

1 **Interpreting oral fluid drug results from prisoners: monitoring current**
2 **drug intake and detection times for drugs self-administered prior to**
3 **detention**

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26

27 **Abstract**

28

29 **Purpose** Urine is the most common matrix for prisoner drug testing, although oral fluid offers
30 a possible alternative. Identifying new drug intake by a prisoner results in negative sanctions.
31 Detection times in oral fluid after chronic drug intake may be extended. Within the prison
32 admission population are chronic drug users. Our aim was to investigate drugs of abuse
33 detection windows in oral fluid from prisoners.

34

35 **Methods** Nineteen frequent drug abusing prisoners provided oral fluid and urine at
36 admission, and each morning for 9 consecutive days.

37

38 **Results** The most positive findings were for amphetamine/ methamphetamine, cannabis and
39 benzodiazepines. Maximum detection times in oral fluid were ≥ 9 days for diazepam,
40 methadone and methamphetamine, with corresponding urinary detection times of ≥ 9 , 7 and 6
41 days. Maximum oral fluid detection times were nine days for clonazepam, eight for
42 oxazepam, three for amphetamine and nitrazepam and two for tetrahydrocannabinol, with
43 positive urinary detection times of 8, ≥ 9 , 5, 7 and ≥ 9 days, respectively. Cocaine, morphine
44 and 6-acetylmorphine were positive only one day in oral fluid, and one and two days,
45 respectively, in urine, while 6-acetylmorphine was not detected in urine.

46 **Conclusion** We confirmed oral fluid as a viable matrix for monitoring drugs of abuse in
47 prisoners. Windows of detection for benzodiazepines and amphetamines were up to one week,
48 an important consideration for evaluating oral fluid drug testing results. Some likely new drug
49 exposures were observed based on urine and oral fluid drug results, but there are few data
50 guiding these interpretations.

51

52 **Introduction**

53

54 Prisoners are frequently drug tested, with urine as the preferred matrix. Observed urine
55 collections are time consuming and many donors consider it as an intrusion of privacy. Due to
56 advances in analytical technology for oral fluid testing, this biological matrix is now a viable
57 alternative to urine testing in several disciplines [1-5]. The easy, fast and gender-neutral oral
58 fluid sample collection can take place in almost any location, with less embarrassment for the
59 donor, giving oral fluid significant advantages over urine.

60

61 In Norway, urine is collected on admission to prison, and creatinine-corrected urine
62 sample concentrations taken at regular intervals thereafter, are interpreted to determine if
63 results are likely to represent new intake within the prison or residual excretion from intake
64 before imprisonment. Replacement of urine with oral fluid as the testing matrix requires a
65 scientific basis, and although data exist on drug elimination in oral fluid from controlled
66 administration studies, these results might not be representative for samples collected from
67 prisoners with chronic and/or high drug intake.

68

69 Drug windows of detection in oral fluid are considered short, and more similar to
70 blood than urine [6; 7]. The detection periods are thus highly dependent upon both the chosen
71 cut-off concentrations, and the dose ingested [8; 9]. Multiple studies documented that oral
72 fluid is a viable matrix for drugs of abuse detection [10-15]. Single and low doses are
73 typically administered in controlled drug studies [16-26], although others investigating drug
74 elimination purported high doses from patients admitted for drug detoxification [27-30] or
75 after chronic frequent use [31; 32] reported increased drug detection times. Since many

76 prisoners use high and/or chronic doses of drugs of abuse before incarceration, elimination
77 and detection times of drugs of abuse in oral fluid from this population provide relevant data
78 for future interpretation of oral fluid tests. The aim of this study was to investigate drugs of
79 abuse windows of detection in oral fluid after possible ingestion of high doses or chronic
80 frequent drug use, at the time of prisoner incarceration and the following 9 days. Drug use is
81 prohibited in prison and inmates are under sustained and monitored abstinence.

82

83

84 **Materials and methods**

85 **Study group**

86 In total, 19 inmates from three prisons were enrolled in the study. Drug consumption prior to
87 incarceration was self-reported. Information regarding prescribed drugs during the study was
88 provided by the prison physician. The only relevant medications reported were buprenorphine
89 and methadone for opioid-dependence treatment and oxazepam.

90

91 Positive drug test results produced no negative consequences for participants, as the
92 prisons did not receive results. Participants received no payment for providing samples. Each
93 participant had a unique code linked to their self-report data and samples, and only one person
94 in each prison had access to these data. Everyone else was blind to prisoner identity, and only
95 participants' unique codes were reported.

96

97 **Sample collection**

98 Sampling occurred the day of and for 9 days after prison admission (reported as day 0 to day
99 9), for a total of 10 oral fluid samples per participant. Since drug intake might have occurred

100 on day 0, positive oral fluid samples collected on day 1 were considered as having a detection
101 time of 1 day. Oral fluid samples were collected each morning, and if possible, each first
102 voided urine also. Oral fluid samples were collected with the commercially available
103 Intercept® Oral Specimen Collection Device (OraSure Technologies, Bethlehem, PA, USA).
104 The cotton pad on a stick was placed between the cheek and gum for 2 min to sample oral
105 fluid according to manufacturer's recommendations. All samples were weighed to obtain the
106 amount of oral fluid collected. The collection pad contains preservatives and citric acid,
107 stimulating oral fluid production, and collecting a mixture of saliva, gingival crevicular fluid
108 and mucosal transudate. After collection, the pad was placed into a vial containing 0.8 mL
109 stabilizing buffer solution and stored at -20°C until analysis. The urine sample was collected
110 in a 120 mL BD-Vacutainer urine collection cup with integrated transfer device (Becton,
111 Dickinson and Company, Franklin Lakes, NJ, USA) and transferred to Vacuette® vials
112 without additives (Med-Kjemi A/S, Asker, Norway) before transport to the laboratory.

113

114 **Analytical methods**

115 Urine samples were screened for amphetamines (EMIT DAU reagents, Siemens, Healthcare
116 AS, Oslo, Norway), cannabis, cocaine, methadone, opiates (EMIT II Plus reagents, Siemens
117 Healthcare AS) and benzodiazepines (CEDIA reagents, Thermo Fisher Microgenics, Fremont,
118 CA, USA) by immunological methods on the Hitachi 917 analyzer (Hitachi, Tokyo, Japan).
119 In addition, pH (DRI® pH-Detect Test; Thermo Fisher Microgenics) and creatinine (DRI®
120 Creatinine-Detect® Test; Thermo Fisher Microgenics) were measured. γ -hydroxybutyrate,
121 GHB, was screened by ultrahigh performance- liquid chromatography–tandem mass
122 spectrometry (UHPLC-MS/MS) [33]. Confirmation analyses were performed by liquid-
123 chromatography–tandem mass spectrometry (LC–MS/MS) for benzodiazepines [34] and

124 UHPLC–MS/MS for opiates and cocaine [35]. Amphetamines, methadone and 11-nor-9-
125 carboxy- Δ 9- tetrahydrocannabinol, THCCOOH, were analysed by internally-validated
126 UHPLC-MS/MS and gas chromatography–mass spectrometry (GC–MS) methods,
127 respectively. Oral fluid samples were analysed for drugs of abuse by a quantitative LC-MS-
128 MS method [36]. Cut-off concentrations in oral fluid and urine are shown in Table 1. Urine
129 validation data for amphetamine, methamphetamine and THC-COOH are presented in Table
130 4, together with urine validation data for the other compounds presented in figures 2 to 4.

131

132 **Statistics**

133 The data were analysed using IBM SPSS Statistics 23 (IBM). Pearson’s correlation was used
134 to investigate the relationship between concentrations in oral fluid and urine.

135

136 **Results**

137 Demographic data and self-reported prior drug intake for the 17 male and 2 female
138 participants are shown in Table 2. Fifteen subjects provided biological samples for all ten
139 days of the study and the remaining four for five, seven, eight and nine days, respectively.

140 The longest detection times for each drug and/or metabolite are reported in Table 3. It is
141 important to emphasize that drugs might have been consumed prior to admission (day 0), and
142 for those drugs still detected on day nine, detection times might be longer, because later
143 samples were not collected or analysed.

144

145 **Amphetamine/methamphetamine**

146 Amphetamine and methamphetamine were detected together in 11 participants’ oral fluid
147 and/or urine, while one participant’s biological samples contained only amphetamine, subject

148 15, and one only methamphetamine, subject 5. Amphetamine was identified in oral fluid
149 from day 0 to 3 days, and for methamphetamine from day zero to nine days. Amphetamine
150 was detected in urine from day zero to day five, and for methamphetamine from day zero to
151 six days. The longest amphetamine detection time was in urine, while for methamphetamine it
152 was in oral fluid. As seen in Fig. 1, the biological sample with the longest detection time
153 varied between subjects. The prisoners self-reported previous amphetamine, but not
154 methamphetamine use. If self-reported ingestion times were considered, detection times were
155 longer, with a maximum of ten days for amphetamine and 15 days for methamphetamine.

156

157

158 <Figure 1 here>

159

160 **Opioids**

161 Morphine and/or 6-acetylmorphine (6-AM) were detected in two participants' samples.

162 Heroin is metabolised rapidly to 6-AM and morphine, and later morphine-3-glucuronide

163 (M3G) and morphine-6-glucuronide (M6G). Subject 8 self-reported heroin consumption the

164 day before admission (day -1) and morphine, M3G and M6G were identified in his/her urine

165 through day two, Table 2. Opioids were not detected in any of his/her oral fluid samples.

166 Subject 11 self-reported heroin ingestion three days before imprisonment (day -3); 6-AM was

167 identified in oral fluid on day 1, but not in any urine sample. Morphine was detected in the

168 oral fluid sample on day 0 and in urine until day 1. Considering the self-reported time of the

169 last exposure, morphine's window of detection was four days in urine, and the detection time

170 for morphine/6-AM was three days in oral fluid. These estimates are, however, uncertain.

171

172 Methadone was detected in samples from two participants. Subject 2 allegedly
173 ingested massive amounts of drugs prior to admission (methadone, heroin, cannabis and
174 diazepam; doses and times of ingestion were not given) and had a nonfatal overdose. Urine
175 samples showed decreasing methadone concentrations detectable until day 7, and methadone
176 was detected in oral fluid through day 9. The other participant, subject 4, received opioid-
177 dependence treatment during the study, and no detection time window can be given.

178

179 In all cases where buprenorphine was detected, it was given as opioid-dependence
180 treatment. This makes it impossible to estimate the window of detection for buprenorphine.

181

182 **Δ 9-Tetrahydrocannabinol (THC)**

183 In 15 participants 11-nor-9-carboxy- Δ 9-tetrahydrocannabinol (THCCOOH) was detected in
184 urine on day 1. Of these, only two participants had detectable THC in oral fluid, and only on
185 day 0. Seven participants had detectable THCCOOH in urine throughout the study. However,
186 most participants claimed last cannabis intake several days before admission. One participant
187 tested positive for THC in oral fluid on days 4 and 5, with prior negative samples. Oral fluid
188 concentrations corrected for dilution were 7.7 and 14.2 μ g/L, respectively. Urine creatinine-
189 corrected THCCOOH increased from 15 ng/mg on day 1 and <cut-off on day 2, to 78,171 and
190 203 ng/mg creatinine on days three, four and five, respectively. Day 3 urine concentrations
191 were assessed as new cannabis intake using the reference values of U2/U1 ratios reported by
192 Smith et al. [37]. Clearly positive THC results in oral fluid on days 4 and 5 also indicate new
193 cannabis intake in prison, Fig. 2.

194

195 <Figure 2 here>

196

197 **Benzodiazepines**

198 Clonazepam

199 Clonazepam was identified in seven prisoners' urine or oral fluid samples, and none received
200 medical treatment with clonazepam in prison. Maximum clonazepam detection time was eight
201 days in urine (range one to eight days), and in oral fluid, at least nine days (range one to nine
202 days) if the positive clonazepam samples for subject 15 on days 0 and 9 only are included.

203 Clonazepam and 7-aminoclonazepam oral fluid concentrations in one participant on day 8
204 were 97 and 6.4 µg/L, respectively, while all prior urine and oral fluid samples were negative.
205 This suggested a new clonazepam intake after admission to prison.

206

207 Nitrazepam

208 Nitrazepam/7-aminonitrazepam were detected in oral fluid from three participants. In urine,
209 detection times ranged from two to seven days, and in oral fluid from one to three days.

210 Considering the subjects' self-reported last intake, the detection time did not change. Fig. 3
211 shows the elimination curves for nitrazepam and 7-aminonitrazepam in oral fluid from
212 participants 14 and 18, the prisoners with the longest detection times, and the corresponding
213 creatinine-normalized urine elimination curves for 7-aminonitrazepam.

214 <Figure 3 here>

215

216 Oxazepam

217 Eleven participants had oxazepam in either urine or oral fluid. Two subjects used oxazepam
218 when admitted to prison. No information about doses was available for subject 7, who,
219 according to our information, stopped taking oxazepam during the study; however, the date

220 was not given. For subject 10, 25 mg Sobril[®] was prescribed morning and evening during the
221 study, making it impossible to determine detection times in either matrix. For the other
222 inmates, oxazepam was found with other diazepam metabolites. Windows of detection for
223 oxazepam ranged from two to \geq nine days in oral fluid and urine, with generally longer
224 detection times in urine than oral fluid samples. Additionally, it was difficult to distinguish
225 the source of oxazepam, as it also is a metabolite of other benzodiazepines including
226 diazepam. One person (subject 14) disclosed oxazepam ingestion the day before incarceration
227 to prison, with detection times of six days in oral fluid and seven days in urine; however,
228 presence of other diazepam metabolites demonstrates that there was intake of other
229 benzodiazepine(s) also.

230

231 Diazepam/*N*-desmethyldiazepam

232 Diazepam or metabolites were identified in eight inmates' samples. Maximum diazepam
233 detection times in oral fluid ranged from four to \geq nine days, and in urine from one to \geq nine
234 days. Participant 4 only had positive *N*-desmethyldiazepam in oral fluid, and 3-
235 hydroxydiazepam and oxazepam in urine, and did not declare diazepam intake. For four
236 participants, diazepam and its metabolites were detected in oral fluid for the entire study
237 period, but there was no self-report of time of last intake. However, one person with positive
238 samples during the entire study claimed that the last diazepam ingestion was at least 13 days
239 before admission.

240

241 **Cocaine**

242 Cocaine and its metabolite benzoylecgonine were detected in subject 5's oral fluid and
243 benzoylecgonine in a urine sample only on day 3, after negative tests the days before. Oral

244 fluid concentrations corrected for dilution were 4.4 µg/L for cocaine and 9.9 µg/L for
245 benzoylecgonine. In urine samples, cocaine was negative, while the benzoylecgonine
246 concentration was 2262 µg/L and the creatinine-normalized result was 1122 ng/mg. This
247 finding was interpreted as ingestion of cocaine after admission.

248

249 **Correlation between oral fluid and urine results**

250 Oral fluid and urine samples collected on the same days were compared. For most drugs, both
251 matrices were initially positive, but last detection varied according to matrices. Cannabis was
252 an exception, as there were many positive urine samples without matching positive oral fluid
253 samples. For amphetamine, a trend towards longer detection time in urine could be seen,
254 while methamphetamine tended to have longer detection times in oral fluid, Fig. 1. Oxazepam
255 had longer detection times in urine, while for *N*-desmethyldiazepam evaluation was difficult
256 as most samples were positive in both oral fluid and urine at the end of the study. For the
257 other compounds, the number of cases was too small to infer any trends. Direct comparison of
258 quantitative results for oral fluid and creatinine-corrected urine concentrations for the four
259 most prevalent drugs is shown in Fig. 4. Pearson's correlation was used to investigate the
260 relationship between concentrations in oral fluid and urine, and we found correlation
261 coefficients of 0.612 (methamphetamine), 0.314 (amphetamine), 0.535 (7-aminoclonazepam)
262 and 0.553 (*N*-desmethyldiazepam). The correlations were significant ($p < 0.01$) for
263 methamphetamine, 7-aminoclonazepam and -desmethyldiazepam, but not ($p = 0.086$) for
264 amphetamine.

265

266 <Figure 4 here>

267

268

269 **Discussion**

270

271 We investigated the detection times of drugs of abuse in oral fluid and urine samples using 19
272 prisoners with a history of drug abuse, while under constant supervision. Individual
273 elimination curves (Figs. 2 and 3) of creatinine-normalized urine results were used for
274 comparison to see if variation in oral fluid results was likely to be the result of new intake
275 during the study. There was a larger variability in elimination curves in oral fluid as compared
276 to creatinine corrected urine curves, in line with the previous findings [27-30]. In addition,
277 after ingestion of high and repeated drug doses, detection times could be several days. Despite
278 significant correlation between oral fluid and urine concentrations for more of the drugs, it is
279 not possible to infer the concentration in urine from oral fluid and vice versa, Fig. 4. At the
280 end of the elimination curve of a drug, a positive sample following after a negative can be
281 found in any matrix, as the concentration fluctuates around the limit of
282 quantification/detection. Oral fluid concentrations tend to be more variable than e.g. blood
283 concentrations, and this effect, is therefore more pronounced in oral fluid. Negative samples
284 interspersed with positive findings were encountered for some in our study, which is
285 consistent with other elimination studies. [16; 27-31]. Detection times were longer than in
286 controlled single dose administration studies [16; 38; 39]. As many prison inmates have a
287 chronic drug problem, these data are important because they represent long term intake of
288 high drug doses based on self-report.

289

290 **Benzodiazepines**

291 Benzodiazepines are popular drugs of abuse, and frequently included in drug testing
292 programs. Few studies investigated windows of benzodiazepine detection in oral fluid [6; 7;

293 29; 40; 41]. A summary by Kidwell et al. [42] reported detection times for diazepam and
294 nitrazepam of 48 and 70 h, respectively, after ingestion of single doses. This is comparable to
295 our previous study of patients undergoing drug detoxification, where diazepam was found as
296 *N*-desmethyldiazepam in oral fluid for the entire nine days, applying a cutoff of 1.3 µg/L [29].
297 Our current research documented a window of detection for diazepam of at least 9 days in
298 oral fluid and urine, and *N*-desmethyldiazepam had the longest detection time in oral fluid
299 compared to urine.

300

301 Detection times for clonazepam were at least 6 days in oral fluid and 8 days in urine,
302 Table 3. One participant was positive for clonazepam in oral fluid on admission day and day 9
303 only; 7-aminoclonazepam fluctuated around the cutoff, extending the detection time to at least
304 9 days in oral fluid. This is comparable to our previous study of patients undergoing drug
305 detoxification, where 7-aminoclonazepam was detected for 6 days [29], with a cutoff of 1.3
306 µg/L.

307

308 Few data are available for nitrazepam elimination in oral fluid. Nitrazepam oral fluid
309 C_{\max} was 1.9 µg/L after 5 mg nitrazepam and the drug was quantifiable up to 70 h (approx. 3
310 days) with a limit of quantification (LOQ) of 0.5 µg/L [43]. As Fig. 3 shows, we also found
311 low 1-2.5 µg/L initial nitrazepam concentrations that decreased over three days. No data were
312 provided about the time of intake of nitrazepam in our study. Nitrazepam/7-aminonitrazepam
313 was detected for three days in oral fluid and 7 days in urine, and 7-aminonitrazepam had
314 higher concentrations than nitrazepam in all samples.

315

316 **Opioids**

317 Only one study to our knowledge investigated heroin's oral fluid window of detection [30],
318 but others reported that 6-AM is more frequently detected in oral fluid as compared to urine
319 [44]. Opioids were only found in samples from two participants. In one case, 6-AM was
320 detected in oral fluid, but not in urine. In the other case, the opposite situation occurred. This
321 documents individual variability that must be taken into account when interpreting results.
322 Our window of detection for methadone in oral fluid of at least 9 days, Table 2, subject 2, is
323 similar or longer than the five and eight days previously reported from patients undergoing
324 drug detoxification [30].

325 **Amphetamines**

327 Few studies investigated windows of detection for amphetamines in oral fluid. Huestis and
328 Cone [24] showed that after sequential daily dosing of 20 mg methamphetamine for four
329 days, a clear accumulation of methamphetamine in oral fluid was observed. Positive
330 specimens were reported for approximately 24 h at a 2.5 µg/L cut-off. Schepers et al. [16]
331 also reported detection times in oral fluid for amphetamine and methamphetamine up to 24 h
332 at the same cut-off after a 20 mg dose of methamphetamine. Methamphetamine was
333 measurable for 36 – 72 h after the last of four doses. As could be expected assuming higher
334 intake, we found a much longer 9 day methamphetamine window of detection than reported
335 in clinical studies, with a 8 µg/L cutoff, Table 3. This is slightly longer than in our previous
336 study from patients undergoing drug detoxification, where the detection window was up to
337 eight days [27]. For amphetamine, a shorter detection window of up to three days was found,
338 Table 3, as compared to the previously reported detection window of up to 8 days for patients
339 undergoing drug detoxification [27]. It might be difficult to differentiate the effects of
340 amphetamine and methamphetamine [45]; thus there was consistency between the

341 participants' self-reports regarding methamphetamine/amphetamine ingestion and the actual
342 findings in oral fluid/urine.

343

344 **THC**

345 THC is metabolized to the inactive metabolite THCCOOH, which can be detected in urine for
346 weeks after stopping chronic frequent cannabis intake [46]. Lee et al. [31] showed that the
347 detection time for THC in oral fluid among chronic frequent cannabis smokers ranged from
348 48 h to 28 days, with negative samples (<0.5 µg/L) interspersed with a few positive samples,
349 raising into question the possibility of reuse despite 24 h surveillance on a closed research
350 unit. In patients undergoing detoxification, Andås et al. [28] reported an oral fluid THC
351 window of detection of 8 days (0.3 µg/L LOQ). In the present study, a 0.9 µg/L cutoff was
352 applied, and THC was detected only in oral fluid samples from two subjects, with the longest
353 detection time of 1 day. The difference could in part attributed to a higher cutoff, but it could
354 also indicate that participants in Lee' and Andås' studies [28; 31] had greater and more
355 frequent cannabis intake.

356 New cannabis intake was suggested for subject 5, with similar findings of THC in oral
357 fluid and urine on days 4 and 5 after admission, Fig. 2. These data support oral fluid as a
358 matrix to reveal drug use in prison. However, the aforementioned possibility of negative
359 samples interspersed with positive findings must also be considered [31].

360

361 **Cocaine/benzoylecgonine**

362 Cocaine or benzoylecgonine were only identified in one participant's oral fluid samples on
363 day 3, Table 2, subject 5; these results were interpreted as new cocaine intake in prison. The

364 transfer of cocaine from blood to oral fluid depends on oral fluid pH. Cocaine has a short
365 detection time in oral fluid, as also occurs for this analyte in blood and urine [22; 47; 48].

366

367 **Limitations**

368 The limitations of the study include the number of participants and single oral fluid and urine
369 samples each day. However, valuable oral fluid detection time data from individuals with
370 histories of potentially high and repeated drug intake are included, as well as comparison of
371 paired oral fluid and urine data. Limited studies investigated this population. Detection times
372 for benzodiazepines and amphetamines in oral fluid were consistent with or somewhat longer
373 than previously reported data, while detection times for opiates and THC were shorter. It is
374 important to emphasize that the study period was 10 days, leading to maximum detection
375 times of at least 9 days (Table 3), while intake was varied prior to imprisonment.

376

377 **Conclusions**

378 Oral fluid was a viable alternative to urine for monitoring drugs of abuse in prison. Oral fluid
379 is easier to collect and much less subject to adulteration than urine. Our study confirms that
380 long detection times, especially for amphetamines and benzodiazepines, can be encountered
381 in this population, although oral fluid cannabinoid results had a much lower prevalence than
382 urine tests. From daily oral fluid concentrations, it might be possible to identify new drug
383 intake, but elimination curves were not as consistent as seen in blood [49] or creatinine-
384 corrected urine. Negative oral fluid samples might be interspersed with positive findings as
385 noted with urine samples, especially when concentrations are close to applied cutoffs.

386

387

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390 Tore Kopperud for organizing inclusion of prisoners into the project and excellent sample
391 collection.

392

393 **Ethical approval**

394 The Norwegian Regional Committee for Medical and Health Research Ethics approved the
395 study. Participation was voluntary, and participants could withdraw at any time without
396 penalty. Written informed consent from 19 inmates from three prisons was obtained prior to
397 inclusion after fully informing participants about the study.

398

399

400 **Conflict of interest**

401

402 The authors declare no conflicts of interest.

403

404 This study has not received any external financial support.

405

406

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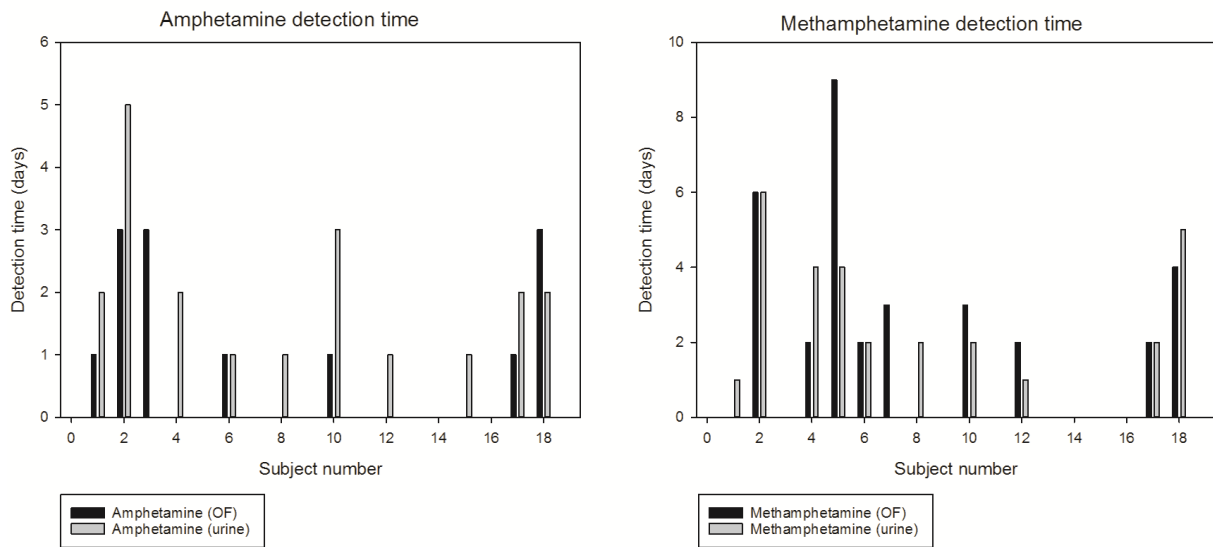
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548 **Figures**

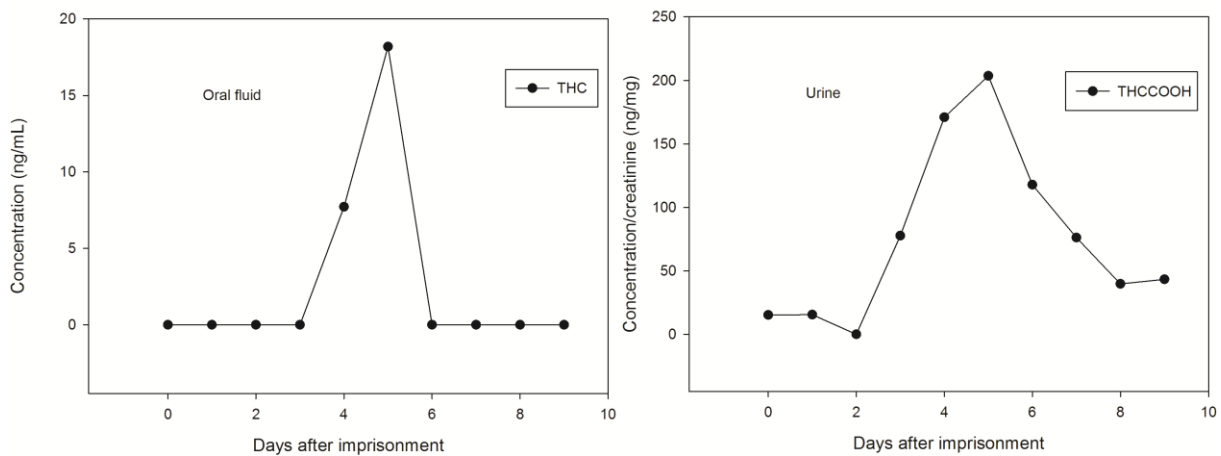


549

550 **Fig. 1** Detection time in oral fluid (OF) and urine for amphetamine (left panel) and

551 methamphetamine (right panel)

Cannabis
Subject 5

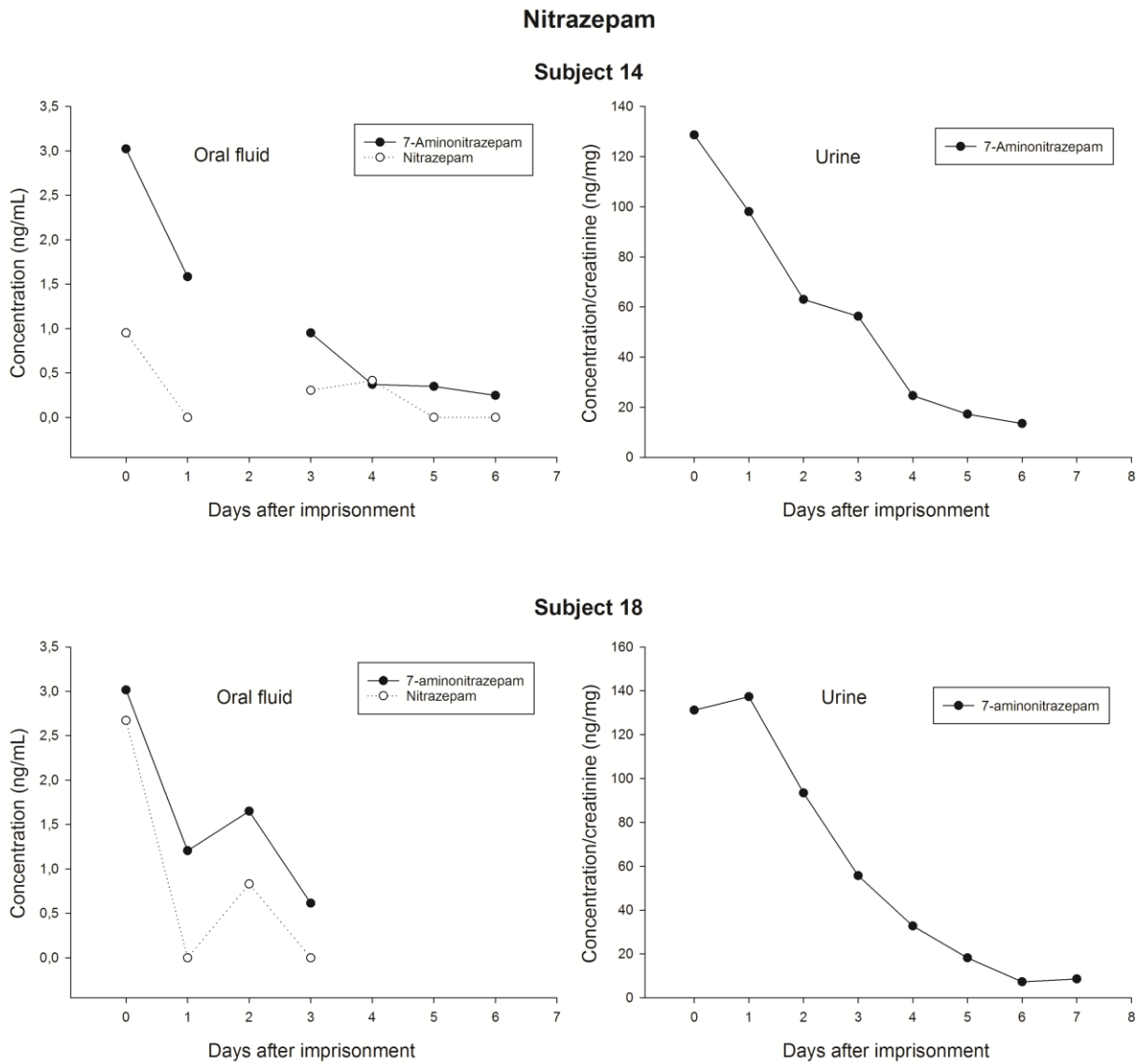


552

553 **Fig. 2** OF and creatinine-normalized urine concentrations from subject 5, with probable new

554 intake of cannabis. THC Δ^9 -tetrahydrocannabinol, THCCOOH 11-nor-9-carboxy- Δ^9 -

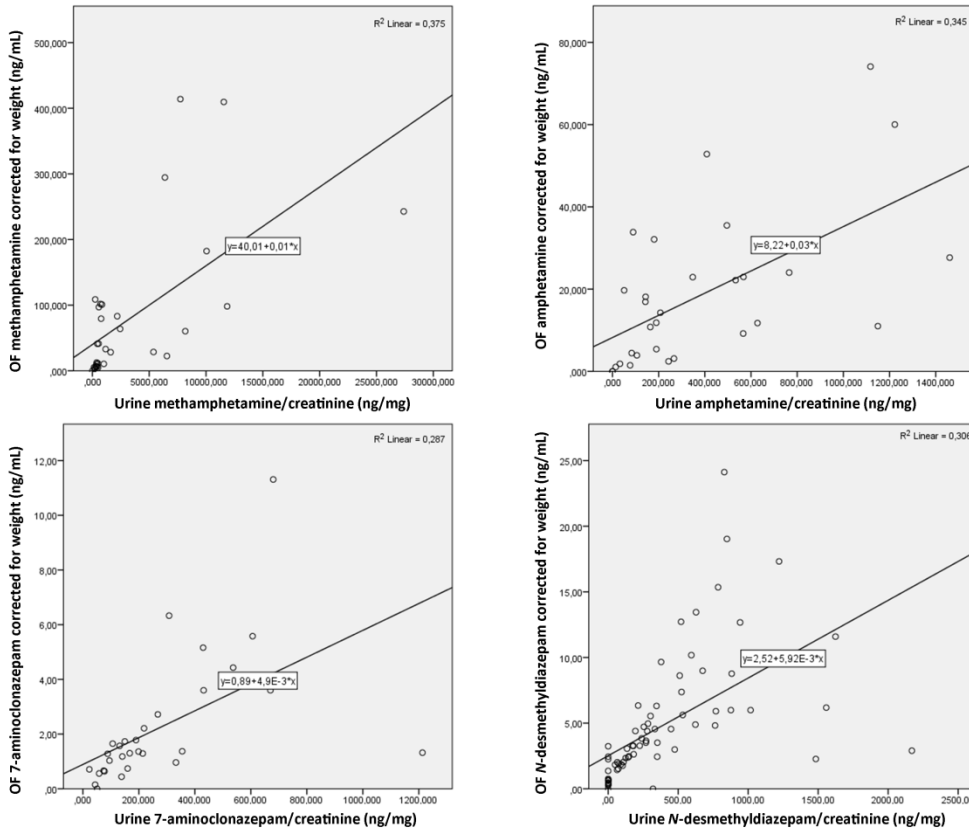
555 tetrahydrocannabinol



557

558

559 **Fig. 3** Elimination curves for nitrazepam and 7-aminonitrazepam in OF and urine samples
 560 from participants 14 and 18, with the longest detection times. No OF results were available on
 561 day 2 due to an analytical error



562

563 **Fig. 4.** Scatter plots and trend lines of the creatinine-normalized urine and OF concentrations
 564 of methamphetamine, amphetamine, 7-aminoclonazepam and N-desmethyldiazepam

565