin suppresses coupling of the redox and proton transfer domains of complex 1, but its overall mechanism of action remains unclear^{179,180}. Recent data suggest that growth/differentiation factor 15 acting via "GDNF family receptor α -like" may mediate the influence of metformin on energy metabolism, suggesting that they could become novel therapeutic targets for safely combating brain energetic deficits associated with NDAs¹⁸¹, but it may also exacerbate AI accumulation¹⁸². Indeed, metformin is potentially protective against cognitive decline in MCI or AD^{183,184} and cognitive impairment due to stroke¹⁸⁵ yet adverse effects have been linked to over-activation of AMPK and vitamin B₁₂ deficiency¹³². Additional studies are needed to further clarify the putative utility of metformin for restoring energetic status in NDAs^{175,186}.

Impaired brain glucose uptake and insulin receptor desensitization could potentially be corrected by targeting nuclear hormone receptors that activate insulin-regulated and IGF-regulated pathways. For example, thiazolidinediones are PPARγ agonists that potentially reduce brain insulin resistance associated with AD and other NDAs³⁶. In contrast, PPARγ agonists, including pioglitazone and rosiglitazone, have produced no cognitive benefit in clinical trials in AD. Inhibitors of sodium–glucose co-transporter 2 such as dapagliflozin increase glucose excretion, improve cardiovascular outcomes and reduce mortality in T2D¹⁸⁷. In addition, these agents induce mild ketonemia¹⁸⁸, suggesting that they should be tested in NDAs, perhaps in combination with ketogenic interventions.

[H3] Incretin hormones. GLP1 receptor agonists such as liraglutide are approved to treat insulin resistance, obesity and T2D^{189,190}. Based on encouraging findings in animal models of AD, PD, HD and ALS (Table 1, Supplementary Table 2), they are also being assessed for treatment of NDAs^{97,191,192}. For example, liraglutide and exenatide reduced neuropathology, neuroinflammation and microvascular pathology and improved cognitive outcomes in a transgenic mouse model of AD (Table 2)¹⁹². A GLP1 receptor agonist reduced Aβ accumulation and reduced mitochondrial pro-apoptotic signaling, while increasing anti-apoptotic signaling and BDNF¹⁹². Semaglutide, a long-acting GLP1 analogue, was more neuroprotective than liraglutide in an animal model of PD, a beneficial effect related to improved mitochondrial function and lower

oxidative stress, apoptosis and neuroinflammation⁹⁷. Inhibitors of GLP1 breakdown are now in clinical trials to treat NDAs¹⁹³.

In mild-to-moderate AD, liraglutide for 6 months attenuated the decline in brain glucose uptake but had no effect on brain A β load or cognitive outcomes¹⁹⁴. Exenatide reduced symptoms of PD in a phase II trial¹⁹⁵. Interestingly, metformin might exert its actions partly via GLP1³⁶. Some dipeptidyl peptidase 4 (DPP4) inhibitors (gliptins) approved to treat T2D are known to prolong the activation of GLP1¹⁹⁶; one such agent, sitagliptin, improved cognition in elderly diabetic individuals with or without AD¹⁹⁷.

GIP agonists have shown similar benefits to GLP1 agonists in mouse models of PD^{198,199}. GIP agonists closely mimic GLP1 agonists in animal models of AD^{200,201}. Dual agonists of both GLP1 and GIP were more effective in rodent models of PD than GLP1 agonists alone^{200,201}. A triple agonist of GLP1, GIP and glucagon receptors had broad neuroprotective activity in a mouse model of AD²⁰². Clinical data are eagerly awaited for these multitarget agents.

[H3] Ghrelin. Ghrelin is neuroprotective and improves cognition in animal models of AD, PD and HD (Supplementary Table 2)²⁰³⁻²⁰⁵. The beneficial effects of ghrelin involve promotion of neuronal glucose uptake, increased expression of uncoupling protein 2, improved mitochondrial function, and enhanced mitophagy^{88,205}. AMPK activation in dopaminergic neurons of the substantia nigra may also contribute to the beneficial effects of ghrelin in PD: AMPK activates PGC1 α to induce mitochondrial biogenesis and increase ATP production, as well as stimulating autophagy to eliminate α -synuclein^{2,205,206}. Since ghrelin counters gastrointestinal dysfunction in PD, relamorelin, a centrally penetrant and selective agonist of the ghrelin receptor (also known as growth hormone secretagogue receptor type 1), is being assessed to treat constipation in PD and T2D. The effects of relamorelin on motor function, neuronal survival and bioenergetics are also being assessed in PD²⁰⁷.

[H3] Leptin and adiponectin. Leptin promotes mitochondrial function, has neuroprotective properties and mitigates the neurotoxic effects of A β accumulation in animal models of AD and PD^{86,87,208}. Synergistic beneficial effects on mitochondrial function have been reported for leptin in combination with PPAR α agonists²⁰⁹. The risk of ALS is reduced in people with T2D, so it is interesting that knocking out the gene encoding leptin (*Lep*), which suppresses appetite in a mouse model of ALS, slowed the progression of ALS symptoms while decreasing energy

expenditure and increasing body weight²¹⁰. This suggests that leptin antagonists should be evaluated as a potential treatment in ALS. Data on adiponectin are currently limited to mouse models of AD in which adiponectin agonists had neuroprotective properties associated with reduced loads of A β and pTau and improved cognitive performance — benefits that were related to increased glucose uptake in the hippocampus and, possibly, to improved insulin sensitivity^{86,211}.

[H3] *Amylin*. The status of amylin as a potential target for treating NDAs is controversial⁹² because native amylin itself is amyloidogenic and aggregated amylin has pro-apoptotic and neuroinflammatory effects and may seed A β aggregation^{91,93}. Moreover, A β binds to amylin receptors and its neurotoxic actions and interference with cognition were blunted by an amylin antagonist^{91,212}. Nevertheless, amylin itself could have potential beneficial properties as a leptin sensitizer, including leptin resistance in AD²¹³. The satiety-stimulating effects of amylin may help to control weight gain as well as improve insulin sensitivity and brain glucose metabolism. Treatment with human amylin or a non-aggregating amylin analogue, pramlintide, has shown both cognitive benefits and reductions in A β pathology in animal models of AD²¹⁴. Amylin and pramlintide also promote A Ξ efflux from the brain, regulate synaptic proteins, reduce oxidative stress and inflammation and improve mitochondrial function^{214,215}. Pramlintide protects against the neurotoxic and memory-disrupting actions of A β ⁹¹. Whether amylin-related mechanisms can be harnessed in the treatment of AD and other NDAs remains to be seen.

[H2] Restoration of downstream signalling

An important fate of glucose distinct from its use as an energy substrate is its utilization to generate *O*-linked β -*N*-acetylglucosamine (*O*-GlcNAc), which is post-translationally and reversibly added to serine and threonine residues of numerous proteins. *O*-GlcNAcylation occurs *via O*-GlcNAc transferase whereas *O*-GlcNAcase removes *O*-GlcNAc residues: both of these enzymes are therapeutically targetable in NDA²¹⁶. *O*-GlcNAcylation is important for axonal stability and synaptic plasticity and for the local and dynamic coupling of glucose utilization to glycolysis and mitochondrial function at both pre-synaptic and post-synaptic sites²¹⁶. In primary cell culture models of PD, *O*-GlcNAcylation of α -synuclein reduced its aggregation and toxicity²¹⁷. In addition, in transgenic mouse models of AD, downregulation of *O*-GlcNAcylation is implicated in the production of A β and pTau²¹⁸. Novel therapeutic agents that aim to restore *O*-GlcNAcylation are currently under investigation in experimental models of NDAs²¹⁶⁻²¹⁸.

[H2] Epigenetic interventions

In addition to driving the TCA cycle, acetyl-CoA is a precursor of brain lipids and a substrate for generation of acetylcholine. Acetyl-CoA is also the source of the acetyl moiety used to acetylate several enzymes that modulate glycolysis, gluconeogenesis and the TCA cycle, tau (acetylation of which promotes its aggregation) and histones²¹⁹. Acetylation is a core component of histone modification, which regulates gene expression, so cellular energetics are affected by the availability of acetyl-CoA for histone acetyl transferases^{220,221}. Accordingly, acetyl-CoA provides a direct link between brain energy balance and the epigenetic control of gene expression — a link reinforced by other components of the TCA cycle, including the intermediates, succinate and citrate²¹⁹. Furthermore, increased activity of the deacetylating enzyme, sirtuin 1, promotes mitochondrial function⁹⁸ and is implicated in the positive influence of the exercise-induced increase in brain lactate on cognition²²². Specific histone deacetylases in the hippocampus may contribute to the improved resilience to stress by lactate in mice²²³.

Short-chain fatty acids produced by the intestinal microbiome also influence the activity of histone deacetylases⁴⁴. For example, BHB modulates the β -hydroxybutyrylation of histones at lysine residues, which couples metabolic status to the control of gene expression²²⁴. Post-translational histone modifications and DNA methylation influence bioenergetic processes that are disrupted in NDA, so pharmacological modulation of these epigenetic mechanisms could potentially improve brain energy status in NDAs^{220,221}.

[H2] RNA-based and DNA-based therapies

Diverse techniques are being developed to alter the level of mRNA encoding proteins that are anomalously expressed in NDAs. These strategies could suppress neurotoxic effects or compensate for a loss of physiological function, thereby improving the energetic status of the brain. Targeting specific classes of microRNAs and long noncoding RNAs that control the translation of dysregulated glycolytic and ATP-generating mitochondrial proteins should also be feasible.

Recent clinical success with oligonucleotide-based therapies in CNS disorders such as spinal muscular atrophy^{225,226}, together with advances in the manipulation of oligonucleotides, such as antagomirs **[G]** and locked nucleic acids **[G]**, make this approach increasingly relevant to NDAs, even for hitherto 'undruggable' targets²²⁷⁻²²⁹. In HD, clinical trials of antisense oligomers directed

against mutant *huntingtin* mRNA are underway to prevent its interference with mitochondrial transport and function²³⁰. In addition, allele-specific strategies that specifically decrease mutant *huntingtin* mRNA while preserving normal *huntingtin* mRNA are under investigation. Some of these interventions use zinc finger nucleases (which act as transcription factors), whereas others rely on small molecules that promote clearance specifically of the mutant proteins²³¹. Antisense oligomers and similar approaches could also be used to target genes containing mutations that disrupt mitochondrial energetics in PD and ALS with FTD (Supplementary Table 1)^{232,233}.

Oligonucleotides and siRNAs that modulate pre-mRNA splicing or neutralize mRNA-directed miRNAs preferentially increase the expression of the intact allele of energy-generating genes down-regulated in NDAs^{226,234}. A more direct mode of gene therapy that aims to restore abnormally low or absent gene expression is to transfer copies of the intact gene into the brain using an adeno-associated virus vector. For example, the approved gene therapy onasemnogene abeparvovec employs an adeno-associated virus vector to deliver intact *SMN1* gene copies to motor neurons to treat spinal muscular atrophy²³⁵. An adeno-associated virus vector loaded with the human *GLUT1* promoter injected directly into the brains of GLUT1-deficient mice led to robust GLUT1 expression in cortico-limbic regions, together with increased CSF glucose and improved motor function²³⁶. A similar strategy that targeted dysfunctional PGC1α helped to restore mitochondrial function in dopaminergic pathways in mouse models of PD²³⁷.

DNA and RNA editing might also become options for the treatment of NDAs, for example, using zinc finger nucleases or CRISPR/Cas9 technologies. One specific approach to improve brain energetics in AD is the conversion of the ApoE4 isoform into ApoE3 as shown in a neuronal cell line^{234,238,239}. Gene editing of *APOE* has not yet been achieved *in vivo* but progress is rapid in this field and a broad range of options for improving glucose metabolism and other abnormalities associated with AD based on neutralization of ApoE4 is being investigated²⁴⁰.

Mitochondrial dysfunction and other disturbances associated with HD may also be linked to excessive translation of mRNAs and overproduction of proteins resulting from inactivation of the eukaryotic translation initiation factor 4E (EIF4E) translational repressor complex. Rapamycin and other repressors of this complex or its components should therefore be assessed to restore mitochondrial energetics and integrity in NDAs²⁴¹. autophagy-targeting chimera. Another approach would be so-called 'autophagy-targeting chimeras' to specifically target and eliminate fragmented mitochondria. In a proof-of-concept study, this approach restored the generation of

functional mitochondria and ATP levels in fibroblasts from Down patients, who invariably develop AD²⁴².

[H2] Photobiomodulation therapy

Low wavelengths of light penetrate brain tissue to a considerable depth and transcranial (intracranial, intra-aural or intranasal) application of near-infrared light is under study as a treatment for various brain disorders^{243,244}. The mechanisms underlying the positive effects of photobiomodulation therapy await further elucidation, but increased brain perfusion, energy availability and oxygen supply have been proposed, in addition to neuronal actions implicating light-absorbing cytochrome C and increased ATP production²⁴³⁻²⁴⁵. The use of photobiomodulation to improve brain energetics has received initial support from small-scale clinical trials^{243,244,246}, but rigorous controlled studies with larger sample sizes are needed. An alternative strategy to improve the energy status of the brain could be non-invasive light and/or auditory stimulation regimes. In a mouse model of AD, this approach improved pathology and reduced neurotoxic proteins, in part by favouring neurovascular coupling and, by inference, brain energetic status²⁴⁷.

[H1] Conclusions and outlook

Impaired brain energy metabolism is now recognised in NDAs and, at least in AD, clearly precedes the onset of clinical symptoms. The metabolic defects occur at multiple levels including reduced neuronal glucose uptake, impaired glycolysis and suboptimal function of the TCA cycle, all of which adversely impact axonal transport, mitochondrial function and ATP production (Figure 3A). The multiple faces of brain glucose hypometabolism present challenges for drug development in NDAs. Indeed, in view of the multiple brain energetic pathways affected, a fundamental question is whether pharmacological approaches that target a single enzyme, receptor or protein could ever be truly clinically effective. By analogy to other multifactorial disorders associated with an increased risk of NDAs, like depression and T2D²⁴⁸ where multi-modal interventions are the most effective, the same could turn out to be true for brain energetic rescue in NDAs. An example would be use of agents that simultaneously clear aggregated toxic proteins and/or suppress ROS and neuroinflammation^{2,249,250}.

The efficacy of a multi-modal approach depends on a better understanding of the cause and effect relationships between the brain glucose deficit and pathophysiological processes^{1,2,103}. In

any event, attempting to promote energetically expensive processes such as microglial clearance, synaptic remodeling, myelin regeneration or axonal transport seems questionable unless the brain has adequate energy resources to fuel the additional work. Therefore, optimization of brain energetics should become a core component of future clinical trials of potential therapies for NDAs, irrespective of their mechanisms of action, because unless the brain energy gap can be reduced (Figure 3), potential benefits of new medications may well be missed. Furthermore, a multi-pronged approach embracing both targeted pharmaceutical treatments and broad improvement in diet and lifestyle is emerging as a viable way to improve both prognosis and clinical symptoms of NDAs^{2,5,249,251}. Insights could also be garnered by considering brain energetics in other neurological diseases (Box 5).

Preclinical studies have demonstrated that brain energy rescue can delay the onset and/or progression of NDAs at two levels: first, by improving neuronal integrity, synaptic plasticity and neuronal–glial interactions linked to cognitive and functional deficits; and second, by disease modification, at least for metformin¹⁷⁶ and certain ketone-based interventions^{146,172}. The benefit of ketone-based interventions in NDAs resembles that seen in other brain disorders including schizophrenia⁸⁴ and epilepsy^{4,252}. It is interesting that ketones and the other traditional black sheep of energy metabolism —lactate — which have complementary roles both as signalling and neuroprotective molecules in brain energetics, have much to offer in developing brain energetic rescue strategies in NDAs^{3,8,253,254}. Hormone-based interventions that modulate appetite and energy expenditure should also be able to contribute to both an early preventive influence by delaying the onset of neuropathology and symptoms and a later benefit by delaying further decline in cognitive function or other functional outcomes. Genetic, epigenetic and other novel strategies are also showing promise for improving brain energetics in NDAs.

Being able to link clinical symptomatic improvement to a clinical read-out or a measurable biomarker (that is, imaging, metabolite or hormone^{76,255}) would accelerate validation of clinical effectiveness and product development. In addition, an ideal biomarker would be able to predict disease modification by an intervention. Ketone PET imaging is a biomarker of brain energy status linked to cognitive outcomes in MCI (Box 4), but it does not demonstrate whether disease modification took place. Given the probable need for a long-term multi-modal strategy including lifestyle intervention, successful compliance and retention in future clinical trials is ultimately likely to depend on the intervention being personalized; for example, exercise or insulin sensitizers only for those who are insulin resistant²⁵⁶.

In conclusion, just as normal neurocognitive development during infancy depends on adequate brain energy supply, the maintenance of cognitive performance and cerebral function during ageing is contingent on the brain continuing to successfully meet its energy needs. Guaranteeing the energy status of the brain should become a cornerstone for trials attempting to delay the onset and progression of NDAs. The observations discussed herein should help us move towards this important goal.

Table 1 | Treatments that improve brain energetics and/or function in preclinical models of

neurodegenerative disorders

Study characteristics	Primary end points: results	References
Mitochondrial function		
AD mice (3xTg) receiving MitoQ (100 μM in drinking water) for 5 months	Mitochondrial function: \forall cognitive decline, \forall oxidative stress, \forall A β accumulation, ψ astrogliosis, ψ synaptic loss, ψ caspase activation, ψ neuropathology, \uparrow mitochondrial function	257
AD mice (APP/PS1) receiving 25 mg/kg/day CP2 for 14 months	Mitochondrial function:↓ complex 1 activity, ↑ AMPK, ↑ mitochondrial bioenergetics.	128
AD mice (APP/PS1) receiving 25-250 μM AP39 in neurons in culture; 100 nM/kg body weight AP39 for 6 weeks	Brain energy status and mitochondrial function: Λ brain ATP, protected mitochondrial DNA, Ψ ROS, Ψ brain atrophy	258
AD mice (APP/PS1) receiving 10 mg/kg or 40 mg/kg mdivi-1 by gavage for 1 month	Mitochondrial dynamics: \checkmark mitochondrial fragmentation, \checkmark loss of mitochondrial membrane potential, \checkmark ROS, \checkmark and synaptic dysfunction. \uparrow ATP, \uparrow learning and memory, mitochondrial function	136
AD mice (APP/PS1) receiving 400 mg/kg/day nicotinamide riboside for 10 weeks	Mitochondrial function and proteostasis: $\bigvee A\beta$ accumulation. \uparrow cognitive function, \uparrow oxidative phosphorylation activity	259
PD rats (hA53T-α-syn) receiving 20 mg/kg mdivi-1 by i.p. injection for 8 weeks	Mitochondrial dynamics: \forall mitochondrial fragmentation, mitochondrial dysfunction, oxidative stress, neurodegeneration and α -syn aggregates; \uparrow motor function	135
HD mice (HD R6/2 and YAC128) receiving DA1 peptides (1 mg/kg/day) via osmotic pump for 2-3 months	Mitochondrial dynamics and function: \uparrow mitochondrial biogenesis and bioenergetics; \downarrow inflammation and neuropathology	260
ALS mice (SOD1 ^{G93A}) receiving MitoQ (500 μM in drinking water) for 30-40 days	Mitochondrial function: ψ nitroxidative stress, neuropathology. \uparrow mitochondrial function and life span	261
SCA1 mice (<i>Sca1</i> ^{154Q/2Q}) receiving MitoQ (500 μM in drinking water) for 16 weeks	Mitochondrial function: ψ neuropathology, oxidative stress, DNA damage, neuronal loss. \uparrow mitochondrial function	262
Seizure model (risk of AD) <i>In vivo</i> study: CD1 mice; 35 energy % C10 in regular diet. <i>In vitro</i> study: astrocytes exposed to 200 mM C8 or C10 for 10 days	Seizures: In vivo study: \checkmark seizures after C10 but not C8; No change in glycolytic enzymes; In vitro study: C8 and C10 \land basal respiration and mitochondrial leak; \land ATP synthesis, antioxidant capacity by C10 but not C8	150
Insulin sensitizers		
AD mice (APP/PS1) receiving 200 mg/kg i.p. metformin for 14 days	Cognitive performance, neuropathology: \uparrow cognitive performance (Morris water maze) \lor hippocampal neuron loss, \lor A β , \lor neuroinflammation	176
HD mice (Hdh150 knock-in) receiving 5 mg/ml metformin in drinking water for 3 weeks	Early network hyperactivation in visual cortex, behaviour: ψ hyperactive neurons, \uparrow normal network patterns, ψ green fluorescent protein-hht protein synthesis, ψ anxiolytic behaviour	177
Ketogenic molecules		
AD mice (APP/PS1) receiving 26 mg/kg/day BHB and pyruvate for 5 weeks	Brain redox status: \uparrow brain nicotinamide adenine dinucleotide phosphate (reduced); \checkmark network hyperactivity (epileptiform discharges)	146
AD mice (3xTgAD) receiving 125 g ketone ester/kg in diet for 8 months	Brain TCA cycle activity, mitochondrial function: BHB ↑ fivefold; 30-40% ↑ brain TCA cycle and glycolytic intermediates; ↑ mitochondrial redox potential; ↓ oxidized lipids/proteins in hippocampus	263
AD mice (Sirt3 ^{+/-} /AppPs1); receiving ketone ester added at 22% of dietary energy for 20 weeks	Neurodegeneration, neuronal network hyperexcitability: \uparrow cortical SIRT3 expression, \checkmark loss of GABAergic neurons, \checkmark seizures and prevented death of Sirt3+/- /AppPs1 mice	264

Mitochondrial function: \uparrow mitochondrial respiration and ATP at Complex 2; \checkmark dopamine neurodegeneration and motor deficit	265
Physical symptoms: mitochondrial function Ψ spinal cord motor neuron loss, \uparrow mitochondrial O ₂ consumption, no change in survival	266
Brain redox status, cognitive performance, Λ cortical redox status; attenuated cognitive decline	267
Brain redox status: normalized cortical NAD ⁺ /NADH; nicotinamide riboside ↑ cognitive function and restored hippocampal synaptic plasticity	142
Brain glucose uptake, cognitive performance: normalized glucose uptake in hippocampus after streptozotocin; prevented spatial/non-spatial learning and memory impairment	268
Physical performance, TCA cycle activity, brain glucose uptake, cognitive performance: \uparrow hind limb grip strength by 2.8 weeks, \uparrow time to loss of balance on rotarod, \uparrow time before weight loss, \uparrow TCA cycle	269
	 ATP at Complex 2; ↓ dopamine neurodegeneration and motor deficit Physical symptoms: mitochondrial function ↓ spinal cord motor neuron loss, ↑ mitochondrial O₂ consumption, no change in survival Brain redox status, cognitive performance, ↑ cortical redox status; attenuated cognitive decline Brain redox status: normalized cortical NAD⁺/NADH; nicotinamide riboside ↑ cognitive function and restored hippocampal synaptic plasticity Brain glucose uptake, cognitive performance: normalized glucose uptake in hippocampus after streptozotocin; prevented spatial/non-spatial learning and memory impairment Physical performance, TCA cycle activity, brain glucose uptake, cognitive performance: ↑ hind limb grip strength by 2.8 weeks, ↑ time to loss of balance on rotarod, ↑ time

Reports shown here exclude those involving the ketogenic diet and lifestyle interventions (see Box 3);

studies involving antioxidants are documented in other reviews⁶⁰. Triheptanoin²⁶⁹ is a seven-carbon

triglyceride. The APP/PS1 AD mice bear the APP-Swedish mutation plus the PS1-L166P mutation. 3XTgAD

mice express three mutations (APP-Swedish, PS1-M146L and tau-P301L). Abbreviations not in main text:

AP39, proprietary mitochondrial-targeted H₂S donor; GSK3 β , glycogen synthase kinase 3 beta; mutUNG1,

mutated mitochondrial DNA repair enzyme; MPTP, 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine; SCA1,

spinocerebellar ataxia type 1; TBI, traumatic brain injury; YAC, yeast artificial chromosome.

Table 2 | Clinical trials reporting improved brain function and/or energetics in

neurodegenerative disorders

Disorder	Study details	Results and comments	Reference s
AD	Single blind RCT of S-equol (n=15) or placebo (n=15) for 2 weeks (NCT 02142777)	Well-tolerated. More participants showed↑cytochrome oxidase activity on S-equol than on placebo; no cognitive change. First study of a mitochondrial intervention as a direct biomarker of mitochondrial engagement in AD.	137
Mild– moderat e AD and MCI	RCT in AD (n=21) or MCI (n=39) receiving long-acting intranasal insulin (20 IU or 40 IU) or placebo for 4 months (NCT01595646)	Dose-dependent 个 memory composite score in ApoE4 ⁺ . No change in functional autonomy or executive function.	38
AD	RCT of liraglutide (n=14) or placebo (n=20) for 26 weeks (NCT01469351)	No change in Aβ load or cognitive scores. ↓ Brain glucose uptake over 26 weeks only on placebo. Underpowered for cognitive outcomes. Liraglutide may delay metabolic decline in brain.	194
AD or MCl	RCT (n=20) of metformin 500 mg or placebo for 8 weeks (NCT01965756)	\uparrow Executive function. No change in cerebral blood flow.	183
PD	RCT of exenatide 2 mg once per week (n=31) or placebo (n=29) for 48 weeks plus 12 week washout (NCT 01971242)	UPDRS motor subscale at 60 weeks \uparrow 1.0 points on drug and \downarrow by 2.1 points on placebo \downarrow motor symptoms.	97
AD	Double-blind RCT of C8 20 g per day (n=77) or placebo (n=63) for 90 days (NCT00142805)	ADAS-Cog ↑ by 3.4 points in ApoE4 ⁻ . Cognitive score varied directly as Ketones ↑ cognition in mild-to-moderate AD.	119
Mild- moderat e AD	Open label (n=10) receiving KD ± C8C10 for 12 weeks (NCT03690193)	↑ ADAS-Cog score; no cardiovascular safety or other metabolic concerns. First reported clinical use of ketogenic diet in AD including medium chain triglyceride supplementation.	157
AD and MCI	RCT of KD (n=9) or NIA low-fat diet (n=5) for 12 weeks (NCT02521818)	↑ Composite cognitive score, particularly memory domain, only in compliant participants and only on KD. First reported clinical use of a KD without medium chain triglyceride supplementation. Feasibility is very challenging but beneficial effects of ketones were clearly present.	162
Mild- moderat e AD	Open label study of C8C10 (n=11) or C8 (n=6) 30 g per day for 4 weeks (NCT02709356)	↑ Ketones 2-fold. Brain ketone uptake ↑ in direct proportion to ↑ ketone and brain glucose utilization. In AD, the brain can utilize additional ketones provided as C8C10.	270
Mild-to- moderat e ApoE4 ⁻ AD	Crossover RCT of MCT 17.3g (n=24) or placebo [canola] (n=25) for 30 days (ChiCTRIOR6009737)	2.62 point improvement on ADAS-Cog (Chinese version) on MCT, 2.57 worsening on placebo. Study restricted to ApoE4(–) patients. Inverse correlation between cognitive changes and plasma lysophosphatidylcholine species.	120
PD	RCT of KD (n=20) or low-fat diet (n=20) for 8 weeks (ACTRN 12617000027314)	↑ UPDRS in both groups, but 41% more on the KD. 86% compliance; tremor ± rigidity intermittently ↑ on KD. First RCT of KD in PD. KD and low-fat diets are safe in PD.	160
HD	Open label study in HD (n=10) and CTL (n=13) receiving triheptanoin 1 g/kg for 1 month (NCT01696708)	MRS ↑ brain high-energy phosphates including ↓ inorganic phosphate/phosphocreatine during visual stimulation	271

These studies reported statistically significant improvements in primary or secondary endpoints with novel treatments or drugs approved for other indications and repurposed for treatment of

neurodegenerative disorders of ageing. The low-fat diet¹⁶² diet was a modified Atkins diet. The three KD trials were all principally feasibility studies not powered for cognitive or metabolic outcomes^{157,161,163}. Abbreviations not in main text: ADAS-Cog, Alzheimer Disease Assessment Scale - Cognitive Subscale; AHAD – American Heart Association diet; ALSFRS-R, ALS functional rating scale –revised; MMKD – modified Mediterranean ketogenic diet; MRS – magnetic resonance spectroscopy; NIA, National Institutes of Aging; OAA, oxaloacetic acid; RCT, randomized controlled trial; SMC – subjective memory complaints; UPDRS; Unified Parkinson disease rating scale.

Table 3 | Clinical trials reporting improved brain function and/or energetics in major riskconditions for neurodegenerative disorders of ageing

Disorder	Study details	Results and comments	Refere nces
T2D ± cognitive impairment	Open label (n=205) DPP4I (sitagliptin) 100 mg per day ± metformin ± insulin (n=101) vs metformin ± insulin only (n=104) for 6 months	↑ MMSE on DPP4I; similar glycemic control in both groups; $↓$ insulin on DPP4I arm; RCT needed.	197
MCI or mild AD (non- diabetic)	Placebo-controlled, parallel- group cross-over study (n=10 per group) of metformin, 2 g per day for 8 weeks (NCT01965756)	CSF β , pTau; CBF; ADAS-Cog and CANTAB [®] cognitive batteries: metformin measurable in CSF; no change in CSF A β 42, pTau; \uparrow CBF in two brain regions at 8 wk; trend to \uparrow executive function, memory and attention. Metformin penetrates brain; underpowered for cognitive outcomes.	186
MCI	RCT of high carbohydrate (n=11) or high fat KD (n=12) for 6 weeks (NCT00777010)	 Feasibility study; cognitive outcomes (executive function, long-term memory), mood: on KD, ↑ memory (paired associate learning); no change in executive function or depression score. Metabolic improvement on KD: ↓ weight, waist circumference, fasting glucose, fasting insulin. 	155
MCI	RCT of C8C10 30 g per day (n=19) or energy-matched non-ketogenic placebo (n=20) for 6 months (NCT02551419)	Brain glucose and ketone status: \uparrow Brain ketone uptake 2- fold. \uparrow executive function, episodic memory, language and processing speed. Several cognitive outcomes improved in direct proportion to [ketone] and/or brain ketone uptake.	154
SMC and MCI	Cross-over RCT with 6 week washout in between 6 weeks of AHAD and 6 weeks of MMKD interventions, in patients with SMC (n=11) or MCI (n=9) (NCT2984540)	 CSF AD markers, neuroimaging markers, peripheral metabolic status, cognition: In MMKD only: ↑ CSF Ab42, ↓ CSF tau, ↑ brain ketone ↑ brain perfusion. In both groups: ↑ metabolic markers, ↑ memory. Compliance ≥90% in both groups; MMKD feasible, acceptable and has prevention effect on AD CSF biomarkers. 	156

ADAS-Cog, Alzheimer Disease Assessment Scale - Cognitive Subscale; AHAD – American Heart Association diet; ALSFRS-R, ALS functional rating scale –revised; MMKD – modified Mediterranean ketogenic diet; MMSE, Mini-mental state examination; MRS – magnetic resonance spectroscopy; OAA, oxaloacetic acid; RCT, randomized controlled trial; SMC – subjective memory complaints; UPDRS; Unified Parkinson disease rating scale.

Box 1 | Generating ATP in the brain

Glucose entering brain cells is phosphorylated to glucose-6-phospate and then enters one of two pathways¹⁴: generation of ATP via aerobic glycolysis and the mitochondrial tricarboxylic acid (TCA) cycle, or the pentose phosphate pathway which generates riboses, nucleic acids and NADPH for antioxidant defence and anabolic reactions (Figures 1, 3). The final step in aerobic glycolysis is generation of pyruvate, converted either to acetyl-CoA via pyruvate dehydrogenase for entry into the TCA cycle or to lactate via lactate dehydrogenase (Figures 1, 3). The TCA cycle pathway predominates in neurons and the lactate pathway in astrocytes, but this is not an absolute distinction. The TCA cycle takes place in the mitochondrial matrix, with oxidative phosphorylation occurring in the inner mitochondrial membrane. Several TCA cycle steps generate NADH and flavin adenine dinucleotide hydride, which are oxidized and donate high-energy electrons to the mitochondrial electron transport chain, which in turn drives conversion of ADP into ATP by oxidative phosphorylation. More than 90% of brain ATP is generated in mitochondria by oxidative phosphorylation. Each molecule of glucose consumed during oxidative phosphorylation generates about 33 ATP molecules²⁷², compared to 2 molecules of ATP produced by aerobic glycolysis. However, oxidative phosphorylation is slower than aerobic glycolysis and occurs at the price of generating reactive oxygen species⁶⁰.

The TCA cycle not only feeds the mitochondrial electron transport chain but also provides carbon for synthesis of glutamate and acetylcholine, a process called cataplerosis **[G]**. Carbon exiting the TCA cycle needs to be replaced in order to maintain TCA cycle activity, a process called anaplerosis. Via pyruvate, glucose is an important contributor to anaplerosis, such that when brain glucose uptake is impaired, not only ATP production but also anaplerosis are both adversely affected. Pyruvate carboxylase generates oxaloacetate, mostly in astrocytes but also oligodendrocytes²⁷. Certain branched-chain amino acids and odd-chain fatty acids (such as heptanoate) are anaplerotic²⁷³. Ketones can replace glucose as a source of acetyl CoA, but they are not anaplerotic.

The astrocyte–neuron lactate shuttle hypothesis postulates that activated glutamatergic neurons stimulate astrocytes to increase their supply of lactate to neurons⁸. Astrocytes release lactate through the low affinity perisynaptic monocarboxylic acid transporters (MCTs) 1 and 4¹⁸. Lactate is taken up by high-affinity neuronal MCT2 and transformed into ATP in neurons by oxidative phosphorylation (Figure 1). ATP produced by metabolism of glucose or glycogen to lactate does not require oxygen, so this route reduces net brain oxygen consumption in cells or

organelles with a high level of aerobic glycolysis, such as hippocampal synapses¹⁵. Lactate derived from glycogen in astrocytes can stimulate neuronal plasticity and learning, but this effect may only occur at relatively high levels of lactate (2–5 mM)⁸. Moreover, some types of neurons, including inhibitory GABAergic interneurons, do not necessarily depend on lactate⁸. Hence, the functional importance of the astrocyte–lactate shuttle *in vivo* requires additional study^{15,274,275}.

Lactate may act as a paracrine regulator through the lactate receptor, hydroxyl carboxylic acid receptor (HCAR) 1 (also known as GPR81). During intense exercise, plasma lactate increases and lactate enters the brain through MCTs at the blood–brain barrier (Figure 3), activating HCAR1 and promoting angiogenesis in the hippocampus and neocortex^{253,254}. Exogenously administered lactate enhances hippocampal synaptic plasticity, neurogenesis and memory formation in rodents⁸.

Box 2 | Measuring brain energy metabolism

[b1] In vitro models

To measure high-energy phosphates in the brain, fresh mitochondria are fixed and visualized by electron microscopy, and adenine nucleotides are measured by high-performance liquid chromatography. Frozen mitochondria can be used to evaluate antioxidants, oxidative damage and the activity of respiratory chain complexes. Frozen brain homogenates are used to determine the activity of glycolytic, tricarboxylic acid (TCA) cycle and electron transport chain complexes, and markers of mitochondrial biogenesis, dynamics and autophagy.

Regulatory mechanisms of energy homeostasis can be studied in single neurons and glia using nanosensors based on fluorescence resonance energy transfer, which detect changes in intracellular concentrations of cyclic AMP, protein kinase A, glucose and lactate in response to activation of G-protein-coupled receptors²⁷⁶. Intact brain cell respiration can be studied using an extracellular flux analyzer to establish rates of oxygen consumption, glycolysis, proton leakage, mitochondrial reserve and other bioenergetic parameters¹²⁹. The use of reprogrammed human cells and brain organoids for advancing bioenergetic drug discovery is considered in Supplementary Box 1.

Fluorescence lifetime imaging microscopy of cells or brain slices measures mitochondrial NADH production in real time and reveals a role for astrocytes in the glycolytic deficits of a mouse model of HD expressing mutant huntingtin⁶⁵. This technique identifies defective glycolysis as leading to mitochondrial dysfunction in AD neurons and supports the potential of pyruvate to bypass impaired glycolysis and maintain mitochondrial respiration¹⁴⁷. The energetics of neuronal network activity can also be studied in cortical slices³⁴. Gamma oscillations have high energy expenditure³⁴, in which pyruvate and BHB can partially replace glucose²⁷⁷.

[b1] Brain imaging in vivo

In living organisms, brain energy metabolism is usually evaluated by positron emission tomography (PET) in which the glucose analogue, deoxyglucose, is labelled with ¹⁸F to make ¹⁸F-fluorodeoxyglucose (FDG). To quantify brain uptake of FDG (or ketones or oxygen) by PET — that is, cerebral metabolic rate [G] (μ mol/100 g/min) — blood samples to measure the tracer must be obtained as near to the brain as possible. Cerebral metabolic rates obtained by PET reflect values obtained by arteriovenous difference across the brain³, but PET provides a visual image of both global and regional brain energy metabolism. A new PET tracer of mitochondrial complex 1

function, ¹⁸F-2-tert-butyl-4-chloro-5-2H pyridazin-3-one²⁷⁸, has been proposed as a marker of mitochondria-specific energy failure arising before the onset of impaired glycolysis in AD, and could be used to validate new therapeutics aiming to correct mitochondrial function.

In healthy ageing individuals, the brain glucose deficit (energy gap; Figure 3) is about 8% and occurs mostly in the frontal cortex, whereas in AD the parietal and temporal lobes are more adversely affected²⁷⁰. A glucose-specific brain energy deficit is also present in young adults with insulin resistance²⁷⁹. Indeed, brain FDG uptake might be a better marker of declining cognitive function in MCI and AD than the A β –PET marker, ¹⁸F-florbetapir²⁸⁰.

Metabolism of multiple energy substrates has been assessed in humans and animals by in vivo ¹³C-magnetic resonance spectroscopy, including brain uptake of ¹³C-labelled glucose and ketones²⁸¹, medium-chain fatty acids¹⁵³, and non-invasive assessment of mitochondrial redox status in the rat²⁸².

Box 3 | Complementary multi-modal lifestyle strategies

Lifestyle interventions may delay the onset of neurodegenerative diseases of ageing (NDAs), as exemplified by the 'Finger' trial which reduced the risk of AD in a typical elderly population^{126,251}. Two lifestyle approaches that improve brain energetics and insulin sensitivity are garnering considerable attention: caloric restriction (and the variant - intermittent fasting)⁵ and physical exercise¹²⁷. Both approaches are neuroprotective and improve cognitive and motor function in preclinical models of AD and Parkinson disease by increasing synaptic spine density⁹⁸, mitochondrial biogenesis²⁸³, neurogenesis in the hippocampus, autophagy of neurotoxic proteins, mitophagy of dysfunctional mitochondria^{98,284}, and activation of ghrelin signalling²⁰⁶. Nutritional ketosis is a feature common to caloric restriction, intermittent fasting and other ketogenic interventions^{3,98,285}.

Exercise helps regulate glucose metabolism and reduces two important risk factors for NDAs: obesity and type 2 diabetes. Exercise also improves executive function, attention and processing speed in NDAs — effects related to enhanced cerebral blood perfusion, notably in the hippocampal dentate gyrus. Exercise increases angiogenesis in several brain regions²⁵⁴ and mitigates the age- and NDA-related decline in cerebral blood flow⁹⁸ which, in turn, may improve synaptic function by providing ketones and lactate^{5,98,286,287}. A three-month exercise regimen increased brain ketone transport by 30% in AD¹⁰⁸, so the improvement in brain energetics by ketones is one possible link between exercise, brain-derived neurotrophic factor (BDNF), neurogenesis and cognitive gains in NDAs²⁸⁷.

The angiogenic effect of exercise is partly mediated by vascular endothelial growth factor^{253,254}. Lactate liberated from skeletal muscle during exercise can also be used by the brain^{5,287} (Box 1). Exogenous lactate mimicked exercise in inducing brain vascular endothelial growth factor and increasing capillary density, actions dependent on hydroxycarboxylic acid 1 (HCAR1) receptors²⁵⁴. The hippocampal myokine, irisin, may also be involved; both irisin and its precursor, fibronectin domain 5, contribute to metabolic homeostasis and neuroprotection²⁸⁷. Lactate recruitment of BDNF is dependent on fibronectin domain 5, thereby interlinking the actions of lactate to irisin in the beneficial effect of exercise on the brain^{222,287,288}.

The goal of mimicking the gains of exercise and fasting in a broadly accessible manner by an appropriate pharmacological intervention ("exercise in a pill") is analogous to using lactate²⁵³ or a ketogenic supplement to mimic and/or augment endogenous ketone production without

severely limiting dietary carbohydrate or food intake^{156,157}. Exercise mimetics could include agents acting *via* myokines, cathepsin B, AMPK or adiponectin^{98,132,287,289}.

Cognitive reserve is the capacity or resilience of the ageing brain to resist functional decline and is directly correlated with higher education and intellectual occupation both early and later in life. Whether improved cognitive reserve can stall AD is currently under exploration²⁹⁰. FDG-PET suggests that cognitive reserve reflects in part the capacity of the brain to maintain normal function in the face of bioenergetic or other deficits^{291,292}. Maintaining and improving cognitive reserve in individuals with NDAs could potentially be enhanced by the brain energy rescue strategies discussed herein.

Box 4 | Deteriorating brain glucose but not brain ketone uptake: an opportunity for brain energy rescue

The decline in brain glucose metabolism associated with NDAs has traditionally been assumed to be a *consequence* of the disease process. The development of PET tracers for assessing ketone uptake (Box 2) provided an opportunity to assess whether brain ketone metabolism was also disrupted. Dual-tracer PET studies of brain glucose (¹⁸F-FDG tracer) and ketone (¹¹C-acetoacetate tracer) uptake show that whereas brain glucose utilization is impaired, brain ketone metabolism is still normal in AD and MCI (see figure, panel **a**). These PET images show the rate constant (min⁻ ¹) for brain glucose uptake (K_{Glc} ; left) and brain acetoacetate uptake (K_{AcAc} ; right) in cognitively healthy, older controls (CTL, n = 24), mild cognitive impairment (MCI; n = 20) and mild–moderate AD (n = 19). The images are paired; that is, one for FDG and one for ¹¹C-acetoacetate obtained from each participant on the same afternoon. Unlike the cerebral metabolic rate (CMR), which is partly dependent on plasma concentrations of the substrate in question, K_{ACAC} is largely independent of plasma levels of glucose or ketones, so it is a better measure of the brain's *capacity* to take up these energy substrates. K_{Glc} is significantly lower in the parietal and temporal cortex as MCI develops and progresses to AD, but K_{AcAc} does not decrease in MCI or AD compared to cognitively unimpaired age-matched controls²⁹³. The CMR of acetoacetate increases in direct proportion to plasma ketone levels in AD after one month of receiving a supplement of 30 g per day of a ketogenic medium-chain triglyceride (C8C10 or C8; see figure, panel **b**). However, there was no change in the CMR of glucose²⁷⁰.

Mitochondrial oxidative phosphorylation is the only way of generating ATP from ketones; that is, there is no extra-mitochondrial pathway for ketones as there is for glucose to lactate. Accordingly, the fact that brain ketone metabolism is normal in MCI and AD indirectly implies that mitochondrial respiration is relatively normal in a significant proportion of brain mitochondria in order for them to be able to generate ATP. Hence, comparisons of brain glucose and brain ketone metabolism offer an opportunity to determine whether mitochondrial function is markedly impaired (in which case both glucose and ketone metabolism would be decreased, regionally or globally) or whether the defect is more at the level of glycolysis or glucose transport (in which case glucose but not ketone metabolism would be impaired). This PET comparison of brain energy substrate uptake could help to clarify whether the onset of mitochondrial dysfunction is an early event in NDAs, whether such dysfunction occurs in the brain regions most affected⁶⁵, and whether

a therapeutic agent being tested corrects the dysfunction or promotes mitochondrial biogenesis or other aspects of mitochondrial health.

Box 5 | Brain energy rescue in other central nervous system disorders

Owing to the success of combinations of antiretroviral drugs, patients infected with human immunodeficiency virus 1 are living longer. However, reflecting persistent neurological sequelae of the virus, about half of them develop cognitive deficits involving impaired glucose metabolism, mitochondrial dysfunction and reduced glial support of neurons. Brain energy rescue therefore might be useful to treat cognitive decline in these patients, and in people suffering long-term neurological repercussions of other classes of viral infection²⁹⁴.

Prions are infectious particles that *lack* DNA or RNA and, in Creutzfeldt–Jacob disease, brain glucose hypometabolism is seen in frontal and parietal regions and is related to sensory and motor dysfunction²⁹⁵. Lower brain perfusion and mitochondrial dysfunction are also observed¹⁰². Mouse models of prion diseases mimic clinical cases in displaying altered metabolism of glucose and fatty acids²⁹⁶. Interestingly, by binding to oligomeric Aβ, prion antagonists suppress synaptic pathology and cognitive deficits in mouse models of AD, underpinning the pertinence of prion disorders to NDAs²⁹⁷.

Both ischemic stroke (caused by blood vessel occlusion) and haemorrhagic stroke (caused by blood vessel rupture) involve an abrupt interruption of the supply of energy and oxygen to neurons, triggering neurodegeneration, acute brain energetic failure and functional deficits. Stroke management is driven by the twin goals of restoring blood flow and protecting mitochondrial function^{60,298} in which brain energetic rescue with ketones is being assessed²⁹⁹.

Like stroke, traumatic brain injury is associated not only with tissue damage but also with focal interruption of brain nutrient and energy supply. Strategies to restore mitochondrial function are under investigation in traumatic brain injury³⁰⁰. Clinical and preclinical studies suggest that ketogenic interventions might be therapeutically beneficial^{140,301}. Furthermore, hypertonic sodium lactate reduces intracranial pressure and compensates for the acute neuronal energetic crisis in traumatic brain injury³⁰².

Inherited GLUT1 deficiencies (de Vivo disease) are associated with neurodevelopmental delay, motor symptoms and seizures during infancy³⁰³. Seizures consume considerable energy, and adult-onset epilepsy is characterized by focal brain hypometabolism, decreased glucose uptake, defective TCA cycling and mitochondrial dysfunction^{34,299}. Seizures also occur with greater frequency in AD, so drugs that reinforce the GABAergic inhibition of hyperactive (energetically costly) networks⁴⁷ warrant assessment in AD. In MCI, task-induced hyperexcitability in the temporal lobe responded favourably to levetiracetam³⁰⁴, an anti-epileptic drug that promotes

vesicular release of GABA. In a mouse model of AD that overexpresses amyloid precursor protein, the addition of pyruvate and BHB to the diet reduced neuronal hyperexcitation and the incidence of epileptiform activity¹⁴⁶. Ketone-based interventions are also under study as a treatment for seizure-related disorders in adults because ketogenic diets are a well-established therapy for treating de Vivo disease and intractable epilepsy in children^{4,252,303}.

Glucose metabolism and the function of mitochondria and GABAergic interneurons are all impaired in the cortex, basal ganglia and other brain structures in schizophrenia^{34,227,305}. As in epilepsy, ketone-based interventions have been proposed as a treatment for schizophrenia^{306,307} and have shown encouraging results in two clinical case reports³⁰⁸.

Migraine is a highly debilitating and widespread form of headache. Energy deficits and/or excessive oxidative stress within the brain are attracting attention as possible triggers and, hence, as targets for metabolically focused therapeutic interventions³⁰⁹.

Finally, the retina is an outpost of the brain, and considerable progress has been made in understanding and potentially treating energetic disorders of the eye, such as age-related macular degeneration³¹⁰ (see also Supplementary Box 3).

FIGURE LEGENDS

Figure 1 | Energy supply and use by neurons and other brain cells. a |Organization of the neurovascular unit which functions to supply glucose (Glc) to neurons. **b** | Astrocytes mainly produce ATP via aerobic glycolysis (glucose to pyruvate), yet also exploit oxidative phosphorylation in mitochondria (ovals) to generate ATP using the tricyclic acid cycle (TCA). Astrocytes supply glucose to neurons and oligodendrocytes from capillaries and endogenous stores of glycogen, a reversible transformation. Astrocytes take up glutamate (Glu) released from synapses and convert it to glutamine (Gln), sent back to neurons: this metabolically costly operation relieves neurons of an energetic burden. Some Glu and Gln contributes carbon to the TCA via the intermediate, alpha-ketoglutarate (α -KG). Astrocytic generation of ketones from acetyl coenzyme A (AcCoA) and uptake of lactate and medium chain fatty acids is not shown for clarity. c | Oligodendrocytes insulate axons with myelin and deliver lactate to axons, transformed into pyruvate and then ATP by mitochondria. Axons promote their own energy supply by releasing glutamate to stimulate N-methyl D-aspartate receptors (NMDAR) on oligodendrocytes: this promotes membrane insertion of GLUT1 and increased oligodendrocyte uptake of glucose delivered as lactate to axons. d | Microglia support neurons by clearing pathogens, waste and toxic proteins but do not provide them with energy. They mainly generate ATP from glucose but also from free fatty acids and glutamine. Fatty acid and fructose (prominent in "modern diets") uptake by microglia is linked to neuroinflammation in NDAs. e | Neurons use transporters to acquire glucose and lactate from astrocytes or the vasculature. Food, adipose tissue and gut microbiota yield short chain fatty acids (SCFA) and triglycerides (TG) transformed by the liver into the ketones, beta-hydroxybutyrate (BHB) and acetoacetate (AcAc), taken up by neurons. Glucose enters the TCA via pyruvate and Acetyl Coenzyme A to yield ATP or the pentose phosphate pathway to provide (via Glycogen-6-phosphate, G6P) the antioxidant, glutathione (GSH), and nucleosides. Microtubule transport of mitochondria and other cargo is driven by ATP, partially generated by "onboard" glycolytic enzymes. See text and Supplementary Figure 1 for details. Abbreviations: AMPA, α -amino-3-hydroxy-5-methyl-4-isoxazolepropionic acid; EAAT, excitatory amino acid transporter; FABP, fatty acid binding protein; LDH1 – lactate dehydrogenase 1; LPL, lipoprotein lipase; MCT, monocarboxylate transporter; MGluR, metabotropic receptor; NAA, Nacetyl-aspartate; NMDAR – N-methyl D-aspartate receptor; Rib5P, ribonucleoside 5 phosphate; SLC, solute carrier; SNAT, sodium-coupled neutral amino acid transporter.

Figure 2 | Causes and consequences of the brain energy gap in neurodegenerative disorders. a Brain glucose hypometabolism occurs in conditions that increase the risk of Alzheimer disease (AD). The chronic brain energy gap and the neuropathological processes (dashed black arrow) both contribute to a vicious cycle leading to brain energy exhaustion and dysfunction. Brain energy rescue strategies (Figure 3, Tables 1–3) attempt to inhibit the positive feedback between the brain energy gap and neuropathology involving A β and phosphorylated tau. Hormones (principally insulin, adipokines and incretins), as well as synthetic agonists and insulin sensitizers, can influence brain energy rescue and inhibit the onset of neuropathology. **b** | Glucose contributes to about 95% of total brain fuel supply in cognitively healthy young adults, and ketones supply the remaining 5%. In cognitively healthy older adults, brain glucose uptake is decreased by about 9%, in people with mild cognitive impairment (MCI) by about 12% and in people with mild-moderate AD by about 18%. The magnitude of the brain energy gap is the difference in total brain fuel uptake (glucose and ketones combined) between healthy young adults and people with mild-to-moderate AD; that is, the therapeutic target for brain energy rescue in MCI and AD. The brain energy gap has not been rigorously quantified in neurodegenerative disorders of ageing other than AD.

Figure 3 | **Brain energy disruption and rescue strategies. a** | Several pathways of brain energy metabolism in neurons are disrupted (shown using dashed black arrows) in neurodegenerative disorders of ageing (specific disorders with declines are shown in the red boxes). Increased production of reaction oxygen species (ROS) and neuroinflammation that negatively affect brain energy levels are shown with a solid black arrow. The combination of impaired ATP production and increased levels of ROS contribute to declining brain function. **b** | Molecules or potential therapies implicated in brain energy rescue strategies target six broad pathways: ATP and redox state (light blue), brain glucose transport and/or aerobic glycolysis (blue; *intranasal insulin, adiponectin, ghrelin, insulin (GLUT4 only), nicotinamide riboside, dichloroacetate, *N*-acetyl-cysteine, oxaloacetate, GLP1, GIP, leptin, amylin, metformin, liraglutide, sitagliptin), anaplerosis and the TCA cycle (purple; C3, C7, C8, BHB, KE), mitochondrial transport and biogenesis (olive), ketogenesis (orange; ** BHB, C8, C10, KE) or protection against ROS and inflammation (light green; *** ghrelin, GLP1, GIP, leptin, adiponectin, metformin, AP-39, mdivi-1, mitoQ, BHB, KD and ketone esters). Details of the molecules or potential therapies are shown in Table 1 (preclinical

studies) and Tables 2 and 3 (clinical studies). Complementary interventions such as caloric restriction, ketogenic diet and exercise are not shown. Neurons take up lactate generated by astrocytes and oligodendrocytes (not shown). Medium-chain fatty acids such as decanoic and octanoic acid in the circulation can enter astrocytes and produce ketones and acetyl CoA. AAV, adeno-associated virus; AD, Alzheimer disease; ALS, amyotrophic lateral sclerosis; ASO, antisense oligonucleotide; ATP Syn, ATP synthase; BHB, D-β-hydroxybutyrate; C3, propionic acid, ETC, electron transport chain; FTD, frontotemporal dementia; GLP1, glucagon-like peptide 1; GIP, glucose-dependent insulinotropic polypeptide; HD, Huntington disease; MCT, monocarboxylate transporter; PD, Parkinson disease; PPP, pentose phosphate pathway.

Glossary

Neuroinflammation

An inflammatory response or state in the brain that involves functional, morphological and energetic shifts in microglia and "reactive" astrocytes, as well as macrophages that migrate into the brain from the periphery. It is a characteristic of neurodegenerative disorders and brain response to infectious agents or injury.

Ketone bodies

(Ketones), β -hydroxybutyrate and acetoacetate. Produced endogenously by fatty acid β -oxidation during caloric or severe carbohydrate restriction, and from medium-chain fatty acids. Exogenous ketones are mostly salts or esters of β -hydroxybutyrate. Acetone is a breakdown product of acetoacetate measurable in plasma and on breath.

Microglia

Resident brain macrophages of mesodermal origin that clear neurotoxic proteins and protect neurons from damaging exogenous molecules, toxins, infectious agents or pathogens. Excess and persistent microglial activation is associated with neuroinflammation, neuronal energetic deterioration and progression of neurodegenerative diseases of ageing.

Oligodendrocytes

Cells that produce myelin to insulate the axon and increase the speed of action potential propagation. They energetically support and communicate with neurons and astrocytes.

Neurovascular coupling

Coordinated response to brain activation involving local capillary dilation and a transitory surge in the flow of oxygenated, glucose-containing blood across the neurovascular unit, thereby replenishing ATP used in neurotransmission.

Insulin resistance

A state in which insulin is ineffective in stimulating glucose use by peripheral tissues and certain populations of neurons in the brain, due mainly to receptor-signalling desensitization. It is

associated with glucose intolerance and type 2 diabetes, and increases the risk of neurodegenerative disorders, particularly Alzheimer disease.

Oxidative phosphorylation

Process by which mitochondria generate ATP by conveying electrons through enzyme complexes (I to IV), thereby creating a proton gradient that powers phosphorylation of ADP to ATP by ATP synthase.

Tricarboxylic acid cycle

(TCA cycle). Process by which acetyl CoA is oxidized to form GTP, FADH₂ and NADH. NADH and FADH₂ feed electrons to the electron transport chain to produce ATP by oxidative phosphorylation. Several neurotransmitters (acetylcholine, glutamate, GABA) are produced by carbon leaving the TCA cycle.

Aerobic glycolysis

Conversion of glucose to pyruvate by the "Emden–Meyerhoff pathway". Pyruvate is either converted to acetyl-CoA and enters the TCA cycle or is reduced to lactate by NADH, a pathway prominent in glia to produce ATP without oxygen. Aerobic glycolysis may also occur in neurons.

Astrocyte-neuron lactate shuttle

Hypothesis that lactate produced in astrocytes is delivered to neurons to support the energy requirements of neurotransmission.

Fast axonal transport

Rapid transport of vesicles, mitochondria and other cargo along axonal microtubules. Vesicles are equipped with molecular motors (kinesin and dynein) and glycolytic enzymes, permitting rapid, local ATP production by aerobic glycolysis.

Incretins

Peptide hormones produced by the small intestine that stimulate pancreatic insulin secretion, regulate glucose metabolism and influence cognition. These include glucagon-like peptide 1 and glucose-dependent insulinotropic polypeptide.

Monocarboxylic acid transporters

Transporters in the cell membrane that facilitate unidirectional, proton-linked transport (uptake) of small monocarboxylic acids such as lactate and ketones.

Short-chain fatty acids

Acetate (2 carbons), propionate (3 carbons), and butyrate (4 carbons). End-products of microbial fermentation of dietary polysaccharides (soluble fiber). Butyrate is ketogenic and propionate is anaplerotic.

Mild cognitive impairment

(MCI). Condition prodromal to Alzheimer disease characterized by a subjective memory complaint and modest deficits in at least one of five main cognitive domains (executive function, memory, language, processing speed or attention). About 50% of cases progress to Alzheimer disease within 5 years.

Caloric restriction

Limiting food intake to a level that does not permit full satiety. Can be self-determined (usually the case in human studies) or imposed relative to the food consumed by a matched group fed *ad libitum* (usually only in animal studies).

Electron transport chain

A series of enzymatic protein complexes in the inner mitochondrial membrane that transfer electrons donated from NADH (complex 1) or fatty acid dehydrogenase (complex 2) to oxygen (complex 4).

Medium-chain triglycerides

Edible oils comprised of saturated fatty acids of 6-14 carbons in length. These have long been used in clinical nutrition to support energy needs in diseases or conditions involving malabsorption. Eight-carbon medium-chain triglycerides are more ketogenic than those of 10 or 12 carbons.

Mitochondrial biogenesis

Renewal of mitochondria. In neurons, mitochondrial biogenesis occurs in the cell body with newly formed mitochondria being transported along the axon to dendritic synapses.

Redox state

Capacity of a molecule to be 'reduced' or acquire electrons; opposite of oxidation. Many biological reactions involve the reduction of one molecular species while another is being simultaneously oxidized. Energy metabolism is highly dependent on the redox state of the cell.

Ketogenic diet

A very low carbohydrate, very high fat diet inciting the liver to produce ketones from free fatty acids released from adipose tissue because there is minimal insulin production. The stricter, medical form of the ketogenic diet developed to treat intractable epilepsy usually also limits dietary protein.

Anaplerosis

Process by which 4 or 5 carbon units enter the TCA cycle independently of acetyl CoA to replenish intermediates used in the synthesis of acetylcholine or lipids (from citrate) or amino acids (from alpha-ketoglutarate and oxaloacetate); opposite of cataplerosis.

Brain energy gap

Deficit in brain energy metabolism on the order of 10% in mild cognitive impairment, and 20% in Alzheimer disease. Also present in other NDAs. It appears to be specific to glucose inasmuch as no studies to date have shown that brain ketone metabolism is affected.

Antagomirs

Also known as anti-microRNAs or blockmirs. Synthetic oligonucleotides engineered to silence endogenous microRNAs or prevent other molecules from binding to a specific mRNA.

Locked nucleic acids

RNAs in which the flexibility of the ribose ring has been restrained by adding a methylene bridge connecting the 2'oxygen and 4' carbon. Oligonucleotides containing locked nucleic acids have improved specificity, sensitivity and hybridization stability.

Cataplerosis

Process by which intermediates (carbon) leave the TCA cycle to support biochemical reactions; that is, acetylcholine and lipid synthesis from citrate, or amino acid synthesis from α -ketoglutarate and oxaloacetate; opposite of anaplerosis.

Cerebral metabolic rate

Quantity of energy substrate consumed by the brain (Imol/100 g/min). Typically refers to glucose, but also used for brain consumption of oxygen, lactate and ketones.

References

- 1 Aldana, B. I. Microglia-specific metabolic changes in neurodegeneration. *J Mol Biol* **431**, 1830-1842, doi:10.1016/j.jmb.2019.03.006 (2019).
- 2 Boland, B. *et al.* Promoting the clearance of neurotoxic proteins in neurodegenerative disorders of ageing. *Nature Reviews Drug Discovery* **17**, 660-688, doi:10.1038/nrd.2018.109 (2018).
- 3 Cunnane, S. C. *et al.* Can ketones help rescue brain fuel supply in later life? Implications for cognitive health during aging and the treatment of alzheimer's disease. *Frontiers in Molecular Neuroscience* **9**, doi:10.3389/fnmol.2016.00053 (2016).
- 4 Zilberter, Y. & Zilberter, M. The vicious circle of hypometabolism in neurodegenerative diseases: Ways and mechanisms of metabolic correction. *Journal of Neuroscience Research* **95**, 2217-2235, doi:10.1002/jnr.24064 (2017).
- 5 Camandola, S. & Mattson, M. P. Brain metabolism in health, aging, and neurodegeneration. *The EMBO Journal* **36**, 1474-1492, doi:10.15252/embj.201695810 (2017).
- 6 Wilson, H., Pagano, G. & Politis, M. Dementia spectrum disorders: Lessons learnt from decades with pet research. *Journal of Neural Transmission* **126**, 233-251, doi:10.1007/s00702-019-01975-4 (2019).
- 7 Johnson, E. C. B. *et al.* Large-scale proteomic analysis of alzheimer's disease brain and cerebrospinal fluid reveals early changes in energy metabolism associated with microglia and astrocyte activation. *Nat Med*, doi:10.1038/s41591-020-0815-6 (2020).
- 8 Magistretti, P. J. & Allaman, I. Lactate in the brain: From metabolic end-product to signalling molecule. *Nature Reviews Neuroscience* **19**, 235-249, doi:10.1038/nrn.2018.19 (2018).
- 9 Wang, A., Luan, H. H. & Medzhitov, R. An evolutionary perspective on immunometabolism. *Science* 363, eaar3932, doi:10.1126/science.aar3932 (2019).
- 10 Tups, A., Benzler, J., Sergi, D., Ladyman, S. R. & Williams, L. M. in *Comprehensive physiology* 741-764 (John Wiley & Sons, Inc., 2017).
- 11 Caron, A. & Richard, D. Neuronal systems and circuits involved in the control of food intake and adaptive thermogenesis. *Annals of the New York Academy of Sciences* **1391**, 35-53, doi:10.1111/nyas.13263 (2016).
- 12 Dodd, G. T. *et al.* Insulin regulates pomc neuronal plasticity to control glucose metabolism. *Elife* **7**, doi:10.7554/eLife.38704 (2018).
- 13 Oyarzabal, A. & Marin Valencia, I. Synaptic energy metabolism and neuronal excitability, in sickness and health. *Journal of Inherited Metabolic Disease* **42**, 220-236, doi:10.1002/jimd.12071 (2019).
- 14 Bordone, M. P. *et al.* The energetic brain a review from students to students. *J Neurochem* **151**, 139-165, doi:10.1111/jnc.14829 (2019).
- Dienel, G. A. Brain glucose metabolism: Integration of energetics with function. *Physiological Reviews* **99**, 949-1045, doi:10.1152/physrev.00062.2017 (2019).
- 16 Engl, E. & Attwell, D. Non-signalling energy use in the brain. *The Journal of Physiology* **593**, 3417-3429, doi:10.1113/jphysiol.2014.282517 (2015).
- 17 Ashrafi, G., Wu, Z., Farrell, R. J. & Ryan, T. A. Glut4 mobilization supports energetic demands of active synapses. *Neuron* **93**, 606-615.e603, doi:10.1016/j.neuron.2016.12.020 (2017).
- 18 Gundersen, V., Storm-Mathisen, J. & Bergersen, L. H. Neuroglial transmission. *Physiological Reviews* **95**, 695-726, doi:10.1152/physrev.00024.2014 (2015).

- 19 Cheng, J. *et al.* Targeting pericytes for therapeutic approaches to neurological disorders. *Acta Neuropathologica* **136**, 507-523, doi:10.1007/s00401-018-1893-0 (2018).
- 20 Lecrux, C., Bourourou, M. & Hamel, E. How reliable is cerebral blood flow to map changes in neuronal activity? *Autonomic Neuroscience* **217**, 71-79, doi:10.1016/j.autneu.2019.01.005 (2019).
- 21 Saab, Aiman S. *et al.* Oligodendroglial nmda receptors regulate glucose import and axonal energy metabolism. *Neuron* **91**, 119-132, doi:10.1016/j.neuron.2016.05.016 (2016).
- 22 Pearson-Leary, J., Jahagirdar, V., Sage, J. & McNay, E. C. Insulin modulates hippocampally-mediated spatial working memory via glucose transporter-4. *Behavioural Brain Research* **338**, 32-39, doi:10.1016/j.bbr.2017.09.033 (2018).
- 23 Barros, L. F., Brown, A. & Swanson, R. A. Glia in brain energy metabolism: A perspective. *Glia* **66**, 1134-1137, doi:10.1002/glia.23316 (2018).
- 24 Waitt, A. E., Reed, L., Ransom, B. R. & Brown, A. M. Emerging roles for glycogen in the CNS. *Frontiers in Molecular Neuroscience* **10**, doi:10.3389/fnmol.2017.00073 (2017).
- 25 Nave, K.-A. & Werner, H. B. Myelination of the nervous system: Mechanisms and functions. *Annual Review of Cell and Developmental Biology* **30**, 503-533, doi:10.1146/annurev-cellbio-100913-013101 (2014).
- 26 Tomassy, G. S. *et al.* Distinct profiles of myelin distribution along single axons of pyramidal neurons in the neocortex. *Science* **344**, 319-324, doi:10.1126/science.1249766 (2014).
- 27 Amaral, A. I., Hadera, M. G., Kotter, M. & Sonnewald, U. Oligodendrocytes do not export NAA-derived aspartate in vitro. *Neurochem Res* **42**, 827-837, doi:10.1007/s11064-016-1985-y (2017).
- 28 Trevisiol, A. *et al.* Monitoring ATP dynamics in electrically active white matter tracts. *eLife* **6**, doi:10.7554/elife.24241 (2017).
- Hinckelmann, M.-V. *et al.* Self-propelling vesicles define glycolysis as the minimal energy machinery for neuronal transport. *Nature Communications* 7, doi:10.1038/ncomms13233 (2016).
- 30 Deczkowska, A. *et al.* Disease-associated microglia: A universal immune sensor of neurodegeneration. *Cell* **173**, 1073-1081, doi:10.1016/j.cell.2018.05.003 (2018).
- 31 Frere, S. & Slutsky, I. Alzheimer's disease: From firing instability to homeostasis network collapse. *Neuron* **97**, 32-58, doi:10.1016/j.neuron.2017.11.028 (2018).
- 32 Jessen, S. B., Mathiesen, C., Lind, B. L. & Lauritzen, M. Interneuron deficit associates attenuated network synchronization to mismatch of energy supply and demand in aging mouse brains. *Cerebral Cortex* **27**, 646-659, doi:10.1093/cercor/bhv261 (2017).
- 33 Micheva, K. D. *et al.* Distinctive structural and molecular features of myelinated inhibitory axons in human neocortex. *eneuro* **5**, ENEURO.0297-0218.2018, doi:10.1523/eneuro.0297-18.2018 (2018).
- 34 Kann, O. The interneuron energy hypothesis: Implications for brain disease. *Neurobiology of Disease* **90**, 75-85, doi:10.1016/j.nbd.2015.08.005 (2016).
- Illarioshkin, S. N., Klyushnikov, S. A., Vigont, V. A., Seliverstov, Y. A. & Kaznacheyeva, E.
 V. Molecular pathogenesis in huntington's disease. *Biochemistry (Moscow)* 83, 1030-1039, doi:10.1134/s0006297918090043 (2018).
- 36 Griffith, C. M., Eid, T., Rose, G. M. & Patrylo, P. R. Evidence for altered insulin receptor signaling in alzheimer's disease. *Neuropharmacology* **136**, 202-215, doi:10.1016/j.neuropharm.2018.01.008 (2018).

- 37 Duarte, A. I., Santos, M. S., Oliveira, C. R. & Moreira, P. I. Brain insulin signalling, glucose metabolism and females' reproductive aging: A dangerous triad in alzheimer's disease. *Neuropharmacology* **136**, 223-242, doi:10.1016/j.neuropharm.2018.01.044 (2018).
- 38 Craft, S. *et al.* Effects of regular and long-acting insulin on cognition and alzheimer's disease biomarkers: A pilot clinical trial. *Journal of Alzheimer's Disease* **57**, 1325-1334, doi:10.3233/jad-161256 (2017).
- 39 Bak, L. K., Walls, A. B., Schousboe, A. & Waagepetersen, H. S. Astrocytic glycogen metabolism in the healthy and diseased brain. *Journal of Biological Chemistry* 293, 7108-7116, doi:10.1074/jbc.r117.803239 (2018).
- 40 Cunnane, S. C. & Crawford, M. A. Energetic and nutritional constraints on infant brain development: Implications for brain expansion during human evolution. *Journal of Human Evolution* **77**, 88-98, doi:10.1016/j.jhevol.2014.05.001 (2014).
- 41 Courchesne-Loyer, A. *et al.* Inverse relationship between brain glucose and ketone metabolism in adults during short-term moderate dietary ketosis: A dual tracer quantitative positron emission tomography study. *Journal of Cerebral Blood Flow & Metabolism* **37**, 2485-2493, doi:10.1177/0271678x16669366 (2016).
- 42 Cani, P. D. Is colonic propionate delivery a novel solution to improve metabolism and inflammation in overweight or obese subjects? *Gut* **68**, 1352-1353, doi:10.1136/gutjnl-2019-318776 (2019).
- Spielman, L. J., Gibson, D. L. & Klegeris, A. Unhealthy gut, unhealthy brain: The role of the intestinal microbiota in neurodegenerative diseases. *Neurochemistry International* 120, 149-163, doi:10.1016/j.neuint.2018.08.005 (2018).
- 44 de Vadder, F. & Mithieux, G. Gut-brain signaling in energy homeostasis: The unexpected role of microbiota-derived succinate. *Journal of Endocrinology* **236**, R105-R108, doi:10.1530/joe-17-0542 (2018).
- 45 Olson, C. A. *et al.* The gut microbiota mediates the anti-seizure effects of the ketogenic diet. *Cell* **174**, 497, doi:10.1016/j.cell.2018.06.051 (2018).
- 46 Soty, M., Gautier-Stein, A., Rajas, F. & Mithieux, G. Gut-brain glucose signaling in energy homeostasis. *Cell Metabolism* **25**, 1231-1242, doi:10.1016/j.cmet.2017.04.032 (2017).
- 47 Zott, B., Busche, M. A., Sperling, R. A. & Konnerth, A. What happens with the circuit in alzheimer's disease in mice and humans? *Annual Review of Neuroscience* **41**, 277-297, doi:10.1146/annurev-neuro-080317-061725 (2018).
- 48 Yu, L., Shen, Z., Wang, C. & Yu, Y. Efficient coding and energy efficiency are promoted by balanced excitatory and inhibitory synaptic currents in neuronal network. *Front Cell Neurosci* **12**, 123, doi:10.3389/fncel.2018.00123 (2018).
- Ta, T.-T. *et al.* Priming of microglia with ifn-γ slows neuronal gamma oscillations in situ.
 Proceedings of the National Academy of Sciences **116**, 4637-4642,
 doi:10.1073/pnas.1813562116 (2019).
- 50 Briston, T. & Hicks, A. R. Mitochondrial dysfunction and neurodegenerative proteinopathies: Mechanisms and prospects for therapeutic intervention. *Biochemical Society Transactions* **46**, 829-842, doi:10.1042/bst20180025 (2018).
- 51 Oliveira, L. T. *et al.* Exogenous β-amyloid peptide interferes with glut4 localization in neurons. *Brain Research* **1615**, 42-50, doi:10.1016/j.brainres.2015.04.026 (2015).
- 52 Ryu, J. C., Zimmer, E. R., Rosa-Neto, P. & Yoon, S. O. Consequences of metabolic disruption in alzheimer's disease pathology. *Neurotherapeutics* **16**, 600-610, doi:10.1007/s13311-019-00755-y (2019).
- 53 An, Y. *et al.* Evidence for brain glucose dysregulation in alzheimer's disease. *Alzheimers Dement* **14**, 318-329, doi:10.1016/j.jalz.2017.09.011 (2018).

- 54 Toppala, S. *et al.* Midlife insulin resistance as a predictor for late-life cognitive function and cerebrovascular lesions. *J Alzheimers Dis* **72**, 215-228, doi:10.3233/jad-190691 (2019).
- 55 Bartzokis, G. Alzheimer's disease as homeostatic responses to age-related myelin breakdown. *Neurobiology of Aging* **32**, 1341-1371, doi:10.1016/j.neurobiolaging.2009.08.007 (2011).
- 56 Klosinski, L. P. *et al.* White matter lipids as a ketogenic fuel supply in aging female brain: Implications for alzheimer's disease. *EBioMedicine* **2**, 1888-1904, doi:10.1016/j.ebiom.2015.11.002 (2015).
- 57 Roy, M. *et al.* Fascicle- and glucose-specific deterioration in white matter energy supply in alzheimer's disease. *Journal of Alzheimer's Disease* (2020).
- 58 Matthews, D. C. *et al.* Fdg pet parkinson's disease-related pattern as a biomarker for clinical trials in early stage disease. *NeuroImage: Clinical* **20**, 572-579, doi:10.1016/j.nicl.2018.08.006 (2018).
- 59 Chu, J. S. *et al.* The metabolic activity of caudate and prefrontal cortex negatively correlates with the severity of idiopathic parkinson's disease. *Aging Dis* **10**, 847-853, doi:10.14336/ad.2018.0814 (2019).
- 60 Murphy, M. P. & Hartley, R. C. Mitochondria as a therapeutic target for common pathologies. *Nature Reviews Drug Discovery* **17**, 865-886, doi:10.1038/nrd.2018.174 (2018).
- 61 Zambon, F. *et al.* Cellular alpha-synuclein pathology is associated with bioenergetic dysfunction in parkinson's ipsc-derived dopamine neurons. *Hum Mol Genet* 28, 2001-2013, doi:10.1093/hmg/ddz038 (2019).
- 62 McColgan, P. *et al.* Brain regions showing white matter loss in huntington's disease are enriched for synaptic and metabolic genes. *Biol Psychiatry* **83**, 456-465, doi:10.1016/j.biopsych.2017.10.019 (2018).
- 63 Morea, V. *et al.* Glucose transportation in the brain and its impairment in huntington disease: One more shade of the energetic metabolism failure? *Amino Acids* **49**, 1147-1157, doi:10.1007/s00726-017-2417-2 (2017).
- 64 Liot, G., Valette, J., Pepin, J., Flament, J. & Brouillet, E. Energy defects in huntington's disease: Why "in vivo" evidence matters. *Biochem Biophys Res Commun* **483**, 1084-1095, doi:10.1016/j.bbrc.2016.09.065 (2017).
- 65 Polyzos, A. A. *et al.* Metabolic reprogramming in astrocytes distinguishes region-specific neuronal susceptibility in huntington mice. *Cell Metabolism* **29**, 1258-1273.e1211, doi:10.1016/j.cmet.2019.03.004 (2019).
- Kedaigle, A. J. *et al.* Bioenergetic deficits in huntington's disease ipsc-derived neural cells and rescue with glycolytic metabolites. *Hum Mol Genet*, doi:10.1093/hmg/ddy430 (2019).
- 67 Saudou, F. & Humbert, S. The biology of huntingtin. *Neuron* **89**, 910-926, doi:10.1016/j.neuron.2016.02.003 (2016).
- 68 Ahmed, R. M. *et al.* Amyotrophic lateral sclerosis and frontotemporal dementia: Distinct and overlapping changes in eating behaviour and metabolism. *The Lancet Neurology* **15**, 332-342, doi:10.1016/s1474-4422(15)00380-4 (2016).
- 69 Jawaid, A., Khan, R., Polymenidou, M. & Schulz, P. E. Disease-modifying effects of metabolic perturbations in ALS/FTLD. *Molecular Neurodegeneration* 13, doi:10.1186/s13024-018-0294-0 (2018).

- 70 Vandoorne, T., De Bock, K. & Van Den Bosch, L. Energy metabolism in als: An underappreciated opportunity? *Acta Neuropathologica* **135**, 489-509, doi:10.1007/s00401-018-1835-x (2018).
- 71 Tefera, T. W., Bartlett, K., Tran, S. S., Hodson, M. P. & Borges, K. Impaired pentose phosphate pathway in the spinal cord of the hsod1(g93a) mouse model of amyotrophic lateral sclerosis. *Mol Neurobiol* **56**, 5844-5855, doi:10.1007/s12035-019-1485-6 (2019).
- 72 Lau, D. H. W. *et al.* Disruption of er–mitochondria signalling in fronto-temporal dementia and related amyotrophic lateral sclerosis. *Cell Death & Disease* 9, doi:10.1038/s41419-017-0022-7 (2018).
- 73 Delic, V. *et al.* Discrete mitochondrial aberrations in the spinal cord of sporadic als patients. *Journal of Neuroscience Research* **96**, 1353-1366, doi:10.1002/jnr.24249 (2018).
- 74 Tefera, T. W. & Borges, K. Metabolic dysfunctions in amyotrophic lateral sclerosis pathogenesis and potential metabolic treatments. *Frontiers in Neuroscience* **10**, doi:10.3389/fnins.2016.00611 (2017).
- 75 Allen, S. P. *et al.* C9orf72 expansion within astrocytes reduces metabolic flexibility in amyotrophic lateral sclerosis. *Brain* **142**, 3771-3790, doi:10.1093/brain/awz302 (2019).
- 76 Sintini, I. *et al.* Regional multimodal relationships between tau, hypometabolism, atrophy, and fractional anisotropy in atypical alzheimer's disease. *Human Brain Mapping*, doi:10.1002/hbm.24473 (2018).
- Carbonell, F., Zijdenbos, A. P., Bedell, B. J. & for the Alzheimer's Disease Neuroimaging,
 I. Spatially distributed amyloid-β reduces glucose metabolism in mild cognitive
 impairment. *Journal of Alzheimer's Disease* 73, 543-557, doi:10.3233/JAD-190560
 (2020).
- 78 Butterfield, D. A. & Halliwell, B. Oxidative stress, dysfunctional glucose metabolism and alzheimer disease. *Nature Reviews Neuroscience* **20**, 148-160, doi:10.1038/s41583-019-0132-6 (2019).
- 79 Velliquette, R. A., O'Connor, T. & Vassar, R. Energy inhibition elevates β-secretase levels and activity and is potentially amyloidogenic in app transgenic mice: Possible early events in Alzheimer's disease pathogenesis. *The Journal of Neuroscience* **25**, 10874-10883, doi:10.1523/jneurosci.2350-05.2005 (2005).
- 80 Correia, S. C., Perry, G. & Moreira, P. I. Mitochondrial traffic jams in alzheimer's disease pinpointing the roadblocks. *Biochimica et Biophysica Acta (BBA) - Molecular Basis of Disease* **1862**, 1909-1917, doi:10.1016/j.bbadis.2016.07.010 (2016).
- 81 Ashraf, A., Fan, Z., Brooks, D. J. & Edison, P. Cortical hypermetabolism in mci subjects: A compensatory mechanism? *European Journal of Nuclear Medicine and Molecular Imaging* **42**, 447-458, doi:10.1007/s00259-014-2919-z (2014).
- Li, H., Liu, C.-C., Zheng, H. & Huang, T. Y. Amyloid, tau, pathogen infection and antimicrobial protection in alzheimer's disease –conformist, nonconformist, and realistic prospects for ad pathogenesis. *Translational Neurodegeneration* 7, doi:10.1186/s40035-018-0139-3 (2018).
- 83 Fülöp, T., Larbi, A. & Witkowski, Jacek M. Human inflammaging. *Gerontology* **65**, 495-504, doi:10.1159/000497375 (2019).
- 84 Millan, M. J., Rivet, J.-M. & Gobert, A. The frontal cortex as a network hub controlling mood and cognition: Probing its neurochemical substrates for improved therapy of psychiatric and neurological disorders. *Journal of Psychopharmacology* **30**, 1099-1128, doi:10.1177/0269881116672342 (2016).

- Yang, L., Wang, H., Liu, L. & Xie, A. The role of insulin/IGF-1/PI3K/AKT/GSK3β signaling in parkinson's disease dementia. *Frontiers in Neuroscience* 12, doi:10.3389/fnins.2018.00073 (2018).
- 86 Forny-Germano, L., De Felice, F. G. & Vieira, M. N. d. N. The role of leptin and adiponectin in obesity-associated cognitive decline and alzheimer's disease. *Frontiers in Neuroscience* 12, doi:10.3389/fnins.2018.01027 (2019).
- 87 McGregor, G. & Harvey, J. Regulation of hippocampal synaptic function by the metabolic hormone, leptin: Implications for health and neurodegenerative disease. *Frontiers in Cellular Neuroscience* **12**, doi:10.3389/fncel.2018.00340 (2018).
- 88 Shi, L., Du, X., Jiang, H. & Xie, J. Ghrelin and neurodegenerative disorders—a review. *Molecular Neurobiology* **54**, 1144-1155, doi:10.1007/s12035-016-9729-1 (2016).
- 89 Suda, Y. *et al.* Down-regulation of ghrelin receptors on dopaminergic neurons in the substantia nigra contributes to parkinson's disease-like motor dysfunction. *Molecular Brain* **11**, doi:10.1186/s13041-018-0349-8 (2018).
- 90 Frago, L. & Chowen, J. Involvement of astrocytes in mediating the central effects of ghrelin. *International Journal of Molecular Sciences* **18**, 536, doi:10.3390/ijms18030536 (2017).
- 91 Fu, W., Patel, A., Kimura, R., Soudy, R. & Jhamandas, J. H. Amylin receptor: A potential therapeutic target for Alzheimer's disease. *Trends in Molecular Medicine* **23**, 709-720, doi:10.1016/j.molmed.2017.06.003 (2017).
- 92 Grizzanti, J., Corrigan, R., Servizi, S. & Casadesus, G. Amylin signaling in diabetes and alzheimer's disease: Therapy or pathology? *Journal of Neurology & Neuromedicine* **4**, 12-16, doi:10.29245/2572.942x/2019/1.1212 (2019).
- 93 Mietlicki-Baase, E. G. Amylin in Alzheimer's disease: Pathological peptide or potential treatment? *Neuropharmacology* **136**, 287-297, doi:10.1016/j.neuropharm.2017.12.016 (2018).
- 94 Kim, M. W. *et al.* Suppression of adiponectin receptor 1 promotes memory dysfunction and alzheimer's disease-like pathologies. *Scientific Reports* **7**, doi:10.1038/s41598-017-12632-9 (2017).
- 95 Ngo, S. T. *et al.* Altered expression of metabolic proteins and adipokines in patients with amyotrophic lateral sclerosis. *Journal of the Neurological Sciences* **357**, 22-27, doi:10.1016/j.jns.2015.06.053 (2015).
- 96 Ma, J. *et al.* Peripheral blood adipokines and insulin levels in patients with Alzheimer's disease: A replication study and meta-analysis. *Current Alzheimer Research* **13**, 223-233, doi:10.2174/156720501303160217111434 (2016).
- 97 Athauda, D. & Foltynie, T. The glucagon-like peptide 1 (GLP) receptor as a therapeutic target in Parkinson's disease: Mechanisms of action. *Drug Discovery Today* **21**, 802-818, doi:10.1016/j.drudis.2016.01.013 (2016).
- 98 Mattson, M. P., Moehl, K., Ghena, N., Schmaedick, M. & Cheng, A. Intermittent metabolic switching, neuroplasticity and brain health. *Nature Reviews Neuroscience* **19**, 80-80, doi:10.1038/nrn.2017.156 (2018).
- 99 Nugent, S. *et al.* Glucose hypometabolism is highly localized, but lower cortical thickness and brain atrophy are widespread in cognitively normal older adults. *American journal of physiology.Endocrinology and metabolism* **306**, E1315-1321, doi:10.1152/ajpendo.00067.2014; 10.1152/ajpendo.00067.2014 (2014).
- 100 Castellano, C.-A. *et al.* Links between metabolic and structural changes in the brain of cognitively normal older adults: A 4-year longitudinal follow-up. *Frontiers in Aging Neuroscience* **11**, doi:10.3389/fnagi.2019.00015 (2019).

- 101 Goyal, M. S. *et al.* Loss of brain aerobic glycolysis in normal human aging. *Cell Metab* **26**, 353-360.e353, doi:10.1016/j.cmet.2017.07.010 (2017).
- 102 de la Torre, J. c. Are major dementias triggered by poor blood flow to the brain? Theoretical considerations. *Journal of Alzheimer's Disease* **57**, 353-371, doi:10.3233/jad-161266 (2017).
- 103 Sweeney, M. D. *et al.* Vascular dysfunction-the disregarded partner of Alzheimer's disease. *Alzheimers Dement* **15**, 158-167, doi:10.1016/j.jalz.2018.07.222 (2019).
- 104 Wingo, A. P. *et al.* Large-scale proteomic analysis of human brain identifies proteins associated with cognitive trajectory in advanced age. *Nature Communications* **10**, doi:10.1038/s41467-019-09613-z (2019).
- 105 de la Monte, S. M. in *Advances in Experimental Medicine and Biology* 45-83 (Springer Singapore, 2019).
- 106 Li, S. & Laher, I. Exercise pills: At the starting line. *Trends in Pharmacological Sciences* **36**, 906-917, doi:10.1016/j.tips.2015.08.014 (2015).
- 107 Vieira, M. N. N., Lima-Filho, R. A. S. & De Felice, F. G. Connecting Alzheimer's disease to diabetes: Underlying mechanisms and potential therapeutic targets. *Neuropharmacology* **136**, 160-171, doi:10.1016/j.neuropharm.2017.11.014 (2018).
- 108 Castellano, C.-A. *et al.* A 3-month aerobic training program improves brain energy metabolism in mild Alzheimer's disease: Preliminary results from a neuroimaging study. *Journal of Alzheimer's Disease* **56**, 1459-1468, doi:10.3233/jad-161163 (2017).
- 109 Penninx, B. & Lange, S. M. M. Metabolic syndrome in psychiatric patients: Overview, mechanisms, and implications. *Dialogues Clin Neurosci* **20**, 63-73 (2018).
- 110 Verdile, G., Fuller, S. J. & Martins, R. N. The role of type 2 diabetes in neurodegeneration. *Neurobiology of Disease* **84**, 22-38, doi:10.1016/j.nbd.2015.04.008 (2015).
- 111 Mosconi, L. *et al.* Increased Alzheimer's risk during the menopause transition: A 3-year longitudinal brain imaging study. *PLoS One* **13**, e0207885, doi:10.1371/journal.pone.0207885 (2018).
- 112 Brinton, R. D., Yao, J., Yin, F., Mack, W. J. & Cadenas, E. Perimenopause as a neurological transition state. *Nature Reviews Endocrinology* **11**, 393-405, doi:10.1038/nrendo.2015.82 (2015).
- 113 Nuriel, T. *et al.* Neuronal hyperactivity due to loss of inhibitory tone in apoe4 mice lacking alzheimer's disease-like pathology. *Nature Communications* **8**, doi:10.1038/s41467-017-01444-0 (2017).
- 114 Wu, L., Zhang, X. & Zhao, L. Human apoe isoforms differentially modulate brain glucose and ketone body metabolism: Implications for alzheimer's disease risk reduction and early intervention. *The Journal of Neuroscience* **38**, 6665-6681, doi:10.1523/jneurosci.2262-17.2018 (2018).
- 115 Zhao, N. *et al.* Apolipoprotein e4 impairs neuronal insulin signaling by trapping insulin receptor in the endosomes. *Neuron* **96**, 115-129 e115, doi:10.1016/j.neuron.2017.09.003 (2017).
- 116 Johnson, L. A. *et al.* Apolipoprotein e4 mediates insulin resistance-associated cerebrovascular dysfunction and the post-prandial response. *Journal of Cerebral Blood Flow & Metabolism* **39**, 770-781, doi:10.1177/0271678x17746186 (2017).
- 117 Nakamura, T., Watanabe, A., Fujino, T., Hosono, T. & Michikawa, M. Apolipoprotein e4 (1-272) fragment is associated with mitochondrial proteins and affects mitochondrial function in neuronal cells. *Mol Neurodegener* **4**, 35, doi:10.1186/1750-1326-4-35 (2009).

- 118 Orr, A. L. *et al.* Neuronal apolipoprotein e4 expression results in proteome-wide alterations and compromises bioenergetic capacity by disrupting mitochondrial function. *Journal of Alzheimer's Disease* **68**, 991-1011, doi:10.3233/jad-181184 (2019).
- 119 Henderson, S. T. *et al.* Study of the ketogenic agent ac-1202 in mild to moderate Alzheimer's disease: A randomized, double-blind, placebo-controlled, multicenter trial. *Nutrition & Metabolism* **6**, 31, doi:10.1186/1743-7075-6-31 (2009).
- 120 Xu, Q. *et al.* Medium-chain triglycerides improved cognition and lipid metabolomics in mild to moderate Alzheimer's disease patients with apoe4(-/-): A double-blind, randomized, placebo-controlled crossover trial. *Clin Nutr*, doi:10.1016/j.clnu.2019.10.017 (2019).
- 121 Larsen, S. B., Hanss, Z. & Krüger, R. The genetic architecture of mitochondrial dysfunction in Parkinson's disease. *Cell and Tissue Research* **373**, 21-37, doi:10.1007/s00441-017-2768-8 (2018).
- 122 Funk, N. *et al.* The Parkinson's disease-linked leucine-rich repeat kinase 2 (Irrk2) is required for insulin-stimulated translocation of glut4. *Sci Rep* **9**, 4515, doi:10.1038/s41598-019-40808-y (2019).
- 123 Joardar, A., Manzo, E. & Zarnescu, D. C. Metabolic dysregulation in amyotrophic lateral sclerosis: Challenges and opportunities. *Current Genetic Medicine Reports* **5**, 108-114, doi:10.1007/s40142-017-0123-8 (2017).
- 124 Diehl-Schmid, J. *et al.* FDG-pet underscores the key role of the thalamus in frontotemporal lobar degeneration caused by c9orf72 mutations. *Translational Psychiatry* **9**, doi:10.1038/s41398-019-0381-1 (2019).
- 125 Koppel, S. J. & Swerdlow, R. H. Neuroketotherapeutics: A modern review of a centuryold therapy. *Neurochemistry International* **117**, 114-125, doi:10.1016/j.neuint.2017.05.019 (2018).
- 126 Ngandu, T. *et al.* A 2 year multidomain intervention of diet, exercise, cognitive training, and vascular risk monitoring versus control to prevent cognitive decline in at-risk elderly people (finger): A randomised controlled trial. *Lancet* **385**, 2255-2263, doi:10.1016/s0140-6736(15)60461-5 (2015).
- 127 Gaitan, J. M. *et al.* Two weeks of interval training enhances fat oxidation during exercise in obese adults with prediabetes. *J Sports Sci Med* **18**, 636-644 (2019).
- 128 Zhang, L. *et al.* Modulation of mitochondrial complex i activity averts cognitive decline in multiple animal models of familial Alzheimer's disease. *EBioMedicine* **2**, 294-305, doi:10.1016/j.ebiom.2015.03.009 (2015).
- 129 Flannery, P. J. & Trushina, E. Mitochondrial dynamics and transport in Alzheimer's disease. *Molecular and Cellular Neuroscience* **98**, 109-120, doi:10.1016/j.mcn.2019.06.009 (2019).
- Baumgart, M. *et al.* Longitudinal rna-seq analysis of vertebrate aging identifies mitochondrial complex i as a small-molecule-sensitive modifier of lifespan. *Cell Syst* 2, 122-132, doi:10.1016/j.cels.2016.01.014 (2016).
- 131 Raule, N. *et al.* The co-occurrence of mtdna mutations on different oxidative phosphorylation subunits, not detected by haplogroup analysis, affects human longevity and is population specific. *Aging Cell* **13**, 401-407, doi:10.1111/acel.12186 (2014).
- 132 Herzig, S. & Shaw, R. J. Ampk: Guardian of metabolism and mitochondrial homeostasis. *Nature Reviews Molecular Cell Biology* **19**, 121-135, doi:10.1038/nrm.2017.95 (2017).
- 133 Burte, F., Carelli, V., Chinnery, P. F. & Yu-Wai-Man, P. Disturbed mitochondrial dynamics and neurodegenerative disorders. *Nature reviews Neurology* **11**, 11-24, doi:10.1038/nrneurol.2014.228 (2015).

- 134 Manczak, M., Kandimalla, R., Yin, X. & Reddy, P. H. Mitochondrial division inhibitor 1 reduces dynamin-related protein 1 and mitochondrial fission activity. *Hum Mol Genet*, doi:10.1093/hmg/ddy335 (2018).
- Bido, S., Soria, F. N., Fan, R. Z., Bezard, E. & Tieu, K. Mitochondrial division inhibitor-1 is neuroprotective in the a53t-alpha-synuclein rat model of Parkinson's disease. *Sci Rep* 7, 7495, doi:10.1038/s41598-017-07181-0 (2017).
- 136 Baek, S. H. *et al.* Inhibition of drp1 ameliorates synaptic depression, abeta deposition, and cognitive impairment in an Alzheimer's disease model. *J Neurosci* **37**, 5099-5110, doi:10.1523/jneurosci.2385-16.2017 (2017).
- 137 Wilkins, H. M. *et al.* A mitochondrial biomarker-based study of s-equol in Alzheimer's disease subjects: Results of a single-arm, pilot trial. *J Alzheimers Dis* **59**, 291-300, doi:10.3233/jad-170077 (2017).
- 138 Wu, B. *et al.* 2,4 dnp improves motor function, preserves medium spiny neuronal identity, and reduces oxidative stress in a mouse model of Huntington's disease. *Experimental Neurology* **293**, 83-90, doi:10.1016/j.expneurol.2017.03.020 (2017).
- 139 Dickey, A. S. *et al.* Ppardelta activation by bexarotene promotes neuroprotection by restoring bioenergetic and quality control homeostasis. *Sci Transl Med* **9**, doi:10.1126/scitranslmed.aal2332 (2017).
- 140 Xin, L. *et al.* Nutritional ketosis increases NAD+/NADH ratio in healthy human brain: An in vivo study by 31p-mrs. *Frontiers in Nutrition* **5**, doi:10.3389/fnut.2018.00062 (2018).
- 141 Dellinger, R. W. *et al.* Repeat dose NRPT (nicotinamide riboside and pterostilbene) increases NAD+ levels in humans safely and sustainably: A randomized, double-blind, placebo-controlled study. *npj Aging and Mechanisms of Disease* **3**, doi:10.1038/s41514-017-0016-9 (2017).
- 142 Hou, Y. *et al.* NAD+ supplementation normalizes key alzheimer's features and DNA damage responses in a new ad mouse model with introduced DNA repair deficiency. *Proceedings of the National Academy of Sciences* **115**, E1876-E1885, doi:10.1073/pnas.1718819115 (2018).
- 143 Schöndorf, D. C. *et al.* The NAD+ precursor nicotinamide riboside rescues mitochondrial defects and neuronal loss in ipsc and fly models of parkinson's disease. *Cell Reports* **23**, 2976-2988, doi:10.1016/j.celrep.2018.05.009 (2018).
- 144 Wilkins, H. M. *et al.* Oxaloacetate enhances neuronal cell bioenergetic fluxes and infrastructure. *Journal of Neurochemistry* **137**, 76-87, doi:10.1111/jnc.13545 (2016).
- 145 Cai, R. *et al.* Enhancing glycolysis attenuates Parkinson's disease progression in models and clinical databases. *J Clin Invest* **129**, 4539-4549, doi:10.1172/jci129987 (2019).
- 146 Zilberter, M. *et al.* Dietary energy substrates reverse early neuronal hyperactivity in a mouse model of Alzheimer's disease. *Journal of Neurochemistry* **125**, 157-171, doi:10.1111/jnc.12127 (2013).
- 147 Theurey, P. *et al.* Systems biology identifies preserved integrity but impaired metabolism of mitochondria due to a glycolytic defect in Alzheimer's disease neurons. *Aging Cell* **18**, e12924, doi:10.1111/acel.12924 (2019).
- 148 Roy, M. *et al.* Rapid adaptation of rat brain and liver metabolism to a ketogenic diet: An integrated study using1H- and 13C-nmr spectroscopy. *Journal of Cerebral Blood Flow & Metabolism* **35**, 1154-1162, doi:10.1038/jcbfm.2015.29 (2015).
- 149 St-Pierre, V. *et al.* Plasma ketone and medium chain fatty acid response in humans consuming different medium chain triglycerides during a metabolic study day. *Front Nutr* **6**, 46, doi:10.3389/fnut.2019.00046 (2019).

- 150 Tan, K. N., Carrasco-Pozo, C., McDonald, T. S., Puchowicz, M. & Borges, K. Tridecanoin is anticonvulsant, antioxidant, and improves mitochondrial function. *J Cereb Blood Flow Metab* **37**, 2035-2048, doi:10.1177/0271678x16659498 (2017).
- 151 Schwarzkopf, T. M., Koch, K. & Klein, J. Reduced severity of ischemic stroke and improvement of mitochondrial function after dietary treatment with the anaplerotic substance triheptanoin. *Neuroscience* **300**, 201-209, doi:10.1016/j.neuroscience.2015.05.014 (2015).
- 152 Mochel, F. Triheptanoin for the treatment of brain energy deficit: A 14-year experience. *Journal of Neuroscience Research* **95**, 2236-2243, doi:10.1002/jnr.24111 (2017).
- 153 Marin-Valencia, I., Good, L. B., Ma, Q., Malloy, C. R. & Pascual, J. M. Heptanoate as a neural fuel: Energetic and neurotransmitter precursors in normal and glucose transporter i-deficient (G1D) brain. *Journal of Cerebral Blood Flow & Metabolism* 33, 175-182, doi:10.1038/jcbfm.2012.151 (2012).
- 154 Fortier, M. *et al.* A ketogenic drink improves brain energy and some measures of cognition in mild cognitive impairment. *Alzheimer's & Dementia* **15**, 625-634, doi:10.1016/j.jalz.2018.12.017 (2019).
- 155 Krikorian, R. *et al.* Dietary ketosis enhances memory in mild cognitive impairment. *Neurobiology of Aging* **33**, 425.e419-425.e427, doi:10.1016/j.neurobiolaging.2010.10.006 (2012).
- 156 Neth, B. J. *et al.* Modified ketogenic diet is associated with improved cerebrospinal fluid biomarker profile, cerebral perfusion, and cerebral ketone body uptake in older adults at risk for alzheimer's disease: A pilot study. *Neurobiol Aging* **86**, 54-63, doi:10.1016/j.neurobiolaging.2019.09.015 (2020).
- 157 Taylor, M. K., Sullivan, D. K., Mahnken, J. D., Burns, J. M. & Swerdlow, R. H. Feasibility and efficacy data from a ketogenic diet intervention inAalzheimer's disease. *Alzheimer's* & *Dementia: Translational Research & Clinical Interventions* **4**, 28-36, doi:10.1016/j.trci.2017.11.002 (2017).
- 158 Ota, M. *et al.* Effects of a medium-chain triglyceride-based ketogenic formula on cognitive function in patients with mild-to-moderate Alzheimer's disease. *Neuroscience Letters* **690**, 232-236, doi:10.1016/j.neulet.2018.10.048 (2019).
- 159 Avgerinos, K. I., Egan, J. M., Mattson, M. P. & Kapogiannis, D. Medium chain triglycerides induce mild ketosis and may improve cognition in Alzheimer's disease. A systematic review and meta-analysis of human studies. *Ageing Res Rev* 58, 101001, doi:10.1016/j.arr.2019.101001 (2020).
- 160 Phillips, M. C. L., Murtagh, D. K. J., Gilbertson, L. J., Asztely, F. J. S. & Lynch, C. D. P. Lowfat versus ketogenic diet in Parkinson's disease: A pilot randomized controlled trial. *Mov Disord* **33**, 1306-1314, doi:10.1002/mds.27390 (2018).
- 161 Krikorian, R. *et al.* Nutritional ketosis for mild cognitive impairment in Parkinson's disease: A controlled pilot trial. *Clinical Parkinsonism & Related Disorders* **1**, 41-47, doi:https://doi.org/10.1016/j.prdoa.2019.07.006 (2019).
- 162 Brandt, J. *et al.* Preliminary report on the feasibility and efficacy of the modified atkins diet for treatment of mild cognitive impairment and early Alzheimer's disease. *Journal of Alzheimer's Disease* **68**, 969-981, doi:10.3233/jad-180995 (2019).
- 163 Shaafi, S. *et al.* The efficacy of the ketogenic diet on motor functions in Parkinson's disease: A rat model. *Iran J Neurol* **15**, 63-69 (2016).
- 164 Ari, C. *et al.* Metabolic therapy with deanna protocol supplementation delays disease progression and extends survival in amyotrophic lateral sclerosis (als) mouse model. *PLoS ONE* **9**, e103526, doi:10.1371/journal.pone.0103526 (2014).

- 165 Ruskin, D. N. *et al.* A ketogenic diet delays weight loss and does not impair working memory or motor function in the r6/2 1j mouse model of Huntington's disease. *Physiology & Behavior* **103**, 501-507, doi:10.1016/j.physbeh.2011.04.001 (2011).
- 166 Ma, D. *et al.* Ketogenic diet enhances neurovascular function with altered gut microbiome in young healthy mice. *Scientific Reports* **8**, doi:10.1038/s41598-018-25190-5 (2018).
- 167 Reger, M. A. *et al.* Effects of β -hydroxybutyrate on cognition in memory-impaired adults. *Neurobiology of Aging* **25**, 311-314, doi:10.1016/s0197-4580(03)00087-3 (2004).
- 168 Page, K. A. *et al.* Medium-chain fatty acids improve cognitive function in intensively treated type 1 diabetic patients and support in vitro synaptic transmission during acute hypoglycemia. *Diabetes* **58**, 1237-1244 (2009).
- 169 Wakade, C., Chong, R., Bradley, E., Thomas, B. & Morgan, J. Upregulation of gpr109a in Parkinson's disease. *PLoS ONE* **9**, e109818, doi:10.1371/journal.pone.0109818 (2014).
- 170 Fu, S.-P. *et al.* Anti-inflammatory effects of bhba in both in vivo and in vitro parkinson's disease models are mediated by gpr109a-dependent mechanisms. *Journal of Neuroinflammation* **12**, 9, doi:10.1186/s12974-014-0230-3 (2015).
- 171 Masino, S. A. & Rho, J. M. Metabolism and epilepsy: Ketogenic diets as a homeostatic link. *Brain Res* **1703**, 26-30, doi:10.1016/j.brainres.2018.05.049 (2019).
- 172 Kashiwaya, Y. *et al.* A ketone ester diet exhibits anxiolytic and cognition-sparing properties, and lessens amyloid and tau pathologies in a mouse model of Alzheimer's disease. *Neurobiology of Aging* 34, 1530-1539, doi:10.1016/j.neurobiolaging.2012.11.023 (2013).
- 173 Cucuzzella, M., Hite, A., Patterson, K., Saslow, L. & Heath, R. A clinician's guide to inpatient low carbohydrate diets for remission of type 2 diabetes :Toward a standard of care protocol. *Diabetes Manag.* **9**, 7-19 (2019).
- 174 Ritze, Y. *et al.* Metabolic and cognitive outcomes of subchronic once-daily intranasal insulin administration in healthy men. *Frontiers in Endocrinology* **9**, doi:10.3389/fendo.2018.00663 (2018).
- 175 Rotermund, C., Machetanz, G. & Fitzgerald, J. C. The therapeutic potential of metformin in neurodegenerative diseases. *Frontiers in Endocrinology* 9, doi:10.3389/fendo.2018.00400 (2018).
- 176 Ou, Z. *et al.* Metformin treatment prevents amyloid plaque deposition and memory impairment in app/ps1 mice. *Brain, Behavior, and Immunity* **69**, 351-363, doi:10.1016/j.bbi.2017.12.009 (2018).
- 177 Arnoux, I. *et al.* Metformin reverses early cortical network dysfunction and behavior changes in Huntington's disease. *Elife* **7**, doi:10.7554/eLife.38744 (2018).
- 178 Campbell, J. M. *et al.* Metformin use associated with reduced risk of dementia in patients with diabetes: A systematic review and meta-analysis. *Journal of Alzheimer's Disease* **65**, 1225-1236, doi:10.3233/jad-180263 (2018).
- 179 Cameron, A. R. *et al.* Metformin selectively targets redox control of complex I energy transduction. *Redox Biol* **14**, 187-197, doi:10.1016/j.redox.2017.08.018 (2018).
- 180 Rena, G., Hardie, D. G. & Pearson, E. R. The mechanisms of action of metformin. *Diabetologia* **60**, 1577-1585, doi:10.1007/s00125-017-4342-z (2017).
- 181 Coll, A. P. *et al.* Gdf15 mediates the effects of metformin on body weight and energy balance. *Nature* **578**, 444-448, doi:10.1038/s41586-019-1911-y (2020).
- 182 Moore, E. M. *et al.* Increased risk of cognitive impairment in patients with diabetes is associated with metformin. *Diabetes Care* **36**, 2981-2987, doi:10.2337/dc13-0229 (2013).

- 183 Koenig, A. M. *et al.* Effects of the insulin sensitizer metformin in Alzheimer disease: Pilot data from a randomized placebo-controlled crossover study. *Alzheimer Dis Assoc Disord* **31**, 107-113, doi:10.1097/WAD.00000000000202 (2017).
- 184 Luchsinger, J. A. *et al.* Metformin in amnestic mild cognitive impairment: Results of a pilot randomized placebo controlled clinical trial. *J Alzheimers Dis* **51**, 501-514, doi:10.3233/JAD-150493 (2016).
- 185 Lin, Y. *et al.* Corrigendum: Evaluation of metformin on cognitive improvement in patients with non-dementia vascular cognitive impairment and abnormal glucose metabolism. *Front Aging Neurosci* **10**, 322, doi:10.3389/fnagi.2018.00322 (2018).
- 186 Arnold, S. E. *et al.* Brain insulin resistance in type 2 diabetes and Alzheimer disease: Concepts and conundrums. *Nature Reviews Neurology* **14**, 168-181, doi:10.1038/nrneurol.2017.185 (2018).
- 187 Wiviott, S. D., Raz, I. & Sabatine, M. S. Dapagliflozin and cardiovascular outcomes in type 2 diabetes. Reply. *N Engl J Med* **380**, 1881-1882, doi:10.1056/NEJMc1902837 (2019).
- 188 Ferrannini, E. *et al.* Shift to fatty substrate utilization in response to sodium–glucose cotransporter 2 inhibition in subjects without diabetes and patients with type 2 diabetes. *Diabetes* **65**, 1190-1195, doi:10.2337/db15-1356 (2016).
- 189 Holst, J. J. & Madsbad, S. Semaglutide seems to be more effective the other GLP-1RAS. Annals of Translational Medicine, 505-505, doi:10.21037/atm.2017.11.10 (2017).
- 190 Goldenberg, R. M. & Steen, O. Semaglutide: Review and place in therapy for adults with type 2 diabetes. *Canadian Journal of Diabetes* **43**, 136-145, doi:10.1016/j.jcjd.2018.05.008 (2019).
- 191 Batista, A. F., Bodart-Santos, V., De Felice, F. G. & Ferreira, S. T. Neuroprotective actions of glucagon-like peptide-1 (GLP-1) analogues in Alzheimer's and Parkinson's diseases. *CNS Drugs* **33**, 209-223, doi:10.1007/s40263-018-0593-6 (2018).
- 192 Yildirim Simsir, I., Soyaltin, U. E. & Cetinkalp, S. Glucagon like peptide-1 (GLP-1) likes Alzheimer's disease. *Diabetes & Metabolic Syndrome: Clinical Research & Reviews* 12, 469-475, doi:10.1016/j.dsx.2018.03.002 (2018).
- Sayed, N. H. *et al.* Vildagliptin attenuates huntington's disease through activation of glp-1 receptor/pi3k/akt/bdnf pathway in 3-nitropropionic acid rat model. *Neurotherapeutics* 17, 252-268, doi:10.1007/s13311-019-00805-5 (2020).
- 194 Gejl, M. *et al.* In alzheimer's disease, 6-month treatment with GLP-1 analog prevents decline of brain glucose metabolism: Randomized, placebo-controlled, double-blind clinical trial. *Frontiers in Aging Neuroscience* **8**, doi:10.3389/fnagi.2016.00108 (2016).
- 195 Athauda, D. *et al.* Post hoc analysis of the exenatide-pd trial-factors that predict response. *European Journal of Neuroscience* **49**, 410-421, doi:10.1111/ejn.14096 (2018).
- 196 Chalichem, N. S. S., Gonugunta, C., Krishnamurthy, P. T. & Duraiswamy, B. DPP4 inhibitors can be a drug of choice for type 3 diabetes: A mini review. *Am J Alzheimers Dis Other Demen* **32**, 444-451, doi:10.1177/1533317517722005 (2017).
- 197 Isik, A. T., Soysal, P., Yay, A. & Usarel, C. The effects of sitagliptin, a dpp-4 inhibitor, on cognitive functions in elderly diabetic patients with or without Alzheimer's disease. *Diabetes Research and Clinical Practice* **123**, 192-198, doi:10.1016/j.diabres.2016.12.010 (2017).
- 198 Jalewa, J., Sharma, M. K. & Hölscher, C. Novel incretin analogues improve autophagy and protect from mitochondrial stress induced by rotenone in sh-sy5y cells. *Journal of Neurochemistry* **139**, 55-67, doi:10.1111/jnc.13736 (2016).

- 199 Verma, M. K., Goel, R., Nandakumar, K. & Nemmani, K. V. S. Effect of d-Ala 2 GIP, a stable GIP receptor agonist on MPTP-induced neuronal impairments in mice. *European Journal of Pharmacology* **804**, 38-45, doi:10.1016/j.ejphar.2017.03.059 (2017).
- 200 Pathak, N. M. *et al.* Novel dual incretin agonist peptide with antidiabetic and neuroprotective potential. *Biochemical Pharmacology* **155**, 264-274, doi:10.1016/j.bcp.2018.07.021 (2018).
- 201 Hölscher, C. Novel dual GLP-1/GIP receptor agonists show neuroprotective effects in Alzheimer's and Parkinson's disease models. *Neuropharmacology* **136**, 251-259, doi:10.1016/j.neuropharm.2018.01.040 (2018).
- Tai, J., Liu, W., Li, Y., Li, L. & Hölscher, C. Neuroprotective effects of a triple GLP 1/GIP/glucagon receptor agonist in the app/ps1 transgenic mouse model of Alzheimer's disease. *Brain Research* 1678, 64-74, doi:10.1016/j.brainres.2017.10.012 (2018).
- 203 Rudenko, O. *et al.* Ghrelin mediated improvements in the metabolic phenotype in the r6/2 mouse model of Huntington's disease. *Journal of Neuroendocrinology* **31**, doi:10.1111/jne.12699 (2019).
- 204 Jeong, Y.-o. *et al.* Mk-0677, a ghrelin agonist, alleviates amyloid beta-related pathology in 5xfad mice, an animal model of Alzheimer's disease. *International Journal of Molecular Sciences* **19**, 1800, doi:10.3390/ijms19061800 (2018).
- Morgan, A. H., Rees, D. J., Andrews, Z. B. & Davies, J. S. Ghrelin mediated neuroprotection - a possible therapy for Parkinson's disease? *Neuropharmacology* 136, 317-326, doi:10.1016/j.neuropharm.2017.12.027 (2018).
- 206 Bayliss, J. A. *et al.* Ghrelin-ampk signaling mediates the neuroprotective effects of calorie restriction in Parkinson's disease. *Journal of Neuroscience* **36**, 3049-3063, doi:10.1523/jneurosci.4373-15.2016 (2016).
- 207 A randomized trial of relamorelin for constipation in Parkinson's disease (move-pd): Trial results and lessons learned. *Parkinsonism & Related Disorders* **37**, 101-105, doi:10.1016/j.parkreldis.2017.02.003 (2017).
- 208 Malekizadeh, Y. *et al.* A leptin fragment mirrors the cognitive enhancing and neuroprotective actions of leptin. *Cerebral Cortex* **27**, 4769-4782, doi:10.1093/cercor/bhw272 (2016).
- 209 Fernandez-Martos, C. M., Atkinson, R. A. K., Chuah, M. I., King, A. E. & Vickers, J. C. Combination treatment with leptin and pioglitazone in a mouse model of Alzheimer's disease. *Alzheimer's & Dementia: Translational Research & Clinical Interventions* **3**, 92-106, doi:10.1016/j.trci.2016.11.002 (2016).
- 210 Lim, M. A. *et al.* Genetically altering organismal metabolism by leptin-deficiency benefits a mouse model of amyotrophic lateral sclerosis. *Human Molecular Genetics* 23, 4995-5008, doi:10.1093/hmg/ddu214 (2014).
- 211 Yoon, G., Shah, S. A., Ali, T. & Kim, M. O. The adiponectin homolog osmotin enhances neurite outgrowth and synaptic complexity via ADIPOR1/NGR1 signaling in Alzheimer's disease. *Molecular Neurobiology* 55, 6673-6686, doi:10.1007/s12035-017-0847-1 (2018).
- 212 Soudy, R. *et al.* Short amylin receptor antagonist peptides improve memory deficits in alzheimer's disease mouse model. *Scientific Reports* **9**, doi:10.1038/s41598-019-47255-9 (2019).
- 213 Levin, B. E. & Lutz, T. A. Amylin and leptin: Co-regulators of energy homeostasis and neuronal development. *Trends in Endocrinology & Metabolism* **28**, 153-164, doi:10.1016/j.tem.2016.11.004 (2017).

- 214 Patrick, S. *et al.* Neuroprotective effects of the amylin analog, pramlintide, on Alzheimer's disease are associated with oxidative stress regulation mechanisms. *Journal of Alzheimer's Disease* **69**, 157-168, doi:10.3233/jad-180421 (2019).
- 215 Wang, E. *et al.* Amylin treatment reduces neuroinflammation and ameliorates abnormal patterns of gene expression in the cerebral cortex of an Alzheimer's disease mouse model. *Journal of Alzheimer's Disease* **56**, 47-61, doi:10.3233/jad-160677 (2017).
- 216 Pinho, T. S., Correia, S. C., Perry, G., Ambrósio, A. F. & Moreira, P. I. Diminished O-GlcNAcylation in Alzheimer's disease is strongly correlated with mitochondrial anomalies. *Biochimica et Biophysica Acta (BBA) - Molecular Basis of Disease* **1865**, 2048-2059, doi:10.1016/j.bbadis.2018.10.037 (2019).
- 217 Levine, P. M. *et al.* α-synuclein O-GlcNAcylation alters aggregation and toxicity, revealing certain residues as potential inhibitors of Parkinson's disease. *Proceedings of the National Academy of Sciences* **116**, 1511-1519, doi:10.1073/pnas.1808845116 (2019).
- 218 Yuzwa, S. A. & Vocadlo, D. J. O-GlcNAc and neurodegeneration: Biochemical mechanisms and potential roles in alzheimer's disease and beyond. *Chem. Soc. Rev.* 43, 6839-6858, doi:10.1039/c4cs00038b (2014).
- 219 Haas, R. *et al.* Intermediates of metabolism: From bystanders to signalling molecules. *Trends in Biochemical Sciences* **41**, 460-471, doi:10.1016/j.tibs.2016.02.003 (2016).
- 220 Cobos, S. N., Bennett, S. A. & Torrente, M. P. The impact of histone post-translational modifications in neurodegenerative diseases. *Biochimica et Biophysica Acta (BBA) Molecular Basis of Disease* **1865**, 1982-1991, doi:10.1016/j.bbadis.2018.10.019 (2019).
- 221 Narayan, P. & Dragunow, M. in *Advances in Experimental Medicine and Biology* 321-336 (Springer International Publishing, 2017).
- 222 El Hayek, L. *et al.* Lactate mediates the effects of exercise on learning and memory through SIRT1-dependent activation of hippocampal brain-derived neurotrophic factor (BDNF). *The Journal of Neuroscience*, 1661-1618, doi:10.1523/jneurosci.1661-18.2019 (2019).
- 223 Karnib, N. *et al.* Lactate is an antidepressant that mediates resilience to stress by modulating the hippocampal levels and activity of histone deacetylases. *Neuropsychopharmacology* 44, 1152-1162, doi:10.1038/s41386-019-0313-z (2019).
- Xie, Z. *et al.* Metabolic regulation of gene expression by histone lysine β-hydroxybutyrylation. *Molecular Cell* 62, 194-206, doi:10.1016/j.molcel.2016.03.036 (2016).
- 225 Poletti, V. & Biffi, A. Gene-based approaches to inherited neurometabolic diseases. *Hum Gene Ther* **30**, 1222-1235, doi:10.1089/hum.2019.190 (2019).
- 226 Mercuri, E. *et al.* Nusinersen versus sham control in later-onset spinal muscular atrophy. *N Engl J Med* **378**, 625-635, doi:10.1056/NEJMoa1710504 (2018).
- 227 Millan, M. J. Linking deregulation of non-coding RNA to the core pathophysiology of alzheimer's disease: An integrative review. *Progress in Neurobiology* **156**, 1-68, doi:10.1016/j.pneurobio.2017.03.004 (2017).
- Fumagalli, M., Lombardi, M., Gressens, P. & Verderio, C. How to reprogram microglia toward beneficial functions. *Glia* **66**, 2531-2549, doi:10.1002/glia.23484 (2018).
- 229 Maniati, M. S., Maniati, M., Yousefi, T., Ahmadi Ahangar, A. & Tehrani, S. S. New insights into the role of micrornas and long noncoding RNAs in most common neurodegenerative diseases. *Journal of Cellular Biochemistry* **120**, 8908-8918, doi:10.1002/jcb.28361 (2019).

- 230 Wild, E. J. & Tabrizi, S. J. One decade ago, one decade ahead in huntington's disease. *Mov Disord* **34**, 1434-1439, doi:10.1002/mds.27849 (2019).
- 231 Zeitler, B. *et al.* Allele-selective transcriptional repression of mutant htt for the treatment of huntington's disease. *Nat Med* **25**, 1131-1142, doi:10.1038/s41591-019-0478-3 (2019).
- 232 Cappella, M., Ciotti, C., Cohen-Tannoudji, M. & Biferi, M. G. Gene therapy for als-a perspective. *Int J Mol Sci* **20**, doi:10.3390/ijms20184388 (2019).
- 233 Savitt, D. & Jankovic, J. Targeting alpha-synuclein in parkinson's disease: Progress towards the development of disease-modifying therapeutics. *Drugs* **79**, 797-810, doi:10.1007/s40265-019-01104-1 (2019).
- 234 Deverman, B. E., Ravina, B. M., Bankiewicz, K. S., Paul, S. M. & Sah, D. W. Y. Gene therapy for neurological disorders: Progress and prospects. *Nature Reviews Drug Discovery* **17**, 641-659, doi:10.1038/nrd.2018.110 (2018).
- 235 Al-Zaidy, S. A. *et al.* Avxs-101 (onasemnogene abeparvovec) for sma1: Comparative study with a prospective natural history cohort. *J Neuromuscul Dis* **6**, 307-317, doi:10.3233/jnd-190403 (2019).
- 236 Nakamura, S. *et al.* Gene therapy for glut1 -deficient mouse using an adeno-associated virus vector with the human intrinsic glut1 promoter. *The Journal of Gene Medicine* **20**, e3013, doi:10.1002/jgm.3013 (2018).
- 237 Choong, C.-J. & Mochizuki, H. Gene therapy targeting mitochondrial pathway in parkinson's disease. *Journal of Neural Transmission* **124**, 193-207, doi:10.1007/s00702-016-1616-4 (2016).
- 238 Rohn, T. T., Kim, N., Isho, N. F. & Mack, J. M. The potential of crispr/cas9 gene editing as a treatment strategy for alzheimer's disease. J Alzheimers Dis Parkinsonism 8, doi:10.4172/2161-0460.1000439 (2018).
- 239 Safieh, M., Korczyn, A. D. & Michaelson, D. M. Apoe4: An emerging therapeutic target for alzheimer's disease. *BMC Med* **17**, 64, doi:10.1186/s12916-019-1299-4 (2019).
- 240 Uddin, M. S. *et al.* Apoe and alzheimer's disease: Evidence mounts that targeting apoe4 may combat alzheimer's pathogenesis. *Mol Neurobiol* **56**, 2450-2465, doi:10.1007/s12035-018-1237-z (2019).
- 241 Creus-Muncunill, J. *et al.* Increased translation as a novel pathogenic mechanism in huntington's disease. *Brain* **142**, 3158-3175, doi:10.1093/brain/awz230 (2019).
- 242 Takhashi, D. *et al.* AUTACS: Cargo-specific degraders using selective autophagy. *Mol Cell* 76, 797-810 (2019).
- 243 de la Torre, J. C. Treating cognitive impairment with transcranial low level laser therapy. *Journal of Photochemistry and Photobiology B: Biology* **168**, 149-155, doi:10.1016/j.jphotobiol.2017.02.008 (2017).
- 244 Hamblin, M. R. Shining light on the head: Photobiomodulation for brain disorders. *BBA Clinical* **6**, 113-124, doi:10.1016/j.bbacli.2016.09.002 (2016).
- 245 Salehpour, F. *et al.* Transcranial low-level laser therapy improves brain mitochondrial function and cognitive impairment in d-galactose–induced aging mice. *Neurobiology of Aging* **58**, 140-150, doi:10.1016/j.neurobiolaging.2017.06.025 (2017).
- 246 Berman, M. H. *et al.* Photobiomodulation with near infrared light helmet in a pilot, placebo controlled clinical trial in dementia patients testing memory and cognition. *Journal of Neurology and Neuroscience* **08**, doi:10.21767/2171-6625.1000176 (2017).
- 247 Martorell, A. J. *et al.* Multi-sensory gamma stimulation ameliorates alzheimer'sassociated pathology and improves cognition. *Cell* **177**, 256-271.e222, doi:10.1016/j.cell.2019.02.014 (2019).

- 248 Clemmensen, C. *et al.* Emerging hormonal-based combination pharmacotherapies for the treatment of metabolic diseases. *Nature Reviews Endocrinology* **15**, 90-104, doi:10.1038/s41574-018-0118-x (2018).
- 249 Cummings, J., Ritter, A. & Rothenberg, K. Advances in management of neuropsychiatric syndromes in neurodegenerative diseases. *Current Psychiatry Reports* **21**, doi:10.1007/s11920-019-1058-4 (2019).
- 250 de Freitas Silva, M., Dias, K. S. T., Gontijo, V. S., Ortiz, C. J. C. & Viegas, C. Multi-target directed drugs as a modern approach for drug design towards alzheimer's disease: An update. *Current Medicinal Chemistry* **25**, 3491-3525, doi:10.2174/0929867325666180111101843 (2018).
- 251 Kivipelto, M., Mangialasche, F. & Ngandu, T. Lifestyle interventions to prevent cognitive impairment, dementia and alzheimer disease. *Nature Reviews Neurology* **14**, 653-666, doi:10.1038/s41582-018-0070-3 (2018).
- 252 McDonald, T., Puchowicz, M. & Borges, K. Impairments in oxidative glucose metabolism in epilepsy and metabolic treatments thereof. *Front Cell Neurosci* **12**, 274, doi:10.3389/fncel.2018.00274 (2018).
- E, L., Lu, J., Selfridge, J. E., Burns, J. M. & Swerdlow, R. H. Lactate administration reproduces specific brain and liver exercise-related changes. *J Neurochem* 127, 91-100, doi:10.1111/jnc.12394 (2013).
- 254 Morland, C. *et al.* Exercise induces cerebral vegf and angiogenesis via the lactate receptor hcar1. *Nat Commun* **8**, 15557, doi:10.1038/ncomms15557 (2017).
- 255 Insel, P. S. *et al.* Time to amyloid positivity and preclinical changes in brain metabolism, atrophy, and cognition: Evidence for emerging amyloid pathology in alzheimer's disease. *Frontiers in Neuroscience* **11**, doi:10.3389/fnins.2017.00281 (2017).
- 256 Liyanage, S. I., Santos, C. & Weaver, D. F. The hidden variables problem in alzheimer's disease clinical trial design. *Alzheimer's & Dementia: Translational Research & Clinical Interventions* 4, 628-635, doi:10.1016/j.trci.2018.09.003 (2018).
- 257 McManus, M. J., Murphy, M. P. & Franklin, J. L. The mitochondria-targeted antioxidant mitoq prevents loss of spatial memory retention and early neuropathology in a transgenic mouse model of alzheimer's disease. *J Neurosci* **31**, 15703-15715, doi:10.1523/jneurosci.0552-11.2011 (2011).
- 258 Zhao, F.-l. *et al.* Ap39, a mitochondria-targeted hydrogen sulfide donor, supports cellular bioenergetics and protects against alzheimer's disease by preserving mitochondrial function in app/ps1 mice and neurons. *Oxidative Medicine and Cellular Longevity* 2016, 1-19, doi:10.1155/2016/8360738 (2016).
- 259 Sorrentino, V. *et al.* Enhancing mitochondrial proteostasis reduces amyloid-beta proteotoxicity. *Nature* **552**, 187-193, doi:10.1038/nature25143 (2017).
- 260 Zhao, Y. *et al.* Atad3a oligomerization causes neurodegeneration by coupling mitochondrial fragmentation and bioenergetics defects. *Nat Commun* **10**, 1371, doi:10.1038/s41467-019-09291-x (2019).
- 261 Miquel, E. *et al.* Neuroprotective effects of the mitochondria-targeted antioxidant mitoq in a model of inherited amyotrophic lateral sclerosis. *Free Radic Biol Med* **70**, 204-213, doi:10.1016/j.freeradbiomed.2014.02.019 (2014).
- 262 Stucki, D. M. *et al.* Mitochondrial impairments contribute to spinocerebellar ataxia type 1 progression and can be ameliorated by the mitochondria-targeted antioxidant mitoq. *Free Radic Biol Med* **97**, 427-440, doi:10.1016/j.freeradbiomed.2016.07.005 (2016).
- 263 Pawlosky, R. J. *et al.* Effects of a dietary ketone ester on hippocampal glycolytic and tricarboxylic acid cycle intermediates and amino acids in a 3xtgad mouse model of

Alzheimer's disease. *Journal of Neurochemistry* **141**, 195-207, doi:10.1111/jnc.13958 (2017).

- 264 Cheng, A. *et al.* Sirt3 haploinsufficiency aggravates loss of gabaergic interneurons and neuronal network hyperexcitability in an alzheimer's disease model. *J Neurosci* **40**, 694-709, doi:10.1523/jneurosci.1446-19.2019 (2020).
- Tieu, K. *et al.* D-β-hydroxybutyrate rescues mitochondrial respiration and mitigates features of parkinson disease. *Journal of Clinical Investigation* **112**, 892-901, doi:10.1172/jci200318797 (2003).
- 266 Zhao, W. *et al.* Caprylic triglyceride as a novel therapeutic approach to effectively improve the performance and attenuate the symptoms due to the motor neuron loss in als disease. *PLoS ONE* **7**, e49191, doi:10.1371/journal.pone.0049191 (2012).
- Gong, B. *et al.* Nicotinamide riboside restores cognition through an upregulation of proliferator-activated receptor-γ coactivator 1α regulated β-secretase 1 degradation and mitochondrial gene expression in alzheimer's mouse models. *Neurobiology of Aging* 34, 1581-1588, doi:10.1016/j.neurobiolaging.2012.12.005 (2013).
- 268 Costa, M. *et al.* N-acetylcysteine protects memory decline induced by streptozotocin in mice. *Chemico-Biological Interactions* **253**, 10-17, doi:10.1016/j.cbi.2016.04.026 (2016).
- 269 Tefera, T. W. *et al.* Triheptanoin protects motor neurons and delays the onset of motor symptoms in a mouse model of amyotrophic lateral sclerosis. *PLOS ONE* **11**, e0161816, doi:10.1371/journal.pone.0161816 (2016).
- 270 Croteau, E. *et al.* Ketogenic medium chain triglycerides increase brain energy metabolism in alzheimer's disease. *Journal of Alzheimer's Disease* **64**, 551-561, doi:10.3233/jad-180202 (2018).
- Adanyeguh, I. M. *et al.* Triheptanoin improves brain energy metabolism in patients with huntington disease. *Neurology* 84, 490-495, doi:10.1212/wnl.000000000001214 (2015).
- 272 Mookerjee, S. A., Gerencser, A. A., Nicholls, D. G. & Brand, M. D. Quantifying intracellular rates of glycolytic and oxidative atp production and consumption using extracellular flux measurements. *Journal of Biological Chemistry* **292**, 7189-7207, doi:10.1074/jbc.m116.774471 (2017).
- 273 Roe, C. R. & Brunengraber, H. Anaplerotic treatment of long-chain fat oxidation disorders with triheptanoin: Review of 15years experience. *Molecular Genetics and Metabolism* **116**, 260-268, doi:10.1016/j.ymgme.2015.10.005 (2015).
- Hyder, F. *et al.* Uniform distributions of glucose oxidation and oxygen extraction in gray matter of normal human brain: No evidence of regional differences of aerobic glycolysis. *Journal of Cerebral Blood Flow & Metabolism* 36, 903-916, doi:10.1177/0271678x15625349 (2016).
- Jha, M. K. & Morrison, B. M. Glia-neuron energy metabolism in health and diseases: New insights into the role of nervous system metabolic transporters. *Exp Neurol* 309, 23-31, doi:10.1016/j.expneurol.2018.07.009 (2018).
- 276 Vardjan, N. *et al.* Enhancement of astroglial aerobic glycolysis by extracellular lactatemediated increase in camp. *Frontiers in Molecular Neuroscience* **11**, doi:10.3389/fnmol.2018.00148 (2018).
- 277 Vodovozov, W. *et al.* Metabolic modulation of neuronal gamma-band oscillations. *Pflügers Archiv - European Journal of Physiology* **470**, 1377-1389, doi:10.1007/s00424-018-2156-6 (2018).

- Terada, T. *et al.* In vivo mitochondrial and glycolytic impairments in patients with alzheimer disease. *Neurology* 94, e1592-e1604, doi:10.1212/wnl.000000000009249 (2020).
- 279 Castellano, C.-A. *et al.* Regional brain glucose hypometabolism in young women with polycystic ovary syndrome: Possible link to mild insulin resistance. *PLOS ONE* **10**, e0144116, doi:10.1371/journal.pone.0144116 (2015).
- 280 Khosravi, M. *et al.* 18f-fdg is a superior indicator of cognitive performance compared to 18f-florbetapir in alzheimer's disease and mild cognitive impairment evaluation: A global quantitative analysis. *J Alzheimers Dis* **70**, 1197-1207, doi:10.3233/jad-190220 (2019).
- 281 Chowdhury, G. M. I., Jiang, L., Rothman, D. L. & Behar, K. L. The contribution of ketone bodies to basal and activity-dependent neuronal oxidationin vivo. *Journal of Cerebral Blood Flow & Metabolism* **34**, 1233-1242, doi:10.1038/jcbfm.2014.77 (2014).
- 282 von Morze, C. *et al.* Direct assessment of renal mitochondrial redox state using hyperpolarized 13 c-acetoacetate. *Magnetic Resonance in Medicine* **79**, 1862-1869, doi:10.1002/mrm.27054 (2018).
- 283 Hasan-Olive, M. M. *et al.* A ketogenic diet improves mitochondrial biogenesis and bioenergetics via the pgc1α-sirt3-ucp2 axis. *Neurochemical Research* 44, 22-37, doi:10.1007/s11064-018-2588-6 (2018).
- McCarty, M. F., DiNicolantonio, J. J. & O'Keefe, J. H. Ketosis may promote brain macroautophagy by activating sirt1 and hypoxia-inducible factor-1. *Medical Hypotheses* 85, 631-639, doi:10.1016/j.mehy.2015.08.002 (2015).
- 285 Stekovic, S. *et al.* Alternate day fasting improves physiological and molecular markers of aging in healthy, non-obese humans. *Cell Metabolism* **31**, 878-881, doi:10.1016/j.cmet.2020.02.011 (2020).
- 286 Lourenco, M. V. *et al.* Exercise-linked fndc5/irisin rescues synaptic plasticity and memory defects in alzheimer's models. *Nat Med* **25**, 165-175, doi:10.1038/s41591-018-0275-4 (2019).
- 287 Pedersen, B. K. Physical activity and muscle–brain crosstalk. *Nature Reviews Endocrinology* **15**, 383-392, doi:10.1038/s41574-019-0174-x (2019).
- 288 Cao, B. *et al.* Comparative efficacy and acceptability of antidiabetic agents for alzheimer's disease and mild cognitive impairment: A systematic review and network meta-analysis. *Diabetes, Obesity and Metabolism* **20**, 2467-2471, doi:10.1111/dom.13373 (2018).
- Li, A., Yau, S.-y., Machado, S., Yuan, T.-F. & So, K.-F. Adult neurogenic and antidepressant effects of adiponectin: A potential replacement for exercise? *CNS & Neurological Disorders Drug Targets* 14, 1129-1144, doi:10.2174/1871527315666151111125533 (2015).
- 290 Kremen, W. S. *et al.* Influence of young adult cognitive ability and additional education on later-life cognition. *Proceedings of the National Academy of Sciences* **116**, 2021-2026, doi:10.1073/pnas.1811537116 (2019).
- 291 Carapelle, E. *et al.* How the cognitive reserve interacts with β-amyloid deposition in mitigating fdg metabolism. *Medicine* **96**, e5876, doi:10.1097/md.000000000005876 (2017).
- 292 Arenaza-Urquijo, E. M. *et al.* The metabolic brain signature of cognitive resilience in the 80+: Beyond alzheimer pathologies. *Brain* **142**, 1134-1147, doi:10.1093/brain/awz037 (2019).

- 293 Croteau, E. *et al.* A cross-sectional comparison of brain glucose and ketone metabolism in cognitively healthy older adults, mild cognitive impairment and early alzheimer's disease. *Experimental Gerontology* **107**, 18-26, doi:10.1016/j.exger.2017.07.004 (2018).
- 294 Cotto, B., Natarajanseenivasan, K. & Langford, D. Hiv-1 infection alters energy metabolism in the brain: Contributions to hiv-associated neurocognitive disorders. *Prog Neurobiol* **181**, 101616, doi:10.1016/j.pneurobio.2019.101616 (2019).
- 295 Renard, D., Castelnovo, G., Collombier, L., Thouvenot, E. & Boudousq, V. Fdg-pet in creutzfeldt-jakob disease: Analysis of clinical-pet correlation. *Prion* **11**, 440-453, doi:10.1080/19336896.2017.1387348 (2017).
- 296 Bourgognon, J.-M. *et al.* Alterations in neuronal metabolism contribute to the pathogenesis of prion disease. *Cell Death & Differentiation* **25**, 1408-1425, doi:10.1038/s41418-018-0148-x (2018).
- 297 Gunther, E. C. *et al.* Rescue of transgenic alzheimer's pathophysiology by polymeric cellular prion protein antagonists. *Cell Reports* **26**, 1368, doi:10.1016/j.celrep.2019.01.064 (2019).
- 298 Norrving, B. Stroke management recent advances and residual challenges. *Nature Reviews Neurology* **15**, 69-71, doi:10.1038/s41582-018-0129-1 (2019).
- 299 Bazzigaluppi, P. *et al.* Imaging the effects of β-hydroxybutyrate on peri-infarct neurovascular function and metabolism. *Stroke* **49**, 2173-2181, doi:10.1161/strokeaha.118.020586 (2018).
- Lamade, A. M. *et al.* Aiming for the target: Mitochondrial drug delivery in traumatic brain injury. *Neuropharmacology* 145, 209-219, doi:10.1016/j.neuropharm.2018.07.014 (2019).
- 301 Deng-Bryant, Y., Prins, M. L., Hovda, D. A. & Harris, N. G. Ketogenic diet prevents alterations in brain metabolism in young but not adult rats after traumatic brain injury. *Journal of Neurotrauma* **28**, 1813-1825, doi:10.1089/neu.2011.1822 (2011).
- 302 Arifianto, M., Ma'ruf, A., Ibrahim, A. & Bajamal, A. Role of hypertonic sodium lactate in traumatic brain injury management. *Asian Journal of Neurosurgery* **13**, 971, doi:10.4103/ajns.ajns_10_17 (2018).
- 303 Koch, H. & Weber, Y. G. The glucose transporter type 1 (glut1) syndromes. *Epilepsy & Behavior* **91**, 90-93, doi:10.1016/j.yebeh.2018.06.010 (2019).
- 304 Bakker, A., Albert, M. S., Krauss, G., Speck, C. L. & Gallagher, M. Response of the medial temporal lobe network in amnestic mild cognitive impairment to therapeutic intervention assessed by fmri and memory task performance. *NeuroImage: Clinical* 7, 688-698, doi:10.1016/j.nicl.2015.02.009 (2015).
- 305 Dean, B., Thomas, N., Scarr, E. & Udawela, M. Evidence for impaired glucose metabolism in the striatum, obtained postmortem, from some subjects with schizophrenia. *Translational Psychiatry* **6**, e949-e949, doi:10.1038/tp.2016.226 (2016).
- 306 Kraeuter, A. K., Loxton, H., Lima, B. C., Rudd, D. & Sarnyai, Z. Ketogenic diet reverses behavioral abnormalities in an acute nmda receptor hypofunction model of schizophrenia. *Schizophrenia Research* **169**, 491-493, doi:10.1016/j.schres.2015.10.041 (2015).
- 307 Wlodarczyk, A., Wiglusz, M. S. & Cubala, W. J. Ketogenic diet for schizophrenia: Nutritional approach to antipsychotic treatment. *Med Hypotheses* **118**, 74-77, doi:10.1016/j.mehy.2018.06.022 (2018).
- 308 Palmer, C. M. Ketogenic diet in the treatment of schizoaffective disorder: Two case studies. *Schizophrenia Research* **189**, 208-209, doi:10.1016/j.schres.2017.01.053 (2017).

- 309 Gross, E. C., Lisicki, M., Fischer, D., Sandor, P. S. & Schoenen, J. The metabolic face of migraine - from pathophysiology to treatment. *Nature reviews. Neurology* **15**, 627-643, doi:10.1038/s41582-019-0255-4 (2019).
- 310 Country, M. W. Retinal metabolism: A comparative look at energetics in the retina. *Brain Research* **1672**, 50-57, doi:10.1016/j.brainres.2017.07.025 (2017).

Acknowledgements

This article is based on the proceedings of a small, focused symposium organized by M.J.M. and supported by an unrestricted grant from Advances in Neuroscience for Medical Innovation, which is affiliated with the Institut de Recherche Servier. S.C.C. was supported by the Alzheimer Association (USA), CIHR, FRQS, NSERC and Nestlé, and thanks Valérie St-Pierre, Mélanie Fortier, Alexandre Castellano, Étienne Myette-Côté, Etienne Croteau, Maggie Roy, Marie-Christine Morin and Camille Vandenberghe in particular for outstanding help. M.J.M. thanks Jean-Michel Rivet for help with preparation of the original graphic artwork and Muriel Gaillot and her team for the ordering and provision of pdfs consulted in the preparation of this manuscript. R.H.S. is supported by P30 AG035982, R01 AG060733, and R01 AG061194 from the NIH. E.T. was supported by the National Institutes of Health (NIH) National Institute on Aging (NIA) grant RF1AG55549 and National Institute of Neurological Disorders and Stroke (NINDS) R01NS107265 and RO1AG062135 grants. Z.B.A. was supported by a Senior Research Fellowship from the National Health and Medical Research Council of Australia (APP1154974). P.I.M. was supported by funding from Alzheimer's Association NIRG-13-282387; FEDER funds through the Operational Programme Competitiveness Factors and by the Portuguese Foundation for Science and Technology (grants PEst-C/SAU/LA0001/2013-2014 and UIDB/04539/2020). G.C. is supported by the NIH 1R15AG050292 and 1R21AG064479 grants. R.D.B. was supported by NIA grants R37AG053589, R01AG057931 and P01-AG026572. J.H.J. was supported by grants from the Canadian Institutes of Health Research, Alberta Prion Research Institute, the Alzheimer's Society of Alberta and Northwest Territories and the University Hospital Foundation (Edmonton, AB, Canada). L.H.B. holds grants from Nasjonalforeningen-Demensforbundet, Norway. A.E. was supported by the Swiss National Science Foundation, grant #31003A-179294. O.K. was supported by the Deutsche Forschungsgemeinschaft (grant CRC 1134, B02). M.P.M. was supported by the Medical Research Council UK (MC U105663142) and by a Wellcome Trust Investigator award (110159/Z/15/Z). F.S. has received funding from the European Research

Council (ERC) under the European Union's Horizon 2020 research and innovation program (ADG grant agreement No. 834317). A.E. was funded by NF grant#31003A-179294. W.F.M. was supported by ERC ADG 666053 and VW 93046. A.P. was supported by the Deutsche Forschungsgemeinschaft (PR1527/5-1) and the German Federal Ministry of Education and Research (AZ.031A318 and 031L0211).

Competing interests

S.C.C. declares that he has consulted for and has received honoraria, test products and/or research funding from Abitec, Accera, Bulletproof, Nestlé and Servier, is the founder of Senotec and is co-inventor on a patent for an MCT formulation. M.J.M. declares that he is a full-time employee of Servier and has no other interests to declare. M.P.M. declares that he holds patents related to therapies targeted at decreasing oxidative damage to mitochondria. G.C. declares that she holds a patent related to compositions and methods for treating cognitive deficits using amylin and other hormones. A.E. declares that she has received honoraria, test products and/or research funding from Schwabe and Vifor. F.S. declares that he has consulted for Servier and TEVA. R.D.B. declares that she holds patents for therapeutics targeting Alzheimer disease and NDAs and is the founder of NeuTherapeutics. E.T. declares that she holds a patent related to compositions and methods for treating complex I inhibitors. All other authors declare no competing interests.

Publisher's note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Supplementary information

Supplementary information is available for this paper at https://doi.org/10.1038/s415XX-

TOC blurb

Accumulating evidence indicates that impaired glucose metabolism in the brain is involved in the aetiology and progression of neurodegenerative disorders of ageing such as Alzheimer disease. This Review discusses the status and prospects of therapeutic strategies for countering neurodegenerative disorders of ageing by rescuing, protecting or normalizing brain energetics.