

CYP1A1 and CYP1B1 in human lung
PAH-bioactivation capacity, sex differences and
steroid receptor mediated regulation

Thesis submitted for the degree of
Philosophiae Doctor
by
Heidi Uppstad



Faculty of Mathematics and Natural Sciences, University of Oslo, Norway
Section of Toxicology and Biological Working Environment, Department of Chemical and
Biological Work Environment. National Institute of Occupational Health, Oslo, Norway

2016

© **Heidi Uppstad, 2016**

*Series of dissertations submitted to the
Faculty of Mathematics and Natural Sciences, University of Oslo
No. 1764*

ISSN 1501-7710

All rights reserved. No part of this publication may be
reproduced or transmitted, in any form or by any means, without permission.

Cover: Hanne Baadsgaard Utigard.
Print production: Reprosentralen, University of Oslo.

Acknowledgements

The work presented in this thesis was carried out at The National Institute of Occupational Health. I am grateful to The Research Council of Norway for supporting this project and The National Institute of Occupational Health for providing excellent facilities.

I would like to express my sincere gratitude to my supervisor Dr. Steen Møllerup. It would have been impossible to finalize this project without his outstanding commitment, knowledge and enthusiasm. His profound expertise on lung carcinogenesis and techniques in molecular biology along with his communication skills have provided the basis for many valuable and inspiring discussions and excellent guiding.

I am thankful to my supporting co-supervisor Professor Johannes Gjerstad and to my co-authors, especially Professor David H. Phillips for his excellent work performed on the measurements of DNA adducts. Professor Steinar Øvrebø and Professor Aage Haugen, thank you for your insightfulness, enthusiasm and valuable help with the manuscripts.

I would like to thank all my former co-workers at Section for Toxicology and Biological Working Environment, especially mentioning Kari Anne Haaland, Kent Hart, Rita Bæra and Unn W. Holm.

My dear friends, Kari Anne Haaland, Tord R. Næss, Marianne L. Møllerløyken, Therese Edvardsen, Dag Storrvik, Eirin D. Johansen and Guro E. R. Birkeland, thank you for making my life full of laughter and precious moments.

To my beloved parents Barbro and Gunnar Uppstad and my brother Bernt Halvar Uppstad. Thank you for your unconditional support, encouragement and love. It has kept me going through both good times and challenging times. I am forever grateful to you.

Table of contents

Acknowledgements	2
List of papers	5
Summary	6
Abbreviations and explanations	9
1.Introduction	11
1.1 Lung cancer	11
1.1.1 Incidence, histology and mortality	11
1.1.2 Lung cancer etiology and lung carcinogens	12
1.1.3 Lung cancer epidemiology and smoking habits are tightly linked	13
1.2 Lung carcinogenesis	15
1.2.1 Stages in carcinogenesis	15
1.2.2 Lung cancer susceptibility	18
1.2.3 Genetic susceptibility to lung cancer	18
1.2.4 Epigenetic alterations	19
1.2.5 Epigenetics in lung cancer	21
1.2.6 Lung cancer and sex differences	21
1.2.7 Animal studies and sex differences in lung cancer biology	23
1.3 Biotransformation of PAH in tobacco smoke.....	24
1.3.1 Chemistry, sources and biotransformation of PAH	24
1.3.2 Aryl hydrocarbon receptor (AHR)	26
1.3.3 Cytochrome P450 1A1 and 1B1 (CYP1A1 and CYP1B1)	27
1.3.4 Metabolism of B[a]P catalyzed by CYP1A1 and CYP1B1	29
1.3.5 B[a]P-DNA adduct formation	30
1.4. Sex steroid receptors and lung cancer	31
1.4.1 Estrogen receptors in lung	31
1.4.2 Hormone replacement therapy (HRT) and antiestrogen treatment	32
1.4.3 A reciprocal association between the signaling pathways of ERs and AHR	34
1.4.4 Androgen receptor in lung	35
1.4.5 A reciprocal association between the signaling pathways of AR and AHR	37
2. Aims of the study	38
3. Summary of papers	39
4. Discussion	42

4.1. The individual roles of CYP1A1 and CYP1B1 in bioactivation of benzo[<i>a</i>]pyrene in human lung cells.....	43
4.2 Sex differences in expression of <i>CYP1A1</i> and capacity to form B[<i>a</i>]P-DNA adducts in human lung cells.....	47
4.3 Sex steroid receptors' impact on <i>CYP1</i> regulation in lung cells	50
5. Conclusions and future perspectives	55
6. References	57
Paper I, II and III.....	76

List of papers

This project has resulted in the following papers, and will be referred to in the text by their roman numbering I, II and III:

Paper I

Heidi Uppstad, Steinar Øvrebø, Aage Haugen and Steen Mollerup. Importance of CYP1A1 and CYP1B1 in bioactivation of benzo[*a*]pyrene in human lung cell lines. *Toxicology Letters* 2010; 192: 221 - 228.

Paper II

Heidi Uppstad, Gro Helene Osnes, Kathleen J. Cole, David H. Phillips, Aage Haugen and Steen Mollerup. Sex differences in susceptibility to PAHs is an intrinsic property of human lung adenocarcinoma cells. *Lung Cancer*. 2011; 71: 264 - 270.

Paper III

Heidi Uppstad, Tove Igeland, Kristine Haugen Anmarkrud, Aage Haugen and Steen Mollerup. Modulatory effect of sex steroid receptors on PAH metabolism genes in human lung cells. *Manuscript*.

Summary

Airway-exposure to polycyclic aromatic hydrocarbons (PAHs) increases the risk of lung cancer in humans. PAHs readily bind to the aryl hydrocarbon receptor (AHR), which is a ligand-inducible transcription factor that regulates transcription of cytochrome P450 (CYP) *1A1* and *CYP1B1*. CYP1A1 and CYP1B1 show similar, but not identical, substrate specificity toward various PAHs, and are responsible for bioactivation of most carcinogenic PAHs to reactive epoxide intermediates. PAH-epoxides generated by CYP1 can react with DNA and form PAH-DNA adducts. Epidemiological data have demonstrated an association between the formation of DNA adducts and an increased risk of lung cancer.

Clinical evidence has accumulated toward a notion of sex differences in the biology of lung cancer. Although not conclusive, studies indicate that women may be at greater risk of developing lung cancer from cigarette smoking than men. Furthermore, women are typically diagnosed at an earlier age than men are. After being diagnosed with lung cancer, however, women appear to display more favorable prognoses than men do. Hence, distinct clinical characteristics differ between men and women.

Although the lung is not considered a typical hormone responsive tissue, steroid receptors such as estrogen and androgen receptors have been reported to be expressed and active in lung cells. The rates and the extent of cellular processes such as metabolic activation of pro-carcinogens like PAHs may influence individual susceptibility to the tumorigenic effects associated with exposure. Studies have indicated that sex steroid receptors may modulate PAH-bioactivation in lung cells and therefore influence lung cancer susceptibility.

The purpose of this thesis was to study the individual roles of the PAH-bioactivation genes *CYP1A1* and *CYP1B1* in human lung, with specific focus on sex differences in expression and a possible modulatory effect of steroid receptors in gene regulation. First we wanted to establish the respective roles of CYP1A1 and CYP1B1 in bioactivation of prototype PAH benzo[*a*]pyrene (B[*a*]P). siRNA was used to knock down expression of *CYP1A1* and *CYP1B1*, gene expression was measured by RT-qPCR and formation of B[*a*]P-metabolites by fluorescence-HPLC. In summary, the results revealed that the overall B[*a*]P-metabolism and

formation of B[a]P-tetrol I-1 (a hydrolysis product of the ultimate B[a]P-carcinogen BPDE I) in human lung cells are primarily dependent on *CYP1A1*. The formation of B[a]P-tetrol I-1 may thus serve as an indicator of the current bioactivation capacity. Hence, the results show that *CYP1A1* plays an important role in bioactivation of B[a]P in lung cells.

The next major objective was to elucidate whether previously reported *in vivo* sex differences in *CYP1A1* expression and PAH-DNA adduct formation are intrinsic characteristics of lung cells. We therefore analyzed basal, cigarette smoke condensate (CSC)- and B[a]P-induced gene expression of *CYP1A1* and *CYP1B1*, CYP1 activity and formation of B[a]P-DNA adducts in eleven non-small cell lung carcinoma (NSCLC) cell lines (five of female and six of male origin). Gene expression was measured by RT-qPCR, CYP1 enzyme activity was analyzed by EROD, and formation B[a]P-adducts was measured by ³²P-postlabelling. Significantly higher levels of basal, CSC- and B[a]P-induced *CYP1A1* gene expression, CSC-induced CYP1 activity and levels of B[a]P-DNA adducts in B[a]P-exposed cells were found in cell lines from women compared to cell lines from men. The results support the hypothesis that the female lung may be more susceptible to PAH than the male lung.

The last main focus was to examine the modulatory role of sex steroid receptors in regulation of *CYP1A1* and *CYP1B1* expression in human lung cells. Knockdown of gene expression was performed with siRNA, and gene expression was measured by RT-qPCR. Knockdown of estrogen receptor β (*ER β*) significantly reduced B[a]P-induced expression of *CYP1A1* and *CYP1B1*, whereas knockdown of androgen receptor (*AR*) significantly enhanced both CSC- and B[a]P-induced expression of *CYP1B1* in H2009 cells. ICI 182780, an antiestrogenic agent known to specifically degrade *ER α* , increased the expression of *CYP1A1* and *CYP1B1*, whereas dihydrotestosterone (DHT) repressed B[a]P-induced expression of both genes in H2009. Together the results may indicate that the presence of *ER β* may be necessary to maintain full B[a]P-induced *CYP1* expression, whereas the observed induction of *CYP1A1* and *CYP1B1* expression by ICI 182780 may indicate an opposite and thus suppressing role of *ER α* on *CYP1* expression. The observed effects inflicted on *CYP1* expression in *AR* knockdown- and DHT exposure experiments coincide, and implicate a common mechanism. The results indicate that the expression of sex steroid receptors and exposure to sex steroid receptor ligands influence regulation of basal and induced expression levels of *CYP1*.

The importance of CYP1A1 in bioactivation of B[a]P, and the relationship between expression levels of *CYP1A1* and elevated levels of B[a]P-DNA adducts in female cell lines compared to male cell lines, mutually strengthen one another. The present results also indicate a sex difference in PAH-bioactivation capacity of human lung adenocarcinoma cell lines, and together with previously published *in vivo* data from our group, they support that the female lung may be more sensitive to PAH-exposure than the male lung. The final study showed how knockdown of *ERβ* or *AR*, respectively, and exposure to sex steroid receptor ligands had a significant impact on expression levels of *CYP1* genes. These results support a role of sex steroid receptors in modulation of xenobiotic bioactivation in lung cells.

This thesis provides evidence indicating that female lung cells have higher PAH-bioactivation capacity than male lung cells, and that sex differences in expression of *CYP1A1* may be mechanistically involved in these differences. Our results also indicate that regulation of important PAH-bioactivation genes is modulated by expression of sex steroid receptors. Further investigations of underlying interactions between existing signaling pathways and novel mechanisms behind sex differences in lung cancer biology should be granted necessary attention.

Abbreviations and explanations

4-OHT;	4-hydroxytamoxifen
AHR;	aryl hydrocarbon receptor
AHRR;	aryl hydrocarbon receptor repressor
AI;	aromatase inhibitor
AKR;	aldo-keto reductase
AP-1;	activator protein-1
AR;	androgen receptor
ARE;	androgen response element
ARNT;	aryl hydrocarbon receptor nuclear translocator
BAG6;	B-cell CLL/lymphoma 2 associated athanogene 6
B[a]P;	benzo[<i>a</i>]pyrene
BPDE I;	B[<i>a</i>]P-7,8-dihydrodiol-9,10-epoxide I
BTE;	basic transcription element
CHRNA;	cholinergic receptor nicotinic alpha
ChIP;	chromatin immunoprecipitation
CLPT1L;	cleft lip and palate transmembrane protein 1-like protein
CSC;	cigarette smoke condensate
CYP;	cytochrome P450 monooxygenase
DEP;	diesel exhaust particle
DHT;	dihydrotestosterone
DNMT;	DNA methyltransferase
DPN;	diarylpropionitrile
E2;	17 β -estradiol
EGFR;	epidermal growth factor receptor
EH;	epoxide hydrolase
ERα;	estrogen receptor alpha
ERβ;	estrogen receptor beta
ERE;	estrogen response element
EROD assay;	ethoxyresorufin-O-deethylase assay
GWAS;	genome wide association study

HAH;	halogenated aromatic hydrocarbon
HBEC;	human bronchial epithelial cells
HDAC;	histone deacetylase
HPLC;	high-performance liquid chromatography
HRT;	hormone replacement therapy
HSP90;	heat shock protein 90
IARC;	International Agency for Research on Cancer
ICI 182780;	ER α antagonist (Fulvestrant, Faslodex)
<i>In vitro</i>;	outside a living organism, in an artificial environment
<i>In vivo</i>;	within a living organism
lncRNA;	long non-coding RNA
LOI;	loss of imprinting
miRNA;	micro RNA
mRNA;	messenger RNA
NNK;	4-(methylnitrosamino)-1-(3-pyridyl)-1-butanone
NSCLC;	non-small cell lung carcinoma
OHF;	hydroxyflutamide
p53;	tumor protein p53
PAH;	polycyclic aromatic hydrocarbon
ROS;	reactive oxygen species
RR;	relative risk
SERD;	selective estrogen receptor degrader
siRNA;	small interfering RNA
SNP;	single nucleotide polymorphism
Sp1;	stimulating protein 1
TCDD;	2,3,7,8-tetrachlorodibenzo- <i>p</i> -dioxin
TERT;	telomerase reverse transcriptase
TSG;	tumor suppressor gene
XAP2;	aryl hydrocarbon receptor interacting protein
XRE;	xenobiotic response element

1.Introduction

1.1 Lung cancer

1.1.1 Incidence, histology and mortality

Lung cancer is a global major health problem and the leading cause of cancer death across the world. Approximately 1.8 million new cases are diagnosed annually and current statistics reveal that lung cancer accounts for 12.9 % of all new cancers diagnosed, and 19.4 % of all cancer deaths registered (Ferlay *et al.* 2015, Fitzmaurice *et al.* 2015, Torre *et al.* 2016). Many developing countries experience a steep increase in lung cancer burden, in part due to rapid industrialization and increasing population growth (Hashim and Boffetta, 2014).

Typically, lung cancers are divided in two distinct histological types, which are further grouped into subcategories. The main histological types are non-small cell lung cancer (NSCLC) and small cell lung cancer (SCLC), with prevalence rates of 86 % (NSCLC) and 14 % (SCLC). NSCLC typically originate from bronchial epithelial-cell precursors and are divided into squamous cell carcinoma, adenocarcinoma and large cell carcinoma. The distribution of histological types has changed over the past decades. Thus, the incidence of squamous cell carcinomas has decreased, while incidence of adenocarcinomas exhibits an increase, especially in women (Torre *et al.* 2016). SCLC usually originate from neuroendocrine-cell precursors, and are subcategorized into small cell carcinoma and combined small cell carcinoma (Travis *et al.* 2013).

Cumulative exposure models estimate that the average latency period for lung cancer is approximately 30 years, but few patients receive their diagnosis at an early stage of their disease. In fact, approximately 50 % of the diagnosed cases are classified as advanced or metastatic (Quint, 2007). Unfortunately, the expected overall 5-year survival rate is 16 % for lung cancer patients, and the need for sensitive and specific diagnostic tools, as well as novel treatments to combat the disease is urgent (Ferlay *et al.* 2015; Sharma *et al.* 2016). An essential component required to provide these tools, is the increased understanding of underlying molecular mechanisms in lung carcinogenesis.

1.1.2 Lung cancer etiology and lung carcinogens

An immense amount of evidence is reaffirming tobacco smoking as the major cause of lung cancer. Globally, 85 % of lung cancers in men and 47 % of lung cancers in women are designated the act of tobacco smoking (Youlden *et al.* 2008). Moreover, the establishment of a causal association between secondhand tobacco smoke and lung cancer is firmly founded, and expected to account for 1.6 % of all lung cancers diagnosed (Boffetta, 2006). The aerosol emerging from the mouthpiece of a cigarette is composed of approximately 4,800 different chemical compounds and typically contains 1,000 particles/ml. Among these compounds are known carcinogens such as polycyclic aromatic hydrocarbons (PAHs), tobacco-specific nitrosamines, 1,3-butadiene, ethyl carbamate, ethylene oxide, nickel, chromium, cadmium, polonium-210, arsenic, and hydrazine (Hecht, 1999).

Occupational exposures are responsible for a considerable fraction of lung cancer incidences (Hashim and Boffetta, 2014). The most important occupational lung carcinogens include mixtures of PAHs, heavy metals, radon, crystalline silica and asbestos. More specifically, occupations such as welding and painting are associated with increased risk of lung cancer. Additionally, uranium miners and nuclear plant workers also have an increased risk of lung cancer due to the exposure to radioactive particulate mass (Algranti *et al.* 2010). It is also worth mentioning, that for most known occupational carcinogens, synergisms with tobacco smoking has been reported (IARC Monographs Vol 83, 2004).

Asbestos are mineral fibers considered to be strong lung carcinogens, and a serious occupational health hazard worldwide. Today, the use of asbestos is in most countries either abandoned or controlled, but it still poses a significant public health threat (Lin *et al.* 2007). Asbestos can act on its own, or synergistically with tobacco smoke to induce lung cancer (Markowitz, 2015). For individuals exposed to asbestos, the relative risk of lung cancer is three times higher than for non-exposed individuals (van Loon *et al.* 1997). Mechanistically, asbestos-induced carcinogenesis is considered to occur through the creation of chronic inflammation, involving oxidative stress and induction of DNA damage (Benedetti *et al.* 2015).

Cumulative exposure to emissions rich in various PAHs present in urban air poses an environmental lung cancer risk (Vineis and Husgafvel-Pursiainen, 2005). Urban air is typically polluted by vehicle emissions such as diesel exhaust particles (DEP) and high concentrations of carcinogenic PAH may be adsorbed on these particles (Ono-Ogasawara and Smith, 2004). DEP are linked to an elevated risk of lung cancer through mechanisms involving oxidative stress and oxidation of DNA (Hesterberg *et al.* 2012; Risom *et al.* 2005). The proportion of lung cancers attributable to urban air pollution in Europe is estimated to be 11 % (Boffetta, 2006).

A considerable fraction of environmental exposure to lung carcinogens comes from arsenic in drinking water and food (Martinez *et al.* 2011). More than 200 million people live in regions with naturally elevated levels of arsenic in their drinking water (Naujokas. *et al.* 2013), and the main cause of death after chronic arsenic intake is lung cancer (Hubaux *et al.* 2012). Arsenic exposure contributes synergistically with other lung carcinogens, such as exposure to tobacco smoke (Ferreccio *et al.* 2013).

Another highly relevant lung carcinogen is radon, a naturally occurring gas usually found in igneous rock and soil and originating from the decay of uranium. Radon itself subsequently decays further to generate radioactive alpha and beta emitting particles, which may damage DNA both directly and through generation of free radicals (Ruano-Ravina *et al.* 2009; Samet *et al.* 2009). Radon is a Class I human carcinogen (IARC Monographs Vol 43, 1988) and studies have shown that domestic and/or occupational inhalation exposure can cause lung cancer (Brauner *et al.* 2012). In the case of non-occupationally exposed never-smokers, studies have revealed that 1/3 of lung cancer deaths might be linked to indoor radon (Lubin *et al.* 1995).

1.1.3 Lung cancer epidemiology and smoking habits are tightly linked

Smoking behavior is the most influential factor in risk of developing lung cancer. It reflects on the trends in lung cancer incidence as well as mortality (IARC Monographs Vol 83, 2004). The first so-called modern cigarette was presented in the early 20th century, and its arrival led to a steep increase in tobacco use, in particular among male inhabitants of the Western world. In the

1940's, women also began their tobacco smoking habits. The trend in the United States shows that the overall habit of cigarette smoking has peaked and declined (Figure 1).

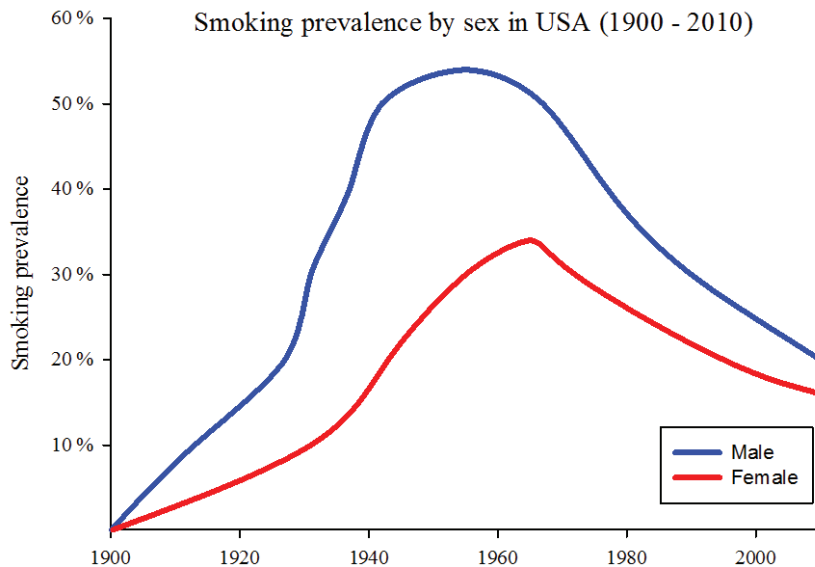


Figure 1. Smoking prevalence by sex in USA (1900 - 2010). Modified from (Thun *et al.* 2012).

Although authorities all over the world have focused on tobacco control issues involving education on the harmful effects of smoking, along with tax increases and smoking restrictions, smokers still exist. Unfortunately, the global image highlights the fact that many developing countries are experiencing a considerable increase in tobacco use following a relative increase in wealth among the inhabitants (O'Connor *et al.* 2010). It is projected that if existing trends remain unchanged, many countries will not manage to reach their goals concerning tobacco control and it is therefore estimated that approximately 1 billion people will remain current smokers in year 2025 (Bilano *et al.* 2015). On the bright side, many developed countries have succeeded in reducing the adult smoking prevalence to less than 25 % (Ng *et al.* 2014). Statistics Norway (SSB) reports that in 2016 only 13 % of inhabitants in Norway smoke on a daily basis (Statistics Norway, Smoking Habits, 2016).

Despite a steady decline in smoking prevalence in recent decades primarily in developed countries (Bray *et al.* 2002), one can spot significant differences in the prevalence among different groups of inhabitants. Education, occupational status and income are all inversely

associated with smoking (Laaksonen *et al.* 2005). Another important demographic feature is that the epidemic most likely will subsist among women in some high-income countries as part of their self-realization process (Bilano *et al.* 2015). In the United States, almost all female smokers started their consumption of cigarettes in their teens, and even though they are well aware of the harmful effects, smoking has become a way to express independence and equality with their grown-up counterparts. The tobacco companies execute extensive marketing toward young girls for example by developing cigarette brands specifically designed for women (Jemal *et al.* 2008).

1.2 Lung carcinogenesis

1.2.1 Stages in carcinogenesis

Continuing discoveries in cancer research constantly renew the knowledge on the complex networks of biological mechanisms involved in carcinogenesis. Hanahan and Weinberg proposed six hallmarks of cancer, which comprise the most important biological capabilities acquired during the multistep development of human tumors (Hanahan and Weinberg, 2000). In 2011, these authors added two more emerging hallmarks of cancer, altogether comprising; sustaining proliferative signaling, evading growth suppressors, resisting cell death, enabling replicative immortality, inducing angiogenesis, activating invasion and metastasis, reprogramming of energy metabolism and evading immune destruction (Hanahan and Weinberg, 2011).

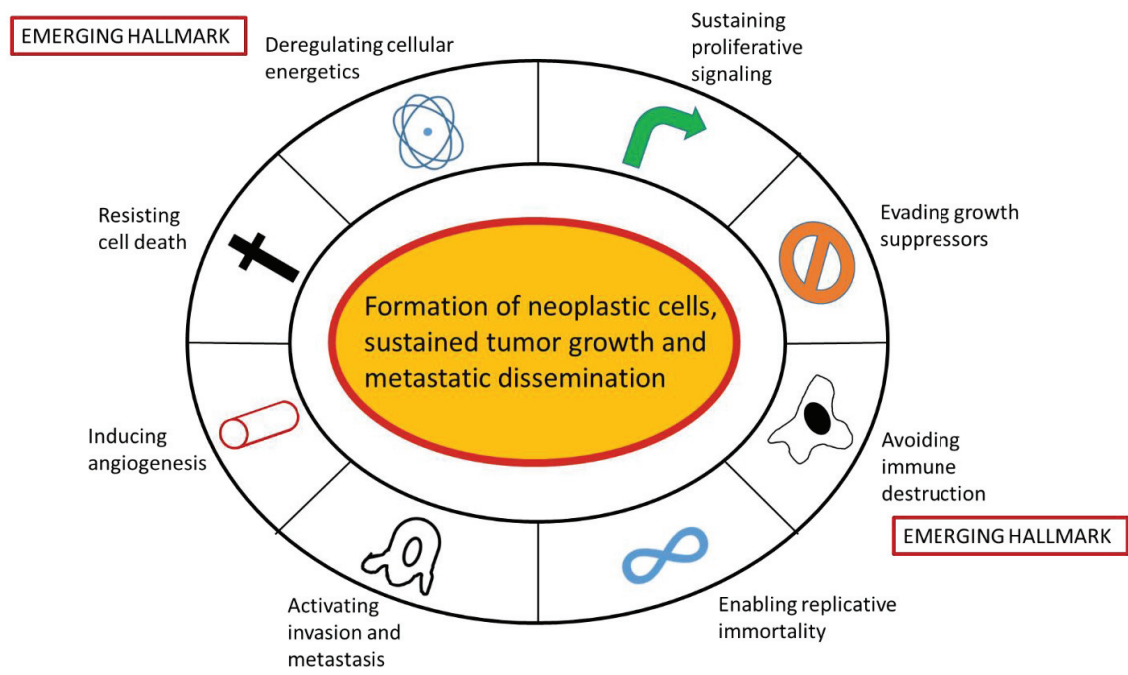


Figure 2. Eight hallmark capabilities of tumor pathogenesis. Modified from (Hanahan and Weinberg, 2011).

A historical and less detailed conceptual model, explicitly described by C.C. Harris in 1991, focus on three main stages of carcinogenesis defined as initiation, promotion and progression (Harris, 1991). The initiation stage of carcinogenesis is characterized by irreversible genetic changes, which predispose susceptible normal cells to impending malignant transformation. An initiated cell is not neoplastic, but it has acquired several successive genotypical and phenotypical changes. Such changes may typically be mutations in DNA-repair genes, oncogenes and tumor-suppressor genes. Cell proliferation and inhibition of apoptosis are essential for this stage, as they both contribute to clonal expansion of initiated cells.

The promotion stage is characterized by increased cell proliferation in susceptible tissues. The increased proliferation will in turn contribute to a fixation of mutations that may enhance alterations in genetic expression (Ochieng *et al.* 2015). Promotion changes DNA in both genetic and epigenetic ways (Hanahan and Weinberg, 2000). In contrast to initiation, promotion is a potentially reversible stage. Representative agents acting in the promotion stage of lung carcinogenesis mostly belong to chemical categories like reactive oxygen species (ROS), redox

active xenobiotics/metals, phorbol esters, polycyclic aromatic compounds, peroxisome proliferators and endocrine disruptors (Hanahan and Weinberg, 2000).

Progression is the last stage of carcinogenesis, and here the neoplastic phenotype is acquired through both genetic and epigenetic mechanisms (Oliveira *et al.* 2007). Progression is irreversible and brings genetic instability, uncontrolled growth, invasion, metastization and altered biochemistry to the affected cells (Oliveira *et al.* 2007; Pitot and Dragan, 1991). In this stage, cell proliferation operates independently from surrounding stimuli (Lutz, 2000). Angiogenesis is essential for progression, although the attainment of an angiogenic phenotype has been found to precede the development of malignancy (Hawighorst, 2001). Agents involved in progression of lung carcinogenesis include both genotoxic and non-genotoxic, promoting compounds.

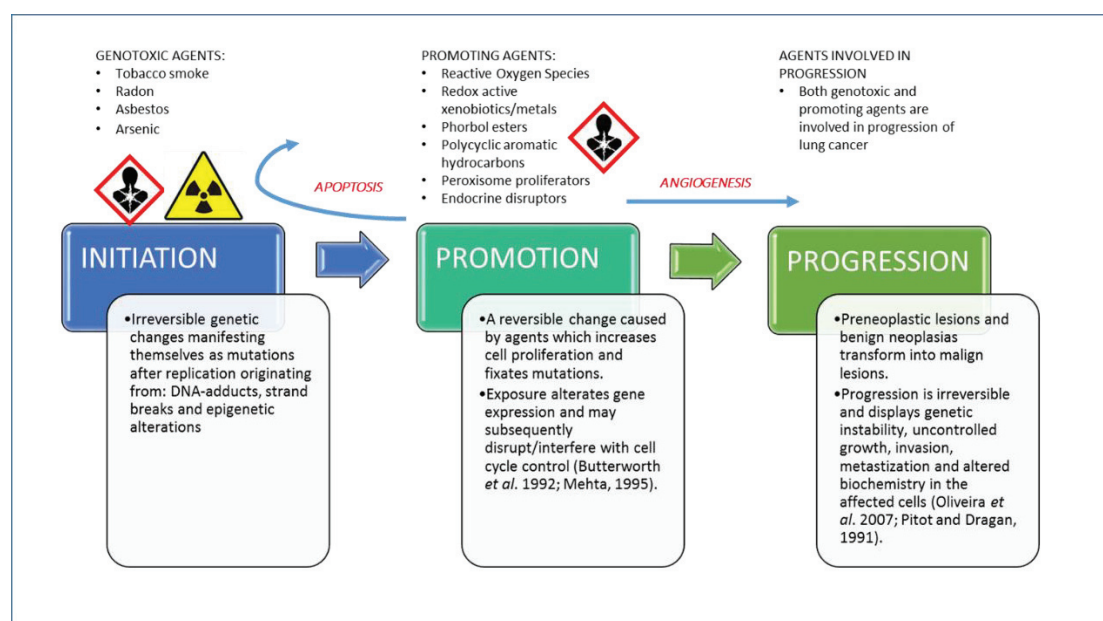


Figure 3. Chemical carcinogenesis divided into three steps. Modified from (Harris, 1991; Oliveira *et al.* 2007).

1.2.2 Lung cancer susceptibility

It has long been recognized that people differ in their susceptibility toward disease. Although different individuals may experience similar environmental and/or occupational exposures, time- and dose-wise, they do not necessarily have the same risk of developing a disease. Susceptibility usually appears as a consequence of genetic predisposition, including mutations, single nucleotide polymorphisms (SNPs) and epigenetic alterations. Their main features are their influence on gene expression or genetic stability. Epidemiological data have revealed that some individuals are predisposed to lung cancer, and susceptibility toward tobacco smoke carcinogens has been debated (Perera and Weinstein, 1982). Epidemiological and pedigree studies suggest that lung carcinogenesis in smokers depend on the combined effects of age, smoking, impaired lung function and genetic factors (Young *et al.* 2009). Eighty-five – 90 % of all NSCLC patients are smokers or former smokers, although only 15 % of lifetime smokers develop NSCLC (Villeneuve and Mao, 1994). Many genes can interfere with the carcinogenic potential of tobacco smoke and the most prominent susceptibility genes are typically involved in carcinogen-metabolism, DNA-repair and cell cycle control (Spitz *et al.* 2003).

1.2.3 Genetic susceptibility to lung cancer

Recent large scale multi-cohort genome wide association studies (GWAS) have revealed several chromosomal regions as lung cancer susceptible loci. In Caucasian smoking populations, the chromosomal regions 15q25, 5p15 and 6p21 have been reported to harbor genes associated with elevated risk of lung cancer (Wei *et al.* 2015; Yang *et al.* 2013). In never smoking females, however, other specific chromosomal regions were highlighted as susceptibility loci (Wei *et al.* 2015; Yang *et al.* 2013).

15q25 contains neuronal nicotinic acetylcholine receptor (*nAChr*) subunits (cholinergic receptor, nicotinic alpha 3 and 5) *CHRNA3* and *CHRNA5* genes. SNPs in these genes can modulate lung cancer risk indirectly through smoking behavior, or through the direct anti-apoptotic or proliferative effects of nicotine (Marshall and Christiani, 2013).

SNPs in cleft lip and palate transmembrane protein 1-like protein (*CLPTLIL*) and telomerase reverse transcriptase (*TERT*) on locus 5p15 have been reported to increase the risk of lung cancer and other cancers. The *CLPTLIL* gene is presumed to have an essential role regarding apoptosis in lung cells (McKay *et al.* 2008). *TERT* is important in maintenance of telomere length, and short telomere length has been associated with a significantly increased risk of lung cancer (Ma *et al.* 2011). SNPs in telomere maintenance genes such as *TERT* are therefore considered to play a role in lung cancer susceptibility (Choi *et al.* 2009; Hosgood *et al.* 2009).

6p21 harbors two candidate genes associated with elevated lung cancer risk. *BCL2*- associated anthanogene 6 (*BAG6*) is involved in induction of apoptosis subsequent to DNA-damage and modulating the response of p53 upon genotoxic stress (Marshall and Christiani, 2013), while the *E. coli* mutS homolog 5, *MSH5* participates in DNA mismatch repair (Yang *et al.* 2013).

1.2.4 Epigenetic alterations

At present, epigenetic alterations are considered as important as genetic changes in carcinogenesis. Among the epigenetic mechanisms involved in lung cancer are changes in DNA methylation, histone modification, nucleosome repositioning, and altered regulation by noncoding RNAs (miRNA, lncRNA) (Robertson, 2005).

Methylation of DNA and acetylation of histones influence gene expression by regulating chromatin structure and accessibility for transcription factors. Different mechanisms of DNA hypomethylation can result in lung carcinogenesis. The first one involve either retrotransposons or activation of repetitive elements (SINE or LINE) with subsequent microsatellite instability. The second mechanism is transcriptional activation and overexpression of oncogenes, and the third is through the loss of imprinting (LOI) (Brena and Costello, 2007; Ehrlich, 2002; Herman and Baylin, 2003). LOI is important in carcinogenesis because it can result in dual allele activation and increased gene expression of oncogenes. LOI can also result in lack of expression of tumor-suppressor genes (TSGs) and DNA repair genes due to lost imprinting and thus increased accessibility to responsive elements that suppress normal transcription (Jeclinic and Shaw, 2007). In conjunction with changes in DNA methylation, histone modifications such as

acetylation and deacetylation may influence gene expression, DNA replication, DNA repair and DNA recombination, and thus lead to cancer (Glozak and Seto, 2007; Kouzarides, 2007).

DNA may become modified through hydroxymethylation of cytosine. Tet methylcytosine dioxygenases typically catalyze the conversion of 5-methylcytosine to 5-hydroxymethylcytosine, and this is believed to be an intermediate state leading to hypometylation (Guibert and Weber, 2013). Chemical exposure to metals, persistent organic pollutants and PAH, can activate Tet methylcytosine dioxygenases, and indirectly contribute to hypometylation of DNA (Ruiz-Hernandez *et al.* 2015).

Furthermore, target mRNAs can be repressed or degraded by ncRNA, such as miRNA. miRNA can control a wide variety of cellular processes such as proliferation and apoptosis which often show irregularities in human cancer (Calin and Croce, 2006; Motoyama *et al.* 2009; Vasudevan *et al.* 2007), and several miRNAs become silenced by CpG hypermethylation in cancer cells (Bueno *et al.* 2008). This system shows great complexity, since one mRNA can be the target of many miRNA's, and one miRNA can target many mRNA's.

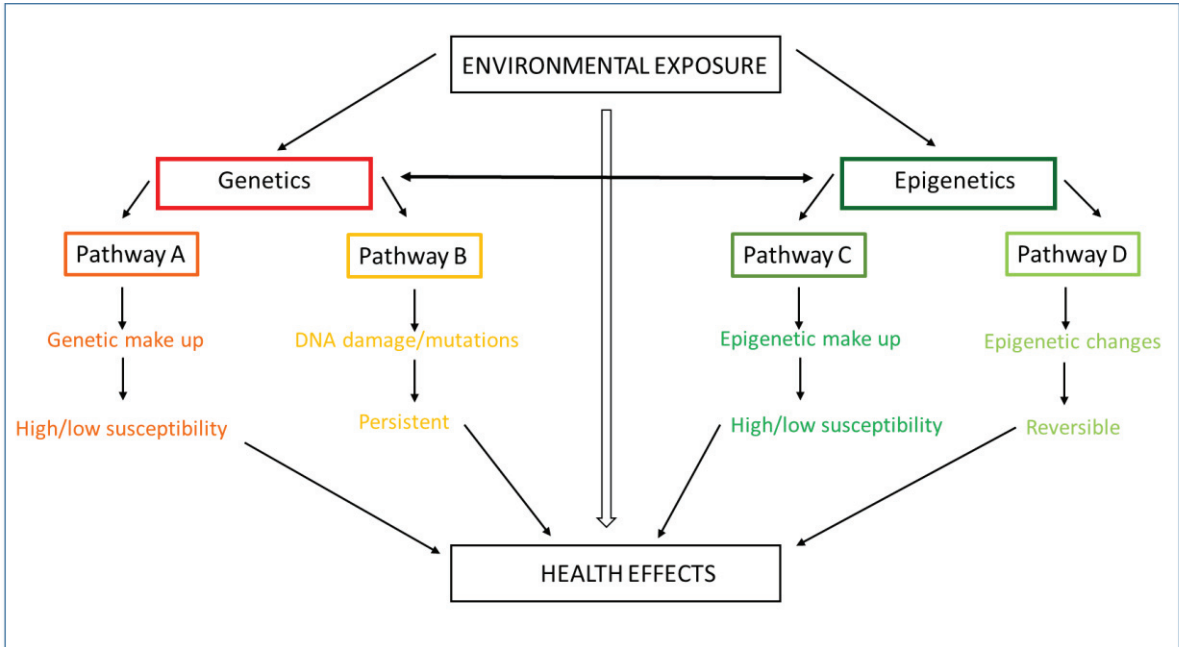


Figure 4. Potential genetic and epigenetic pathways linking environmental exposure to health effects. Modified from (Bollati and Baccarelli, 2010).

1.2.5 Epigenetics in lung cancer

Recent data suggest that smoking directly alters and causes aberrant expression of epigenetic modulators and their downstream targets during initiation and progression of lung cancer (D'Alessio and Szyf, 2006; Izzotti *et al.* 2009). Furthermore, in a study by Watanabe *et al.* (2012) fifty-five miRNA were reported epigenetically silenced in NSCLC patients with a smoking history (Watanabe *et al.* 2012).

Several recent studies suggest that epigenetic changes detected in sputum, bronchoalveolar lavage and saliva may serve as biomarkers for lung cancer (Langevin *et al.* 2015). A growing number of hypermethylated TSG promoters and single gene hypomethylations that develop during early stages of lung carcinogenesis have been identified (Brzezianska *et al.* 2013). Detection of such alterations using non-invasive diagnostic tools could contribute to earlier detection of lung cancer (Langevin *et al.* 2015).

Epigenetic alterations are not only important in diagnostics, but also as therapeutic targets in lung cancer. Considering treatment, histone deacetylase- (HDAC) and DNA methyltransferase- (DNMT) inhibitors could principally restore TSG expression in lung cancer patients. In the clinic, both types of drugs have been tested in treatment of NSCLC, however none of them are currently approved (Lawless *et al.* 2009; NIH, National Cancer Institute, cancer drugs approved by the Food and Drug Administration (FDA) for lung cancer, 2016).

1.2.6 Lung cancer and sex differences

Trend analyses show that lung cancer still is the most common cancer in men worldwide. Globally, approximately 1.2 and 0.6 million new cases of lung cancer are diagnosed in males and females each year, respectively. The incidence rate in women is thus generally lower, but nonetheless, lung cancer is the fourth most frequent cancer and the most common cause of death from cancer among women in developed regions (Bray *et al.* 2013; Ferlay *et al.* 2015; Patel, 2005).

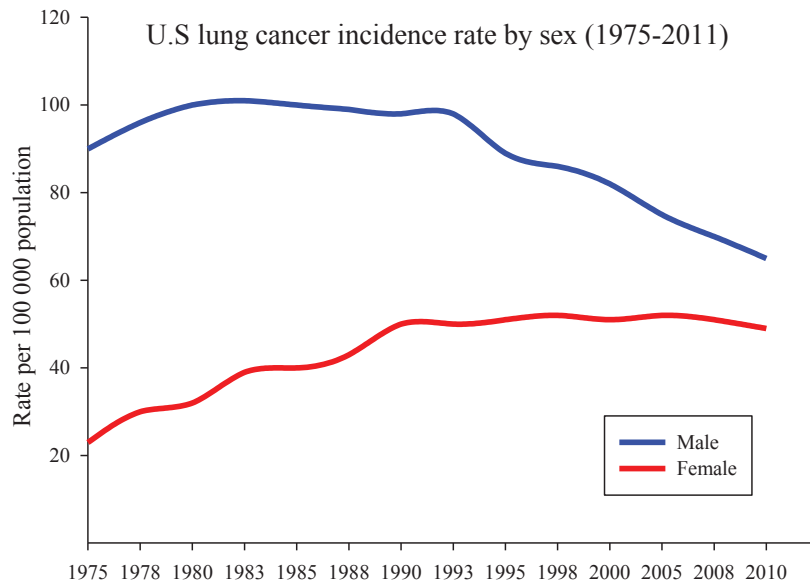


Figure 5. Lung cancer incidence rate by sex in USA 1975 – 2011. Modified from (National Institutes of Health, National Cancer Institute, U.S. SEER Cancer Statistics Review, 1975-2011, 2014).

It has been debated for several decades whether there are sex differences in lung cancer risk. Several case-control studies have shown that women could be at greater risk of developing lung cancer from cigarette smoking than men (Novello and Baldini, 2006; Patel *et al.* 2004; Ramchandran and Patel, 2009). A recent study of patients diagnosed with lung adenocarcinoma, concluded that female smokers were significantly younger than their male counterparts regarding onset of the disease (Nagy-Mignotte *et al.* 2011). Another recent report showed that women were overrepresented amongst NSCLC patients under 40 years (Thomas *et al.* 2015). Furthermore, an elevated incidence of lung cancer in female never-smokers compared to male never-smokers has been described (Koo and Ho, 1990; Stockwell *et al.* 1990; Wakelee *et al.* 2007) which at least in some parts of the world may be associated with exposure to domestic cooking oil- and coal vapors (Gao *et al.* 1987).

In contrast, prospective studies have not been as conclusive regarding sex differences in risk of developing lung cancer (Bain *et al.* 2004; Henscke *et al.* 2006), and a study by De Matteis *et al.* (2013) supports this notion and reports no significant sex difference in risk of lung cancer (De Matteis *et al.* 2013). Discussions regarding study designs, risk estimate measures, model of interactions and potential confounders (e.g., depth of inhalation, tobacco type) that are

preferable or should be taken into consideration, still withstand (De Matteis *et al.* 2013), and the matter of sex differences in risk of lung cancer remains controversial.

A number of hypothesis regarding underlying reasons for a possible sex difference in human lung cancer risk have been proposed. One suggestion is that the combination of exposure to environmental risk factors and genetic differences may account for unequal RR for developing lung cancer (Kiyohara and Ohno, 2010; Kreuzer *et al.* 2002; Siegfried, 2001; Stabile and Siegfried 2003). Other possible explanations have focused on under-reporting of smoking habits and a potentially greater exposure to passive smoking among women (Haugen, 2002; Patel, 2005;).

Although increased risk of lung cancer among women is debatable, women appear to have improved survival rates in every stage of lung cancer, with women over the age of 60 years having a survival advantage over men and younger women (Nakamura *et al.* 2011; Patel *et al.* 2004). Additionally, women are more likely than men to be diagnosed with adenocarcinoma compared with other histological lung cancer subtypes, and in Asian populations, women with lung adenocarcinoma seem to have a higher incidence of tumors with *EGFR* mutations. These lung tumors are more responsive to *EGFR* tyrosine kinase inhibitors, and this may account for a significant fraction of the survival advantage observed at least in Asian women (Marquez-Garban *et al.* 2007; Mitsudomi, 2014). To sum up, epidemiological data indicate that lung tumor biology differs between males and females; however, the matter of sex differences in risk of lung cancer is still not completely sorted out. This is an important area of research, which would benefit from greater dedication.

1.2.7 Animal studies and sex differences in lung cancer biology

Sex differences in lung cancer risk are supported by animal studies. One study reported lower lung tumor incidence and reduced tumor size in androgen receptor (*Ar*) knockout mice co-exposed to 4-(methylnitrosamino)-1-(3-pyridyl)-1-butanone (NNK) and benzo[*a*]pyrene (B[*a*]P) compared to wild-type mice receiving the same treatment (Yeh *et al.* 2012). Moreover, female A/J mice were confirmed to be more susceptible to NNK-induced lung carcinogenesis than male mice. In males, lung carcinogenesis was increased by castration, whereas in females,

malignant transformation of lung proliferative lesions tended to be inhibited by ovariectomy. These results suggested that NNK-induced lung carcinogenesis is inhibited by testosterone and accelerated by E2 (Ninomiya *et al.* 2013), confirming that female A/J mice are more susceptible to NNK-induced lung carcinogenesis than males. These findings indicate the possibility that sex hormones play important roles in determining sex differences in lung carcinogenesis in the A/J mice initiated by NNK (Ninomiya *et al.* 2013).

1.3 Biotransformation of PAH in tobacco smoke

1.3.1 Chemistry, sources and biotransformation of PAH

PAHs are formed by incomplete combustion of organic material, and typical sources comprise urban and industrial air pollution, tobacco smoke and diet (Bostrom *et al.* 2002). The most prominent PAH exposure routes for humans are inhalation, digestion and skin absorption (Bostrom *et al.* 2002). Most PAHs are nonpolar organic molecules consisting of two or more benzene rings arranged in different configurations. Most PAHs are insoluble in water, and their aqueous solubility decreases logarithmically as the molecular mass increases. Consequently, PAHs tend to persist in the environment (Johnsen *et al.* 2005).

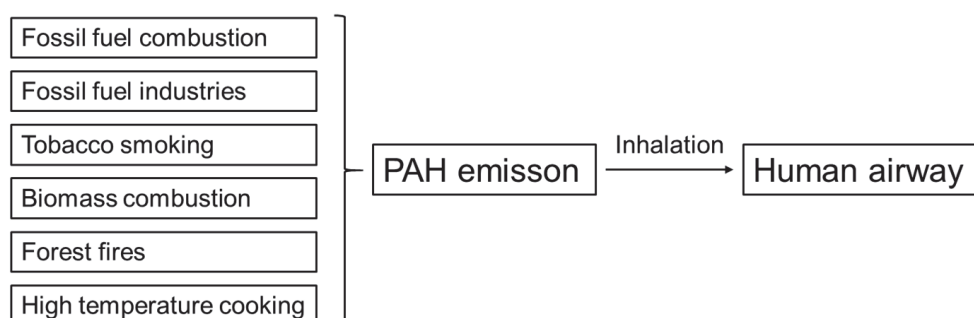


Fig. 6. Main sources of airway PAH-exposure in humans (Bostrom *et al.* 2002).

Airway-exposure to PAH increases the risk of lung cancer in humans, and IARC has listed thirteen PAHs as human carcinogens or potential carcinogens, comprising benz[*a*]anthracene, benzo[*b*]fluoranthene, benzo[*j*]fluoranthene, B[*a*]P, dibenz[*a,h*]anthracene, 7H-dibenzo[*c,g*]carbazole, dibenzo[*a,h*]pyrene, dibenzo[*a,i*]pyrene, indeno[*1,2,3-cd*]pyrene,

benzo[*k*]fluoranthene, dibenzo[*a,e*]pyrene, dibenzo[*a,l*]pyrene and 5-methylchrysene (IARC Monographs Vol 32, 1983).

Tobacco smoke is a significant source of PAH. Cigarettes with regular tar content typically contain approximately 100 ng of total PAHs per gram of tobacco, and a smoker may thus daily inhale ~0.26 µg of B[*a*]P per pack of 20 cigarettes (Grimmer *et al.* 1988; Piccardo *et al.* 2010).

PAHs can diffuse passively across cell membranes due to their lipophilic nature. After entering the cells they usually remain as inert procarcinogens unless bioactivated by endogenous enzymes. Biotransformation of PAH encompasses several metabolic enzymes and includes at least three different pathways. Prominent enzymes present in the first pathway are cytochrome P450 1A1 (CYP1A1), CYP1B1 and epoxide hydrolase (EH) (Shimada and Fujii-Kuriyama, 2004). The second pathway is referred to as the CYP-peroxidase pathway and is catalyzed by a completely different battery of enzymes (Moorthy *et al.* 2015), while the third pathway is called the aldo-keto reductase pathway and the main enzymes involved are aldo-keto reductase 1A1 (AKR1A1) and AKR1C1-1C4 (Palackal *et al.* 2001, 2002).

B[*a*]P is the most extensively studied PAH, and it often serves as a reference for the carcinogenic potential of other PAHs (Bostrom *et al.* 2002). B[*a*]P has the prospective of being bioactivated to the ultimate carcinogen B[*a*]P-diolepoxide I (BPDE I). BPDE I binds to DNA and forms predominantly covalent (+) *trans* adducts at the N2 position of guanine, and strong evidence from molecular epidemiologic studies show that lung tissue in smokers have significantly higher PAH–DNA adduct levels than lung tissue in non-smokers (Dipple, 1995; Perera *et al.* 1987; Wolfe *et al.* 2004).

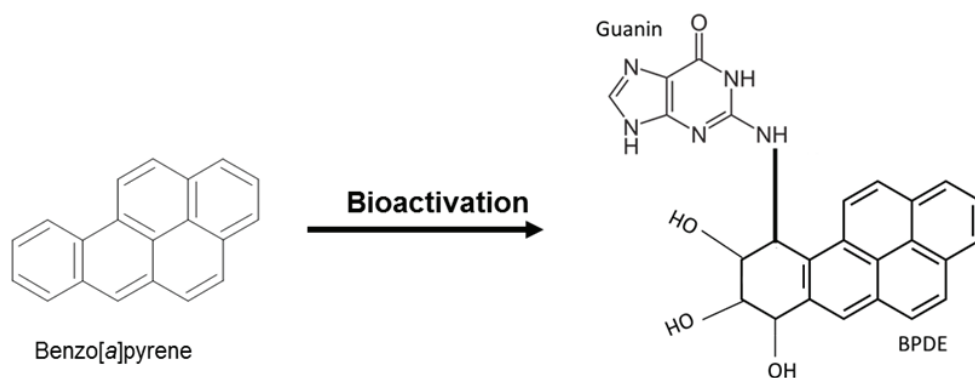


Figure 7. Bioactivated B[a]P forms BPDE-DNA adduct. Modified from (Hecht, 1999; Poirier, 2004).

PAHs exhibit great variability regarding their carcinogenic potential (Deutsch-Wenzel *et al.* 1983; Grimmer *et al.* 1988). Furthermore, exposure often includes mixtures of several different PAHs. In a study conducted by Tarantini *et al.* (2011) different compounds present in PAH-mixtures were shown to affect each other's potential carcinogenicity both synergistically and antagonistically, making the evaluation of cancer risk for PAHs difficult to establish (Tarantini *et al.* 2011).

1.3.2 Aryl hydrocarbon receptor (AHR)

The AHR is a Class E basic helix-loop-helix protein with the highest expression found in liver, adipose tissue, and bronchial epithelial cells (Tsay *et al.* 2013). The human gene encoding the AHR is localized to chromosome position 7p15, and the full-length protein consists of 848 amino acids and weighs 96147 Da. The mRNA sequence corresponding to the full-length protein consists of 11 exons and at least two additional splice variants have been identified (National Center for Biotechnology Information, PubMed Gene ID: 196, 2016)

The AHR is a transcription factor normally found in a cytosolic inactive protein complex comprising a heat shock protein 90 (HSP 90) dimer, co-chaperone p23 and hepatitis B virus X-associated protein 2 (XAP2). When hydrophobic ligands such as PAHs enter the cell by simple diffusion, they can readily bind to the receptor. Ligand binding results in phosphorylation of two protein kinase C sites on the AHR adjacent to nuclear localization sequences. Subsequently, the AHR complex translocates to the nucleus where the receptor dissociates from its chaperone complex and dimerizes with the AHR nuclear translocator (ARNT). The AHR/ARNT

heterodimer is the active transcription factor able to recognize xenobiotic response elements (XREs) in promoter regions of *AHR* responsive genes and thereby alter their expression levels (Esser and Rannug, 2015).

AHR has many physiological effects in the lungs, including altered cell proliferation, differentiation, cell-cell adhesion and xenobiotic metabolism (Chiba *et al.* 2011; Wong *et al.* 2010). Furthermore, recent studies have implicated *AHR* in the development of lung cancer (Tsay *et al.* 2013) *AHR* is highly expressed in lung cancer patients, and AHR-agonists induce lung cancer cell growth through activation of multiple pathways (Chuang *et al.* 2012; Portal-Nunez *et al.* 2012; Wang *et al.* 2009). Overall, activation of AHR may cause a number of downstream effects, which influence tumorigenesis, inflammation, formation of DNA-adducts and cell proliferation (Tsay *et al.* 2013). Some of the genes regulated by the AHR are cytochrome P450 (CYP) *1A1*, *CYP1A2*, and *CYP1B1* (Shimada and Fujii-Kuriyama, 2004). However, the extent of AHR/ARNT-activation is both tissue-specific and dependent upon co-regulators present in different cell types (Whitlock, 1999).

It is apparent, however, that the AHR binds not only toxicants like PAHs, but also endogenous compounds, dietary flavonoids, phytochemicals and many pharmaceuticals (Denison and Nagy, 2003; Esser and Rannug, 2015; Safe *et al.* 2013). Consequently, it has been proposed that this receptor in fact may prove to be an important drug target (Safe *et al.* 2013).

1.3.3 Cytochrome P450 1A1 and 1B1 (CYP1A1 and CYP1B1)

The CYP1A1 and CYP1B1 enzymes belong to the cytochrome P450-superfamily, which catalyzes a variety of reactions, including metabolism of xenobiotics and biosynthesis of endogenous compounds essential to the cells. CYP1A1 and CYP1B1 catalyze the incorporation of one atom of molecular oxygen into the substrate and they typically convert hydrophobic compounds to more polar metabolites that may be readily excreted (Hukkanen *et al.* 2002).

The *CYP1A1* gene is located on chromosome 15q24.1, and the full-length protein consists of 512 amino acids and weighs 58165 Da. The mRNA sequence corresponding to the full-length

variant of the protein consists of seven exons. At least two additional splice variants of the protein have been identified in human leukocytes (Bauer *et al.* 2007; National Center for Biotechnology Information, PubMed Gene ID: 1543, 2016). The *CYP1B1* gene contains three exons and is located on chromosome 2p22.2. This protein consists of 543 amino acids and has a molecular mass of 60846 Da. Four transcript variants of *CYP1B1* have been identified. (National Center for Biotechnology Information, PubMed Gene ID: 1545, 2016).

CYP1A1 and *CYP1B1* are mainly expressed in extrahepatic organs. In human lung, the enzymes are highly expressed in alveolar type I and II cells, ciliated columnar bronchoalveolar epithelial cells, and alveolar macrophages (Hukkanen *et al.* 2002). Both enzymes typically localizes to the endoplasmic reticulum. *CYP1A1* is highly inducible by PAHs and halogenated aromatic hydrocarbons (HAHs) via AHR-mediated gene transcription. *CYP1A1* metabolizes PAHs, but also endogenous compounds such as arachidonic acid, eicosapentaenoic acid and bilirubin are metabolized by *CYP1A1* (National Center for Biotechnology Information, PubMed Gene ID: 1543, 2016; Rifkind, 2006; Shrenk, 1998). *CYP1B1* expression is also highly inducible by PAHs through the AHR, and in addition to its role in metabolism of several procarcinogens such as PAH, the *CYP1B1* enzyme is central in the bioactivation of 17 β -estradiol to 4-hydroxyestradiol (National Center for Biotechnology Information, PubMed Gene ID: 1545, 2016; Shimada and Guengerich, 2006). This catechol is considered important in breast and uterine carcinogenesis. Quinone intermediates derived from 4-hydroxyestradiol may react with DNA and form highly mutagenic adducts (Cavalieri *et al.* 2000).

Several studies have shown that *CYP1A1* and *CYP1B1* are responsible for bioactivation of most carcinogenic PAHs to epoxide intermediates (Shimada and Fujii-Kuriyama, 2004). *CYP1A1* and *CYP1B1* show similar, but not identical, substrate specificity regarding various PAHs (Nebert *et al.* 2004; Shimada *et al.* 2001). Analyses of human lung tissue have showed significantly higher expression of *CYP1A1* in both smokers and ex-smokers compared to never-smokers. Similar results were found in cells from bronchoalveolar lavage and bronchial biopsies in smokers versus non-smokers (Mollerup *et al.* 2006; Thum *et al.* 2006). *CYP1B1* expression in human lung displays a similar pattern (Kim *et al.* 2004). The expression of *CYP1A1* is associated with increased lung cancer risk (London *et al.* 2000; San Jose *et al.* 2010; Vineis *et al.* 2003; Yang *et al.* 2004) and *CYP1B1* is often overexpressed in tumor tissues, and

is therefore be considered to be a potential histopathological tumor marker (McFadyen and Murray, 2005).

1.3.4 Metabolism of B[a]P catalyzed by CYP1A1 and CYP1B1

The lipophilic nature of PAHs initiate cellular enzymatic metabolism to enhance clearance and excretion of the compounds. Metabolism of the prototype PAH B[a]P catalyzed by CYP1A1 and CYP1B1 illustrates how procarcinogenic PAH may become bioactivated during this process (Shimada and Fujii-Kuriyama, 2004).

The major B[a]P-metabolites formed by CYP1A1 and CYP1B1 are B[a]P-4,5-epoxide, B[a]P-7,8-epoxide and B[a]P-9,10-epoxide. The next enzymatic step catalyzed by EH forms the respective dihydrodiols B[a]P-(-)-*trans*-4,5-dihydrodiol, B[a]P-(-)-*trans*-7,8-dihydrodiol, and B[a]P-(-)-*trans*-9,10-dihydrodiol. Five B[a]P-phenols; 1-OH-B[a]P, 3-OH-B[a]P, 6-OH-B[a]P, 7-OH-B[a]P, and 9-OH-B[a]P are frequently formed during metabolism of B[a]P, usually by spontaneous rearrangement of B[a]P-dihydrodiols. These phenols can further be converted to their corresponding quinones; the most common being B[a]P-1,6-quinone, B[a]P-3,6-quinone, and B[a]P-6,12-quinone (Gelboin, 1980). The primary B[a]P-epoxides formed can be conjugated with glutathione by glutathione-S-transferases, and B[a]P-phenols and dihydrodiols can be conjugated to water-soluble compounds by either sulfate- or glucuronide conjugation.

B[a]P-7,8-dihydrodiol and B[a]P-9,10-dihydrodiol have the ability to become activated to highly reactive dihydrodiol-epoxides, also catalyzed by CYP1A1 and CYP1B1. The epoxide groups formed are either *cis* or *trans*, some which will be hydrolyzed by EH to yield B[a]P-tetrols. If not hydrolyzed, B[a]P-7,8-dihydrodiol-9,10-epoxide and B[a]P-9,10-dihydrodiol-7,8-epoxide can readily react with protein or DNA and form adducts. The B[a]P-*trans*-7,8-dihydrodiol-9,10-epoxide I is often referred to as the ultimate B[a]P-carcinogen, abbreviated BPDE I. BPDE I is highly mutagenic due to the bay-region of the molecule. This region provides an area of steric hindrance for detoxifying enzymes and the epoxide is very susceptible to nucleophilic attack from DNA and proteins. The mutagenic potential of B[a]P-9,10-

dihydrodiol-7,8-epoxide III (BPDE III) is low compared to BPDE I (Conney, 1980; Gelboin, 1980; Wei *et al.* 1991).

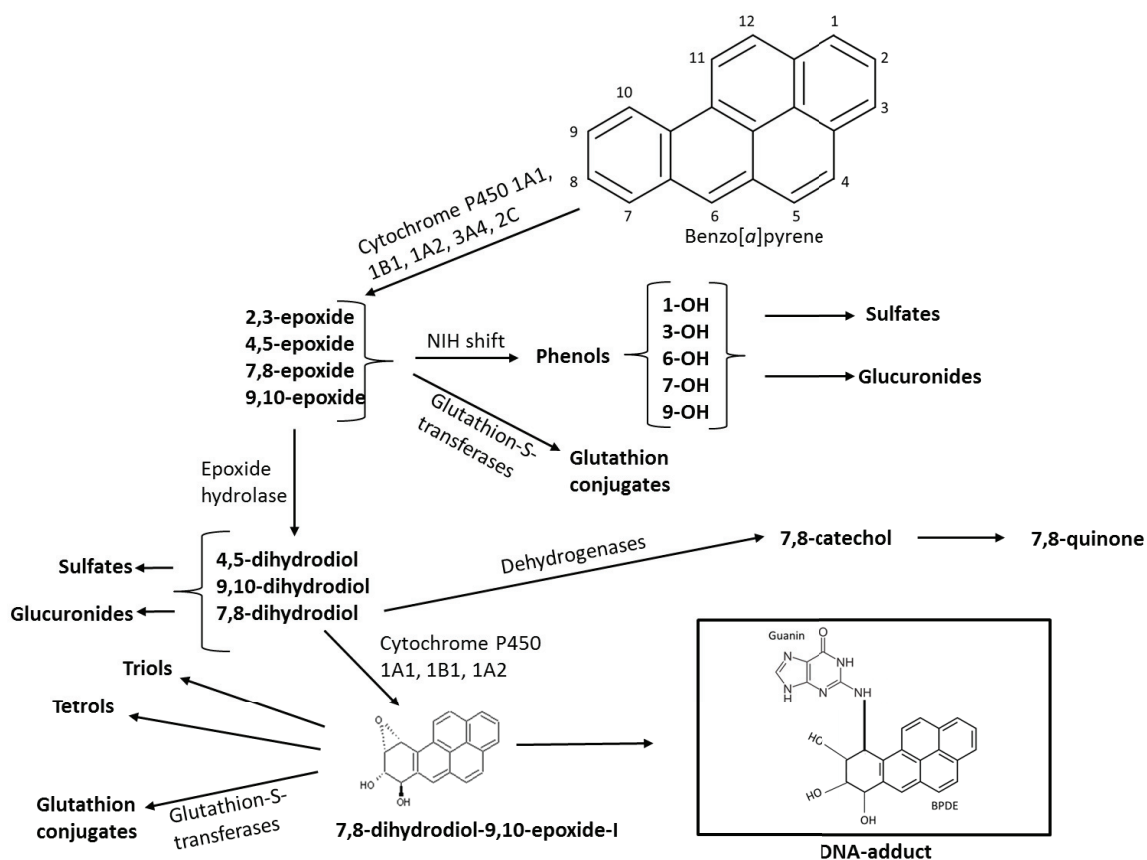


Figure 7. Metabolic pathways of B[*a*]P. Modified from (Hecht, 1999; Poirier, 2004).

1.3.5 B[*a*]P-DNA adduct formation

When BPDE I reacts with DNA, it preferentially becomes covalently linked to the exocyclic amino groups of deoxyguanosine and form adducts (Dipple, 1995; Wolfe *et al.* 2004). If the BPDE-DNA-adducts remain unrepaired, they can cause a miscoding during the replication process. The result may be transversion mutations and if the mutation occurs in a critical region of important genes, like oncogenes and tumor suppressor genes, the carcinogenesis process may thus be initiated (Dong *et al.* 2004; Greenblatt *et al.* 1994). Several studies have shown that DNA-adduct formation is associated with tumor induction in a dose-related fashion (Poirier, 2004; Poirier and Beland, 1992; Swenberg *et al.* 2008). Furthermore, epidemiological data have demonstrated a significant association between the formation of DNA-adducts and an increased risk of lung cancer (Kyrtpoulos, 2006; Veglia *et al.* 2008).

1.4. Sex steroid receptors and lung cancer

1.4.1 Estrogen receptors in lung

Estrogen receptor α ($ER\alpha$) was discovered by Elwood Jensen in the late 1950s, whereas estrogen receptor β ($ER\beta$) was discovered several decades later in 1996 (Jensen and Jacobson, 1960, Kuiper *et al.* 1996). The $ER\alpha$ gene ($ESR1$) is localized to chromosome 6q25.1. The gene corresponding to the full-length protein consists of eight exons. This protein contains 595 amino acid and has a molecular weight of 66216 Da. Twelve distinct splice variants have been identified (National Center for Biotechnology Information, PubMed Gene ID: 2099; Weickert *et al.* 2008). The $ER\beta$ gene ($ESR2$) also contains eight exons and is localized to chromosome 14q23.2. The corresponding full-length protein contains 535 amino acids with a molecular weight of 59216 Da. To date, at least six different splice variants of the receptor have been identified (National Center for Biotechnology Information, PubMed Gene ID: 2100, 2016).

The ERs are ligand inducible transcription factors and members of the nuclear steroid receptor superfamily. They typically mediate cellular responses to estrogens and share a high degree of sequence homology except for their NH₂-terminal domains. Furthermore, they have similar affinities for 17 β -estradiol (E₂) and bind the same estrogen response elements (ERE) on DNA (Heldring *et al.* 2007). $ER\alpha$ and $ER\beta$ can in some cases, however, show opposite actions. This has been explained by the receptors' individual capabilities to adopt a multitude of response states dependent on the bound ligand, with subsequent unique receptor conformational changes and recruitment of distinct coactivators and corepressors to the receptor-transcription complex in a cell-specific manner (Heldring *et al.* 2007; Liu *et al.* 2002).

Ligand binding to ER are followed by hetero- or homodimerization. In the nucleus the dimers bind to EREs on DNA and recruit co-activators to help induce target gene expression (Heldring *et al.* 2007). Alternatively, ERs can interact with transcription factors already bound to the DNA, such as activator protein-1 (AP-1) and stimulating protein 1 (Sp1). In fact, several

estrogen responsive genes do not contain classical EREs, but instead they often contain ERE half-sites, AP-1- and Sp1-sites or combinations of the two (O'Lone *et al.* 2004).

ER α and ER β exhibit tissue- and cell-type specific expression, but both receptors are expressed in histologically normal lung, lung tumor tissue, normal bronchial epithelial cells (HBECs) and lung tumor cell lines (Kawai *et al.* 2005; Mollerup *et al.* 2002; Stabile *et al.* 2002). ER α is, however, primarily reported in cytoplasm in human lung tissue (normal and tumor), whereas in HBECs, both receptors were predominantly found in cytoplasm (Ivanova *et al.* 2009; Stabile *et al.* 2002). Higher expression levels of both ER α and ER β have been demonstrated in tumor tissue compared with normal lung (Stabile *et al.* 2011).

Ivanova *et al.* (2009) reported that the antiestrogens 4-hydroxytamoxifen (4-OHT) and ICI 182780, respectively, inhibited proliferation of HBECs, and thus indicated endogenous ERs to be transcriptionally active (Ivanova *et al.* 2009). Moreover, E2-mediated transcription and proliferation in lung tumor cell lines were blocked by exposure to antiestrogens, providing evidence that ERs in lung cancers are functional and transcriptionally active (Hershberger *et al.* 2005; Shen *et al.* 2010). In addition, mouse lung was reported to be an estrogen-responsive tissue, with a 5-fold increase in luciferase activity in the lungs of transgenic mice upon E2-treatment (Ciana *et al.* 2001). Studies in *ER* knockout mice have revealed a sexual dimorphism in mouse lung. More specifically, in female mice the expression of *ERs* was required for formation of alveoli of appropriate size and number, whereas in male mice, the receptors had a smaller effect on alveolar dimensions (Massaro and Massaro, 2006).

1.4.2 Hormone replacement therapy (HRT) and antiestrogen treatment

The first two reports of a possible correlation between the use of hormone replacement therapy (HRT) and increased risk of lung cancer were proposed in the late 1980s (Adami *et al.* 1989; Wu *et al.* 1988). The evidence of an elevated lung cancer risk in smokers as a consequence of HRT was further strengthened in a small case-control study conducted by Taioli and Wynder in 1994 (Taioli and Wynder, 1994). Newer reports support the notion that HRT-use (combined estrogen and progesterone for > 5 years) elevates the risk of lung cancer and negatively affects prognosis and overall survival (Chlebowski *et al.* 2009; Ganti *et al.* 2006; Slatore *et al.* 2010).

There are, however, studies that do not support an association between HRT-use and increased risk of lung cancer. In a case-control study, self-reported use of HRT in lung cancer patients and healthy age-matched controls suggested an association between the use of HRT and a decreased risk of lung cancer (Schabath *et al.* 2004). Likewise, in a study by Rodriguez *et al.* (2008), HRT-use in postmenopausal women and lung cancer incidence by smoking status was reported inversely correlated (Rodriguez *et al.* 2008). Additionally, the Women's Health Initiative clinical trials demonstrated no increase in the risk of lung cancer in postmenopausal women treated with HRT (Schwartz *et al.* 2015), and a recently published study comprising 0.6 million female never-smokers found that current use of HRT in postmenopausal participants did not increase the risk of lung cancer (Pirie *et al.* 2016). The conflicting results emerging from these studies have emphasized the need for knowledge regarding the specific type of HRT used and the time and duration of use relative to lung tumor development (Siegfried and Stabile, 2014). In that respect, further mechanistic studies on ER and HRT in lung carcinogenesis and lung tumor biology are warranted. Evidently, there are highly complex interactions between multiple factors that may modulate the possible effect of HRT-use on lung cancer risk.

The role of estrogen in lung carcinogenesis has been reinforced by the observation that the use of antiestrogens is associated with decreased lung cancer mortality. Women diagnosed with breast cancer between 1980 and 2003 in the Geneva Cancer Registry, received antiestrogen treatment for their disease, and the epidemiological data showed that the women who received antiestrogen treatment had a 5-fold reduction of lung cancer mortality compared with the expected rates of the general population (Bouchardy *et al.* 2011). An additional study performed by Lothar *et al.* (2013) also found a significant decrease in lung cancer mortality among women who were treated with antiestrogens both prior to and after their lung cancer diagnosis (Lothar *et al.* 2013).

A recent clinical study reported that the inclusion of antiestrogens (tamoxifen) to standard chemotherapy regimens in treatment of NSCLC is promising. NSCLC patients who received chemotherapy with inclusion of tamoxifen had significantly higher median survival (13.1 months) compared to patients that received chemotherapy only (9.5 months) (Kadzhoian and Shevchenko, 2014, only abstract in english). Moreover, the aromatase enzyme that is essential

for the conversion of testosterone to estrogen, is active in normal lung tissue, lung cancer cell lines and lung tumors (Mah *et al.* 2007; Marquez-Garban *et al.* 2009; Weinberg *et al.* 2005). Estrogen stimulate growth in lung tumor cell lines and high aromatase activity has been detected in lung tumors (Mah *et al.* 2007; Marquez-Garban *et al.* 2009). Both aromatase inhibitors and antiestrogens are therefore considered potential anticancer drugs in NSCLC treatment (Lothar *et al.* 2013; Weinberg *et al.* 2005). Clinical and epidemiological data thus suggest an increased recognition of estrogens in lung carcinogenesis.

1.4.3 A reciprocal association between the signaling pathways of ERs and AHR

Several studies have reported a crosstalk between ER and AHR signaling pathways, but the molecular mechanisms behind this interaction are not fully understood. Most studies support a theory where *ER* is required for complete AHR-activity. This is partially based on the lack of 2,3,7,8-tetrachlorodibenzo-*p*-dioxin (TCDD)-induced *CYP1A1* expression in ER-negative cells that was restored by transient expression of *ERα* (Jana *et al.* 1999b, Thomsen *et al.* 1994).

ERα has been shown to indirectly impact on gene expression of *CYP1A1* in estrogen-dependent tissues such as breast and endometrium (Ricci *et al.* 1999; Thomsen *et al.* 1994). Moreover, a study by Matthews *et al.* (2007), showed that stable knockdown of *ERα* in HC11 mouse mammary cells, was followed by significant downregulation of induced *Cyp1a1* expression (Matthews *et al.* 2007). In addition, the antiestrogen 4-hydroxytamoxifen (4-OHT) has been shown to alter the transcriptional activity of AHR and induce expression of both *CYP1A1* and *CYP1B1* in breast cancer cell lines. Likewise, the antiestrogen ICI 182780 was reported to induce *CYP1A1* expression (DuSell *et al.* 2010).

Only a few studies have analyzed the ER-AHR crosstalk in lung cells. Normal bronchial epithelial cells transfected with an *ERα* adenoviral constructs, showed increased constitutive and cigarette smoke extract-induced mRNA and protein expression of *CYP1B1*, and chromatin immunoprecipitation assay (ChIP) confirmed *ERα* bound to the *CYP1B1* promoter. *ERα* transfection did not alter the *CYP1A1* mRNA levels, but increased the protein levels (Han *et al.* 2005). In contrast, transient overexpression of *ERα* in immortalized bronchial epithelial cells (BEAS-2B and BEP2D) had no effect on either basal or TCDD-induced expression of *CYP1A1*

and *CYP1B1*. Likewise, exposure of these cell lines to E2 did not influence constitutive or TCDD-induced levels of *CYP1A1* and *CYP1B1* (Berge *et al.* 2004), indicating that the crosstalk may be cell and/or tissue specific. *In vivo*, female *ERβ* knockout mice, but not male mice were protected against development of lung tumors after *in utero* exposure to the polycyclic hydrocarbon dibenzochrysene, indicating that *ERβ* plays a role in lung carcinogenesis in females (Benninghoff and Williams, 2013).

One possible mechanism for the observed crosstalk was proposed by Wormke *et al.* (2003). In this study TCDD was reported to induce an interaction between the AHR and *ERα*, while exposure to E2 did not induce such interactions. The study suggested a mechanism were TCDD induces formation of a nuclear AHR-complex which coordinately recruits *ERα* and the proteasome complex, resulting in degradation of both receptors (Wormke *et al.* 2003).

The similarities in molecular actions of ER and AHR, suggest that the two receptors may compete for coactivators necessary for recruitment of the transcription complex to the promoter regions of their respective target genes. Competition for common coactivators may thus provide another explanation for the interaction between the two signaling pathways (Monostory *et al.* 2009).

Besides being under the regulatory control of the AHR, the *CYP1B1* gene promoter contains an ERE that ER can bind to and initiate transcription upon exposure to E2 (Tsuchiya *et al.* 2004). Furthermore, the antiestrogens 4-OHT and ICI 182780 have been shown to effectively inhibit E2-induced *CYP1B1* expression in breast cancer cell lines, and when not co-exposed with E2, 4-OHT exhibited an agonistic function, upregulating *CYP1B1* expression (Tsuchiya *et al.* 2004). This proposes a different mode of action, other than an ER-AHR crosstalk, responsible for estrogenic effects on *CYP1B1* expression.

1.4.4 Androgen receptor in lung

The androgen receptor (AR) is the main mediator of male sex hormones. It is a zinc-finger transcription factor that belongs to the superfamily of nuclear receptors. The *AR* gene is

localized to the X-chromosome (Xq12), and the mRNA corresponding to the full-length protein contains 9 exons. The full-length protein has a molecular weight of 98989 Da and consists of 919 amino acids. To date, at least seven splice variants have been identified (National Center for Biotechnology Information, PubMed Gene ID: 367, 2016; Watson *et al.* 2010).

In the absence of ligand, the AR resides in the cytoplasm associated with heat-shock and other chaperone proteins. Once androgens bind to the AR, a conformational change takes place, chaperone proteins dissociate and nuclear localization sequences become exposed (Davey and Grossmann, 2016). In the nucleus, homodimers of AR form and bind to androgen response elements (AREs) in the promoter regions of target genes, to which they recruit various coregulatory proteins to facilitate transcription (Tan *et al.* 2015).

Both human and murine lung show expression of *AR*, principally in type II pneumocytes and bronchial epithelium (Mikkonen *et al.* 2010; Wilson and McPhaul, 1996). Several studies have shown that lung tumors express *AR*, and Mikkonen *et al.* (2010) reported functionality of the receptor in the human adenocarcinoma cell line A549 that exhibited androgen-dependent gene expression (Mikkonen *et al.* 2010; Rades *et al.* 2012). These findings confirmed earlier documented presence of *AR* in human lung and in NSCLC cell lines (Kaiser *et al.* 1996).

Androgens are well known regulators of lung development, specifically they delay lung maturation in males compared to females (Carey *et al.* 2007; Rodriguez *et al.* 2001). The *AR* has been implicated to act as a low penetrance cancer susceptibility gene in different tissues, in addition to prostate cancer where it plays a prominent role (Ferroa *et al.* 2002). Furthermore, *in vitro* studies have shown that AR may play a stimulatory role in lung cancer metastasis (Chang *et al.* 2014). Although NSCLC cell lines express *AR*, and expression levels have been reported to correlate with progression of NSCLC (Yan *et al.* 2008), only limited reports exist on AR's role in lung carcinogenesis.

1.4.5 A reciprocal association between the signaling pathways of AR and AHR

The effect of TCDD and other potent AHR-agonists` on AR transcriptional activity has been extensively studied. TCDD has been reported to decrease androgen-induced *PSA* expression in LNCaP cells, suggesting that TCDD might disrupt AR transcriptional activity in prostate cancer cells (Jana *et al.* 1999a). In lung cells, B[a]P has been shown to reduce both constitutive and testosterone induced expression of *AR* in NSCLC cell line H1355 and in immortalized human bronchial epithelial cell line BEAS-2B. Additionally, exposure to a carcinogenic metabolite of B[a]P (B[a]P-7,8-dihydrodiol-9,10-epoxide) in H1355 cells and human lung fibroblasts WI-38, was shown to significantly repress *AR* expression in both cell lines (Lin *et al.* 2004).

Studies in knockout mice revealed that *Ahr* expression was required for maintaining the inhibitory effect of TCDD on AR-driven gene transcription (Lin *et al.* 2002), which suggests a potential crosstalk between AHR and AR. Further support of such a crosstalk was found by Barnes-Ellerbe *et al.* (2004), who reported that AHR directly inhibited AR-activity, and that elevated levels of AHR-ARNT alone were adequate to downregulate AR-function (Barnes-Ellerbe *et al.* 2004). It has however also been suggested, that AP-1 is at least partially responsible for the antiandrogenic action of PAHs (Kizu *et al.* 2003).

AR has been shown to interact with the main modulator of *CYP1* expression, the AHR, upon exposure to the antiandrogen hydroxyflutamide (OHF) in hepatocellular carcinoma cell lines (Koch *et al.* 2015). Sanada *et al.* (2009) showed that dihydrotestosterone (DHT) repressed 3-methylcholantrene (3-MC) induced expression of *CYP1* genes in LNCaP and T47D cells. This inhibitory effect of DHT was abolished by knockdown of *AR*, and a significantly facilitated formation of AR-AHR complex in 3-MC treated cells was reported (Sanada *et al.* 2009). Furthermore, testosterone treatment in immature pigs *in vivo* significantly decreased the hepatic expression of *Cyp1a1* and *Cyp1a2* in both sexes (Kojima *et al.* 2008). However only limited data exist on AR`s influence on *CYP1* expression in lung. Although evidence toward a reciprocal association between the signaling pathways of AHR and AR exists, further elucidation of molecular mechanisms are necessary.

2. Aims of the study

The aim of this thesis was to clarify associations between PAH-bioactivation, sex differences in susceptibility to PAH and regulatory involvement of steroid receptors. A more detailed outline of the aims included:

- To establish the individual roles of *CYP1A1* and *CYP1B1* in bioactivation of PAH in human lung cells.
- To elucidate whether sex differences in capacity to bioactivate PAH are intrinsic characteristics of human lung cells.
- To examine the modulatory role of sex steroid receptors in regulation of *CYP1A1* and *CYP1B1* expression in human lung cells.

3. Summary of papers

Paper I

Importance of CYP1A1 and CYP1B1 in bioactivation of benzo[*a*]pyrene in human lung cell lines.

In paper I, the individual roles of *CYP1A1* and *CYP1B1* in bioactivation of benzo[*a*]pyrene (B[*a*]P) were analyzed in immortalized human bronchial epithelial cell line BEP2D and NSCLC cell line H2009. siRNA-technique was used to knockdown expression of *CYP1A1* and *CYP1B1*. Gene expression was measured with RT-qPCR (and normalized to the expression of β -actin), and formation of B[*a*]P-metabolites in cell culture medium was analyzed by fluorescence-HPLC. Several metabolites of B[*a*]P were formed in both cell lines. Knockdown of *CYP1A1* in BEP2D cells significantly reduced the formation of B[*a*]P-tetrol I-1 (hydrolysis product of the ultimate B[*a*]P carcinogen B[*a*]P-7,8-dihydrodiol-9,10-epoxide I; BPDE I) by 86 %, whereas knockdown of *CYP1B1* significantly reduced the formation by 35 %. When both *CYP1A1* and *CYP1B1* were knocked down the reduction of B[*a*]P-tetrol I-1 formation was 98 % in BEP2D cells, and similar results were found for H2009 cells. Three other B[*a*]P-tetrols (also hydrolysis products of B[*a*]P-7,8-dihydrodiol-9,10-epoxide) were measured, following a similar pattern, although at lower concentrations. Knockdown of *CYP1A1* caused higher levels of both B[*a*]P-*cis*-7,8-dihydrodiol and B[*a*]P-*trans*-7,8-dihydrodiol compared to control cells indicating a considerable role for *CYP1B1* in the formation of B[*a*]P-7,8-dihydrodiol. Significantly higher concentrations of unmetabolized B[*a*]P were found in cell culture media where both *CYP1A1* and *CYP1B1* were knocked down. In conclusion, in this paper we showed that the overall B[*a*]P-metabolism and formation of B[*a*]P-tetrol I-1 in human lung BEP2D and H2009 cells is primarily dependent on *CYP1A1*.

Paper II

Sex differences in susceptibility to PAH is an intrinsic property of human lung adenocarcinoma cells.

In paper II, we examined if sex differences in gene expression of important polycyclic aromatic hydrocarbon (PAH) metabolizing enzymes and ability to bioactivate B[*a*]P are intrinsic characteristics of NSCLC cell lines. Basal, cigarette smoke condensate (CSC)- and B[*a*]P-

induced gene expression was measured by RT-qPCR and normalized to the expression of β -*actin* (arbitrary units). CYP1-enzyme activity after exposure to CSC or B[a]P, respectively, was analyzed by EROD and formation B[a]P-adducts was measured by 32 P-postlabelling in 11 adenocarcinoma cell lines (5 of female and 6 of male origin). Significantly higher levels of *CYP1A1* gene expression were found in lung adenocarcinoma cell lines from women compared to cell lines from men: basal expression (median = 4.40 versus median = 0.23), B[a]P-induced (median = 73.64 versus median = 1.51) and CSC-induced (median = 208.78 versus median = 29.81). No significant sex differences in *CYP1B1* expression were observed in either control cells or B[a]P-/CSC-exposed cells. EROD activity after treatment with CSC was significantly higher in cell lines of female origin compared to those of male origin. Furthermore, significantly higher levels of B[a]P-DNA adducts were found in B[a]P-exposed lung adenocarcinoma cell lines from women (median = 403.6 adducts/ 10^8 nucleotides) compared to cell lines from men (median = 24.2 adducts/ 10^8 nucleotides). No significant sex differences in gene expression of *AHR*, *ARNT* or *AHRR* were detected. In conclusion, in this paper we showed a significant sex difference in B[a]P-bioactivation capacity in lung adenocarcinoma cell lines of female origin compared to cell lines of male origin. These results confirm previously published data showing sex differences in *CYP1A1* gene expression and hydrophobic DNA-adducts in female compared to male human lung tissue.

Paper III

Modulatory effect of sex steroid receptors on PAH metabolism genes in human lung cells.

In paper III, we studied if sex steroid receptors and their ligands could influence expression and induction of PAH bioactivating genes in human lung cells *in vitro*. Gene expression of estrogen receptors (*ER α* and *ER β*) and androgen receptor (*AR*) mRNA was measured by RT-qPCR in nine non-small cell carcinoma (NSCLC) cell lines and three immortalized bronchial epithelial cell lines (HBEC). Gene expression was normalized to the expression of β -*actin*. The effect of siRNA-mediated knockdown of *ER β* and *AR* on basal, CSC- and B[a]P-induced gene expression of *CYP1A1* and *CYP1B1*, respectively, was tested in the NSCLC cell line H2009. Furthermore, we examined how sex steroid receptor agonists/antagonists modulated gene expression of *CYP1A1* and *CYP1B1* in H2009. All NSCLC cell lines, with exception of H23 (negative for *ER α*) and H838 (negative for *AR*), showed detectable expression levels of *ER α* ,

ERβ and *AR*. However, large interindividual differences in expression levels were evident between the cell lines. When analyzed by sex of the donors of the NSCLC cell lines, no significant differences in expression were apparent. All three HBEC cell lines showed detectable expression of *ERα*, *ERβ* and *AR*. Knockdown of *ERβ* significantly reduced B[a]P-induced expression of *CYP1A1* (56 %) and *CYP1B1* (22 %) whereas knockdown of *AR* significantly enhanced both CSC- and B[a]P-induced expression of *CYP1B1* by 78 % and 64 %, respectively. Exposure to ICI 182780, which is a selective *ERα* protein degrader, induced expression of *CYP1A1* and *CYP1B1* by 3-fold. Exposure to the androgen dihydrotestosterone (DHT) significantly reduced B[a]P-induced expression of *CYP1A1* by 56 % and *CYP1B1* by 52 % when compared to cells that only received B[a]P. Thus, knockdown of *ERβ* and exposure to the *ERα* specific selective estrogen receptor degrader (SERD) resulted in contradictory effects on *CYP1* expression, which may indicate that the two receptors influence transcription in opposite directions. *ERβ* may be necessary for optimal induction of *CYP1* upon PAH-exposure, whereas *ERα* could be hypothesized to play a suppressing role on *CYP1* expression. The observed alterations inflicted on *CYP1* expression in *AR* knockdown- and DHT exposure experiments coincide, suggesting a common mechanism. In conclusion, in this paper we showed that sex steroid receptors might play a role in lung carcinogenesis through the modulation of xenobiotic bioactivation in lung cells.

4. Discussion

In lung cancer etiology, exposure to polycyclic aromatic hydrocarbons (PAHs) plays a central role (Veglia *et al.* 2008). The sources of inhalation exposure are mainly designated cigarette smoke, urban air and certain occupational environments (Bostrom *et al.* 2002; Hashim and Boffetta, 2014; Hecht, 2003). The ultimate carcinogenic potential of PAHs generally depend on enzymatic bioactivation yielding highly reactive intermediates that may react with DNA and form PAH-DNA adducts (Shimada and Fujii-Kuriyama, 2004). Cytochrome P450 (CYP) 1A1 and CYP1B1 are important PAH-metabolizing enzymes in human lung, and a clear association between *CYP1* expression, formation of PAH-DNA adducts and lung carcinogenesis has been established (Moorthy *et al.* 2015; Veglia *et al.* 2008). PAHs that enter lung cells can readily bind to the aryl hydrocarbon receptor (AHR), a ligand-activated transcription factor. The expression of *CYP1A1* and *CYP1B1* in human lung is highly inducible by PAH-exposure through the action of AHR (Shimada and Fujii-Kuriyama, 2004).

Distinct clinical characteristics and unequal lung cancer mortality rates between men and women have suggested the presence of sex differences in tumor biology and risk of lung cancer (Burns and Stabile, 2014). Both epidemiological data and animal studies support the notion that lung tumor biology differs between males and females (Amr *et al.* 2008; Kiyohara and Ohno, 2010; Marquez-Garban *et al.* 2007; Ninomiya *et al.* 2013; Patel *et al.* 2004; Ramchandran and Patel, 2009; Wakelee *et al.* 2007; Yang *et al.* 2005). Although not conclusive, studies indicate that women may be at greater risk of developing lung cancer from cigarette smoking than men (Novello and Baldini, 2006; Patel *et al.* 2004; Ramchandran and Patel, 2009). Furthermore, women are typically diagnosed at an earlier age than men are (Nagy-Mignotte *et al.* 2011; Thomas *et al.* 2015). The rates and the extent of cellular processes such as metabolic activation of procarcinogens, detoxification, DNA-adduct formation and DNA-repair may influence individual susceptibility to the tumorigenic effects associated with PAH-exposure (Benhamou *et al.* 2002). In that respect, significantly higher *CYP1A1* gene expression and levels of PAH-DNA adducts have been reported in female lung cancer patients compared to male patients suggesting a higher susceptibility to PAH among women (Mollerup *et al.* 1999, 2006).

It has been hypothesized that steroid receptors may play a role in the possible sex differences in lung cancer risk and susceptibility to PAH (Townsend *et al.* 2012). Both estrogen receptors (ER α and ER β) (Ivanova *et al.* 2009; Mollerup *et al.* 2002; Stabile *et al.* 2002) and androgen receptor (AR) (Mikkonen *et al.* 2010; Wilson and McPhaul, 1996) are expressed and functional in human lung cells. Furthermore, *in vitro* studies have indicated interactions between the signaling pathways of AHR and ERs resulting in altered gene expression of *CYP1* genes (Matthews *et al.* 2007; Ricci *et al.* 1999; Thomsen *et al.* 1994). A reciprocal association between AHR and AR has also been suggested by several studies (Lin *et al.* 2002; Lin *et al.* 2004). Although the extent and the mechanisms involved are not fully understood, sex steroid receptors may thus be implicated in the regulation of *CYP1* genes and metabolic activation of PAH in lung cells.

The major topic of this thesis is associations between PAH-bioactivation, sex differences in susceptibility to PAH and regulatory involvement of steroid receptors in human lung cells. The first paper is designated the respective roles of CYP1A1 and CYP1B1 in bioactivation of B[a]P. The second paper is assigned the elucidation of whether previously reported *in vivo* sex differences in PAH-DNA adduct formation and *CYP1A1* expression are intrinsic characteristics of human lung cells. The last paper is based on an analysis of sex steroid receptors' potential incursion on *CYP1* expression in lung cells.

4.1. The individual roles of CYP1A1 and CYP1B1 in bioactivation of benzo[a]pyrene in human lung cells

Benzo[a]pyrene (B[a]P) is the most thoroughly studied prototype PAH and is often present at high concentrations in PAH polluted environments (Conney, 1982; Kleiner *et al.* 2004; Pelkonen and Nebert, 1982). CYP1A1 and CYP1B1 enzymes are considered to play important roles in bioactivation of B[a]P to B[a]P-dihydrodiol-epoxides (BPDEs). These electrophilic intermediates, in particular BPDE I, have the capacity to react with DNA and form B[a]P-DNA adducts, which are considered molecular dose-indicators predictive of cancer risk (Veglia *et al.* 2008). Individual differences in the ability to bioactivate lung carcinogens like PAH and formation of DNA adducts may be a determining factor in lung cancer susceptibility (Kouri *et*

al. 1982; Shimada and Fujii-Kuriyama, 2004). Therefore it was important to establish the respective roles of CYP1A1 and CYP1B1 in bioactivation of B[a]P in human lung cells.

In **Paper I** we showed how siRNA-mediated knockdown of *CYP1A1* reduced the formation of B[a]P-tetrol I-1 (hydrolysis product of B[a]P-7,8-dihydrodiol-9,10-epoxide I; BPDE I) by 86 %, whereas knockdown of *CYP1B1* reduced the formation by 35 % in BEP2D (immortalized human bronchial epithelial) cells. When both *CYP1A1* and *CYP1B1* were knocked down the reduction of B[a]P-tetrol I-1 formation was 98 %, with similar results in H2009 non-small cell lung carcinoma (NSCLC) cells. The three other B[a]P-tetrols measured (B[a]P-tetrol I-2, II-1 and II-2) followed a similar pattern, although at much lower concentrations. The highest concentrations of unmetabolized B[a]P were found in cells where both *CYP1A1* and *CYP1B1* were knocked down. To sum up, in BEP2D and H2009 cells the overall B[a]P-metabolism and formation of B[a]P-tetrol I-1 are primarily dependent on *CYP1A1*. However, the results also indicated a significant role for CYP1B1 in the formation of B[a]P-7,8-dihydrodiol (B[a]P-7,8-DHD), a precursor of BPDE I.

An early study by Gautier *et al.* (1996) analyzed the metabolism of B[a]P and B[a]P-7,8-DHD in the presence of human microsomal epoxide hydrolase and either CYP1A1, 1A2, 2C8, 2C9, 2C18, 2C19, 2D6 or 3A4 expressed in *Saccharomyces cerevisiae* (Gautier *et al.* 1996). CYP1A1 was shown to have the highest turnover numbers for the formation of all B[a]P metabolites (Gautier *et al.* 1996), and this is in agreement with the observed significance of CYP1A1 in B[a]P-metabolism reported in **Paper I**. Further support of a causal relationship between *CYP1A1* expression and bioactivation in lung cells was demonstrated in normal lung tissue from lung cancer patients where expression of *CYP1A1* and levels of PAH-related DNA adducts showed a strong, significant correlation (overall Spearman ranked correlation coefficient: $r^2=0.66$) (Mollerup *et al.* 2006). Likewise, Hodek *et al.* (2013) demonstrated that in rat liver and intestine, increased activity of CYP1A1 significantly enhanced the bioactivation and B[a]P-DNA adduct formation upon oral exposure to B[a]P (Hodek *et al.* 2013).

Paper I indicated a significant role for CYP1B1 in the formation of B[a]P-7,8-DHD. This coincides with data from Shimada *et al.* (2001) who showed that CYP1B1, together with

epoxide hydrolase, catalyzed the conversion of B[a]P to B[a]P-7,8-DHD at much higher (10-fold) rate than CYP1A1 in a cell free system of microsomes (Shimada *et al.* 2001).

In contrast to observations in **Paper I**, studies in knockout mice orally exposed to B[a]P reported that Cyp1a1 may have a more prominent role in detoxification than bioactivation. *Cyp1a1*^{-/-} mice given oral B[a]P died within 30 days, whereas *Cyp1a1* wild-type mice survived throughout the one-year long experiment. Furthermore, B[a]P-DNA adducts were significantly higher in internal organs of *Cyp1a1*^{-/-} mice compared to wild-type mice, despite of a significantly lower B[a]P-metabolism and slower clearance (Uno *et al.* 2004, 2006). The authors concluded that *in vivo*, Cyp1a1 have a protective role against orally ingested PAH. The elevated level of adducts found in *Cyp1a1*^{-/-} mice was explained by reduced phase II conjugation due to the lack of coupling between phase I and phase II enzymes in the knockout models (Nebert *et al.* 2004).

Studies in *Ahr* knockout mice support the notion that the distributions and levels of B[a]P and B[a]P-protein adducts are clearly dependent on the route of exposure (Godschalk *et al.* 2000; Sagredo *et al.* 2006, 2009). Sagredo *et al.* (2006) reported that orally exposed *Ahr* knockout mice had higher levels of B[a]P and protein adducts than intraperitoneal or skin- treated knockout mice (Sagredo *et al.*, 2006). Furthermore, in adduct-analysis of skin topically applied with B[a]P, the *Ahr* wild-type mice showed significantly higher levels of protein adducts compared to *Ahr*^{-/-} mice. In contrast, analysis of skin distal to the B[a]P-application site revealed significantly lower levels of protein adducts in *Ahr* wild-type mice compared to *Ahr*^{-/-} mice (Sagredo *et al.* 2009). These results coincide with an important role of liver and skin in a first-pass effect that reduces the systemic uptake of B[a]P and thus subsequent formation of adducts in wild-type mice (Nebert, 1989). Our results reveal a central role of CYP1A1 in bioactivation of B[a]P in human lung cells, which differs from studies in *Cyp1a1* knockout mice (Uno *et al.* 2004, 2006). However, the reliability of our results is ensured by studies reporting that differences in target organ toxicity can be designated the route of administration used in the respective experiments (Nebert *et al.* 2004; Routledge and Shand, 1979; Sagredo *et al.* 2006, 2009).

In **Paper I** CYP1B1 was designated a minor role in bioactivation of B[a]P in human lung cells compared to CYP1A1. In other studies however, a PAH-bioactivating role of *Cyp1b1* has been strengthened. Studies in *Cyp1b1*^{-/-} mice showed that they exhibited increased protection against 7,12-dimethylbenz[a]anthracene toxicity and dibenzo[a,l]pyrene induced tumors compared to *Cyp1b1* wild-type mice (Buters *et al.* 2002; Page *et al.* 2003). Furthermore, exposure to dibenzo[def,p]chrysene (DBC) in *Cyp1b1* knockout mice resulted in significantly reduced adduct formation in lung when compared to exposed wild-type mice. Cyp1b1 is thus indicated as the primary Cyp1 enzyme involved in bioactivation of DBC to DBC-diolepoxide (Harper *et al.* 2015). Together, these data illustrate the distinct substrate specificities of CYP1A1 and CYP1B1 toward different PAHs. The respective roles of CYP1A1 and CYP1B1 in bioactivation of different PAHs depend on the chemical structure of the compound (Murray *et al.* 2001; Shimada *et al.* 2001; Shimada and Fujii-Kuriyama, 2004). A possible reason for the enzymes' respective preferences has been argued to be related to differences of hydrogen bond donor/acceptor side chains in the two enzymes putative active site regions (Lewis, 1999). Studies conducted in cell free systems however, indicate that CYP1A1 and CYP1B1 have relatively similar substrate specificities toward various PAHs. The enzymes exhibit the same activity in metabolism of PAH-dihydrodiols such as 7,12-DMBA-3,4-dihydrodiol, DB[a,l]P-11,12-dihydrodiol, 5-methylchrysene-1,2-dihydrodiol and B[a]A-3,4-dihydrodiol (Shimada *et al.* 2001).

In **Paper I**, cells transfected with either *CYP1A1* or a combination of *CYP1A1* and *CYP1B1* siRNA showed the highest levels of unmetabolized B[a]P in culture media. When interpreting the results one should however bear in mind that the measured quantities of B[a]P and B[a]P-metabolites, are not an absolute measure due to low recovery during preparation on Sep-Pak C18 cartridges. Nevertheless, the result reveal that the overall metabolism of B[a]P was significantly reduced in cells with downregulated expression levels of *CYP1A1*. This underlines the importance of CYP1A1 in degradation of B[a]P in human lung cells. Both cell lines, immortalized human bronchial epithelial cells and NSCLC cells, showed similar metabolic profiles and similar effects upon transfection with *CYP*-siRNA. Together with results from other *in vitro* and *in vivo* studies, **Paper I** provides evidence that CYP1A1 plays an important role in bioactivation of PAH and thus show a stimulating effect on PAH-induced lung carcinogenesis.

4.2 Sex differences in expression of *CYP1A1* and capacity to form B[a]P-DNA adducts in human lung cells

Smoking is considered the main causative agent regarding development of lung cancer, and among the potential human carcinogens present in cigarette smoke are ten different PAHs (IARC Monographs Vol 83, 2004). The majority of PAHs present in cigarette smoke have the ability to induce *CYP1* expression by ligand-activation of the transcription factor AHR, and CYP1 enzymes are essential in bioactivation of PAH (Shimada and Fujii-Kuriyama, 2004). It has been hypothesized that sex differences in lung cancer tumor biology may partially originate from different abilities to bioactivate procarcinogenic PAHs. Differences in PAH bioactivation could affect susceptibility to PAH, and this hypothesis is strengthened by studies where the expression levels of *CYP1A1* and the levels of aromatic/hydrophobic DNA adducts were reported to be significantly higher in the lungs of female smokers compared to the lungs of male smokers. In addition, a strong and significant correlation between *CYP1A1* expression and levels of formed PAH-DNA adducts was found, irrespective of sex and smoking status (Mollerup *et al.* 2006).

In **Paper II** we verified that previously reported sex differences in *CYP1A1* expression and PAH-DNA adduct formation in human lung also are preserved in NSCLC cell lines. More specifically, **Paper II** showed significantly higher levels of basal, CSC- and B[a]P-induced *CYP1A1* gene expression, CSC-induced CYP1 activity and levels of formed B[a]P-DNA adducts in lung adenocarcinoma cell lines from women compared to cell lines from men. In contrast, no significant sex difference in expression levels of *CYP1B1*, *AHR*, *ARNT* or *AHRR* mRNA were identified. These results are in agreement with previous studies conducted on normal lung tissue from current smokers, where females had a significantly higher median level of *CYP1A1* mRNA and significantly higher levels of PAH-DNA adducts compared to currently smoking males (Mollerup *et al.* 2006).

In **Paper II**, the observation of high *CYP1A1* expression, together with high levels of B[a]P-DNA adducts and high CYP1 enzyme activity, mutually support one another. This association is reinforced by previous studies where coherence between CYP1A1 activity and pulmonary PAH-associated DNA adductions was reported (Bartsch *et al.* 1992 a,b). Furthermore, current smoke exposure is considered a major determinant of *CYP1A1* induction in human lung, and

levels of *CYP1A1* expression and DNA-adducts have been shown to significantly correlate in current smokers (McLemore *et al.* 1990; Mollerup *et al.* 1999). Likewise, formation of B[a]P-DNA adducts was reported to correlate with CYP1A1 protein expression, but not with expression levels of CYP1B1 protein in normal mammary epithelial cells and breast cancer cell line MCF-7 (Divi *et al.* 2014).

The degree of interindividual variability in basal expression of *CYP1A1* and *CYP1B1* and the large differences in *CYP1A1* inducibility between the cell lines in the present study coincide with data from lung tissue (Mollerup *et al.* 2006). Coherence with *in vivo* data, also applies to the lack of observable sex differences in gene expression of *CYP1B1* in both **Paper II** and Mollerup *et al.* (2006).

An increased induction of *CYP1A1* mRNA expression and CYP1 activity upon exposure to CSC, compared to B[a]P, was observed in **Paper II**. The CSC used in our experiments has been extracted from the particulate phase of mainstream smoke from Kentucky reference cigarettes 2R4F, and contains at least 13 different PAHs (Roemer *et al.* 2014). The yield of CSC per cigarette is 26.1 mg, and analysis show that this comprises 8.9 mg tar per cigarette, which makes up 1/3 of its constituents (Richter *et al.* 2010). Additionally, the CSC contains a complex composition of other compounds, and halogenated aromatic hydrocarbons (HAH) present in CSC may contribute to the observed elevated induction of *CYP1A1* in comparison to B[a]P (Wilson *et al.* 2008).

Several animal studies support the notion of a sex difference in *Cyp1a1* expression. In mature Meishan pigs, the hepatic gene expression of *Cyp1a1* was reported to be higher in females than in males, but not in castrated males. No such sex difference was reported in immature pigs, but treatment with testosterone markedly decreased the expression levels of *Cyp1a1* in both sexes (Kojima *et al.* 2008). Furthermore, recent studies in mice found that *Cyp1a1* gene expression was significantly higher in homogenates from female whole embryos compared to male embryos (Macak-Safranko *et al.* 2011; Penalozza *et al.* 2014). Also in rat kidney, throughout the whole life cycle, female rats showed in average 4-fold higher expression of *Cyp1a1* than male rats (Kwekel *et al.* 2013).

The mechanisms behind the observed sex difference in *CYP1A1* expression observed both *in vivo* (Mollerup *et al.* 1999, 2006) and *in vitro* (**Paper II**) are not clear, although several mechanisms have been proposed. One possible explanation for the observed sex differences in expression of *CYP1A1* could be related to the AHR. However, we were not able to detect any sex differences in measured expression of *AHR* mRNA in **Paper II**, and the levels of *AHR* mRNA were relatively homogeneous across all the cell lines. **Paper II** shows significant correlations between expression levels of *AHR* mRNA and *CYP1A1* mRNA, and between *AHR* mRNA and *CYP1B1* mRNA. These results confirm that AHR plays an important role in transcriptional regulation of *CYP1*.

DNA-methylation status of gene promoter regions modulate the accessibility for transcription factors and thus expression of genes. Sex distinct patterns of DNA-methylation in the promoter of *Cyp1a1* have been reported. The gene expression of *Cyp1a1* was reported higher in both embryonal and adult lung cells from female mice compared to male mice. Four distinct sites in the promoter region of *Cyp1a1* were methylated in female mice, whereas in males only three sites showed methylation (Penaloza *et al.* 2014). When exposed to E2, the methylation status was altered, and the sex difference in *Cyp1a1* gene expression diminished between female and male mice. This may indicate that E2 normally helps maintain the sex difference in gene expression since only female mice usually become exposed to E2 (Penaloza *et al.* 2014). In contrast, quantitative analysis of *CYP1A1* enhancer DNA methylation levels in normal lung tissue from NSCLC patients revealed no such sex differences (Tekpli *et al.* 2012). The results, however, indicated that low levels of methylation were associated with high mRNA expression of *CYP1A1*. The study also reported that the methylation levels were significantly higher in never-smokers indicating that smoking may alter DNA-methylation status of this gene.

A hypothesis of the involvement of steroid hormones in lung tumorigenesis and sex differences in risk of lung cancer has been proposed (Haugen, 2002; Rivera, 2013). This hypothesis may be extended to the observed sex differences in *CYP1A1* gene expression and PAH-DNA adduct formation *in vitro* (**Paper II**) and *in vivo* (Mollerup *et al.* 1999, 2006). Accordingly, the main objective of **Paper III** was to analyze whether sex steroids and their receptors could influence *CYP1* expression in lung cells.

To sum up, the importance of CYP1A1 in bioactivation of B[a]P reported in **Paper I**, and the relationship between expression levels of *CYP1A1* and elevated levels of B[a]P-DNA adducts in female cell lines compared to male cell lines reported in **Paper II**, mutually strengthen one another. Combined, the *in vitro* data from **Paper I** and **Paper II** lend support to the previously published *in vivo* data (Mollerup *et al.* 1999, 2006), and strengthen the hypothesis that female lung may be more sensitive to PAH-exposure than male lung. The vast majority of present knowledge on biochemical processes and signaling pathways in human cells have been obtained in cell lines with no knowledge on or attention to their sex origin (Shah *et al.* 2014). The significant sex differences in gene expression, enzyme activity and DNA adduction observed in **Paper II**, encourage the notion that the sex of cell lines used during *in vitro* studies should be considered to the same extent as in the case of *in vivo* studies.

4.3 Sex steroid receptors' impact on *CYP1* regulation in lung cells

Evidence has accumulated toward sex differences in the biology of lung cancer, presented by distinct clinical characteristics and unequal mortality rates. Experimental studies indicate that sex steroid receptors may play a role in the pathogenesis of lung cancer, although the exact role of sex steroids and their receptors remains unclear (Burns and Stabile, 2014). In **Paper III** we studied the modulatory effect of ER and AR on PAH metabolism in human lung cells through siRNA-mediated receptor knockdown and exposure to a panel of sex steroid receptor ligands.

In **Paper III** we report that *ER α* , *ER β* and *AR* mRNA are expressed in both NSCLC and human bronchial epithelial (HBEC) cells. This coincides with earlier gene expression analyses of sex steroid receptors in lung tissue and lung cell lines (Ivanova *et al.* 2009; Mikkonen *et al.* 2010; Mollerup *et al.* 2002; Stabile *et al.* 2002). Furthermore, we show that knockdown of *ER β* and *AR* influence the mRNA levels of *CYP1A1* and *CYP1B1* in the human NSCLC cell line H2009. However, the two receptors seem to modulate the expression levels in divergent directions. More specifically, siRNA-mediated *ER β* knockdown significantly downregulated B[a]P-induced gene expression of both *CYP1A1* and *CYP1B1*, whereas *AR* knockdown significantly upregulated both CSC- and B[a]P-induced gene expression of *CYP1B1*. Exposure to sex steroid receptor ligands also modulated the expression levels of *CYP1A1* and *CYP1B1*. The ER α

specific degrader ICI 182780 induced gene expression of both *CYP1A1* and *CYP1B1*, whereas exposure to dihydrotestosterone (DHT) significantly decreased B[a]P-induced mRNA levels of *CYP1A1* and *CYP1B1* in H2009 cells. Together these observations support the hypothesis of a role of sex steroid receptors in modulation of *CYP1* expression.

The NSCLC cell lines analyzed in **Paper III** showed no sex difference in expression levels of *ER α* , *ER β* or *AR*. This is in agreement with former studies where no sex difference in gene expression of *ER* has previously been reported in human lung (Ivanova *et al.* 2009, 2010; Mollerup *et al.* 2002; Stabile *et al.* 2002). However, Dougherty *et al.* (2006) reported an increased transcriptional activity of ER present in NSCLC cell lines of female origin compared to cell lines of male origin, and the cellular localization of the sex steroid receptors may play a role in their transcriptional activity and hence their functionality (Dougherty *et al.* 2006; Ivanova *et al.* 2009, 2010; Kawai, 2014). Regarding expression of the *AR*, at present little or no information on sex differences in expression and/or activity in human lung exists.

In **Paper III**, *ER β* and *AR* were chosen as knockdown targets based on their expression levels in lung cells and their previously described importance in lung development and maturation (Patrone *et al.* 2003; Rodriguez *et al.* 2001). Furthermore, exclusion of *ER α* knockdown in this study was decided based on previous transfection experiments with *ER α* in lung cells conducted by our group, which showed no alterations on the expression levels of neither *CYP1A1* nor *CYP1B1* upon overexpression of *ER α* (Berge *et al.* 2004).

Knockdown of *ER β* mRNA resulted in subsequent downregulation of B[a]P-induced *CYP1A1* and *CYP1B1* gene expression. Downregulation of *CYP1B1* may be related to presence of an active estrogen responsive element (ERE) in the *CYP1B1* promoter region (Tsuchiya *et al.* 2004). Likewise, Penaloza *et al.* (2014) identified an ERE in the promoter region of *Cyp1a1* in mice, indicating that ERs also directly interact with transcriptional initiation of *Cyp1a1* expression (Penaloza *et al.* 2014). Limited amounts of available ER β after siRNA-treatment may thus inhibit normal transcription of *CYP1A1* and *CYP1B1*. Additionally, a basic transcription element (BTE) has been reported to control the constitutive expression of *CYP1A1* (Yanagida *et al.* 1990). The BTE binds Sp1, a transcription factor known to directly interact with ER forming an ER-Sp1 complex (Imataka *et al.* 1992; Safe and Kim, 2004). Thus, a

decrease in *ERβ* may reduce binding to this BTE, and hence reduce normal expression levels of *CYP1A1*.

We found that ICI 182780 induced expression of *CYP1A1* and *CYP1B1* in the NSCLC cell line H2009. ICI 182780 degrades *ERα* (Yeh *et al.* 2013), and simultaneously induces expression of *CYP1A1* and *CYP1B1*. The presence of *ERα* may thus contribute in the opposite direction as *ERβ*, suppressing *CYP1* expression. ICI 182780 may also function as an AHR-agonist. Several synthetic ER ligands, including ICI 182780, possess the ability to activate AHR and significantly enhance expression of *CYP1A1* in MCF-7 breast cancer cells (DuSell *et al.* 2010).

Upregulated levels of B[a]P-/CSC-induced *CYP1B1* gene expression in cells pretreated with *AR* siRNA, may be related to the presence of several steroidogenic factor-1 (SF-1) responsive elements in the upstream enhancer region of *CYP1B1* (Zheng and Jefcoate, 2005). The AR has been reported to suppress transcription of genes regulated by SF-1, involving a mechanism where AR directly binds to already DNA-bound SF-1 in the promoter region of the distinct genes, and inhibits transcription initiation (Jorgensen and Nilson, 2001).

In **Paper III**, DHT repressed B[a]P-induced gene expression of both *CYP1A1* and *CYP1B1*. The observed alterations inflicted on *CYP1* expression in *AR* knockdown- and DHT exposure experiments coincide, and implicate a common mechanism for the modulatory effect on *CYP1* expression. The mode of action may involve whether or not an AR-SF-1 interaction takes place in the promoter region of *CYP1B1*. DHT-bound AR may translocate to the nucleus where it will bind directly to SF-1 proteins already present on the promoter of *CYP1B1*, and elicit the observed reduced induction. The mechanism underlying DHT-induced repression of *CYP1A1* is unknown. However, a possible involvement of activator protein-1 (AP-1) in the anti-androgenic effects of AHR-agonists have been suggested. More specifically, this mechanism is suggested to involve AHR-stimulated *AP-1* expression and AP-1 is known to inhibit binding of AR to androgen responsive elements (AREs) by protein-protein interaction with the receptor (Kizu *et al.* 2003).

Animal studies have indicated that 4-(methylnitrosamino)-1-(3-pyridyl)-1-butanone (NNK)-induced lung carcinogenesis may be inhibited by testosterone (Ninomiya *et al.* 2013). These data, together with reports showing the importance of androgens in lung development and maturation (Rodriguez *et al.* 2001) and results presented in **Paper III**, support that lung tissue is responsive to androgen exposure.

Several studies have reported that the signaling pathways of ER and AHR may interact in a crosstalk (DuSell *et al.* 2010; Jana *et al.* 1999b; Matthews *et al.* 2007; Ricci *et al.* 1999; Thomsen *et al.* 1994; Wormke *et al.* 2003). In normal bronchial epithelial cells, Han *et al.* (2005) reported increased constitutive and cigarette smoke extract-induced mRNA and protein expression of *CYP1B1* in cells transfected with *ERα*-adenoviral constructs. In addition, *ERα* was confirmed to bind to the *CYP1B1* promoter. Still, *ERα* transfection did not alter the *CYP1A1* mRNA levels, but increased *CYP1A1* protein levels (Han *et al.* 2005). A reciprocal influence has also been reported between the AR and AHR, but only limited data exist (Lin *et al.* 2002; Lin *et al.* 2004). A theory of competition for common nuclear receptor coactivators necessary for recruitment of the transcription initiation complex to the promoter regions of their respective target genes has been proposed as an underlying mechanism for such crosstalks. Nuclear receptor coactivator-1 (NCOA1), CREB binding protein (CBP/p300) and Sp1 are all regulatory proteins involved in transcription initiation mediated by several nuclear receptors including AHR, ERs and AR (Esser and Rannug, 2015, Heldring *et al.* 2007). A rivalry for common coactivators may provide an explanation for the observed alterations on AHR transcriptional activity (Monostory *et al.* 2009).

Cancer susceptibility as a consequence of sex steroid receptor expression and exposure to sex steroids has been indicated in studies conducted on *Ar* knockout mice. The results revealed that exposure to NNK-B[a]P contributed to lower lung tumor incidence and reduced tumor size when compared to wild-type mice receiving the same treatment (Yeh *et al.* 2012). Additionally, female A/J mice were confirmed to be more susceptible to NNK-induced lung carcinogenesis than male mice (Ninomiya *et al.* 2013). NNKs like PAHs, are bioactivated by CYP1A1, and in male mice, lung carcinogenesis was significantly increased by castration, whereas in females, malignant transformation of lung proliferative lesions tended to be inhibited by ovariectomy (Ninomiya *et al.* 2013).

In conclusion, **Paper III** showed that knockdown of *ERβ* or *AR* and exposure to sex steroid receptor ligands, respectively, have a significant impact on *CYP1* gene expression. The results support a role of sex steroid receptors in modulation of xenobiotic bioactivation in lung cells. These mechanisms could also be speculated to play a role in the previously reported higher levels of *CYP1A1* expression and PAH-DNA adduction in both female NSCLC patients and NSCLC cell lines derived from women compared to their male counterparts (Mollerup *et al.* 1999, 2006, **Paper II**). However, further clarification of mechanisms behind the modulatory effects is warranted. The matter of sex differences in risk of lung cancer is still not completely sorted out. Of particular relevance for this thesis, a specific mechanistic linkage between steroids and sex differences in *CYP1A1* expression and formation of PAH-DNA adducts is still incomplete and should be further examined.

5. Conclusions and future perspectives

The work presented in this thesis has provided new insight regarding regulation of PAH bioactivation in lung cells. In conclusion, we have:

- ✓ Established that CYP1A1 plays a major role in bioactivation of B[a]P in lung cells.
- ✓ Verified that significantly higher levels of *CYP1A1* expression and significantly higher levels of B[a]P-induced DNA adducts is an intrinsic capacity in lung cell lines from women compared to cell lines from men.
- ✓ Showed that sex steroid receptors may modulate the expression of important PAH-metabolism genes in lung cells.

Data provided by this thesis indicate that female lung cells have higher PAH-bioactivation capacity than male lung cells, and that sex differences in expression of *CYP1A1* may be mechanistically involved in these differences. However, our results warrant further exploration, and the need for clarification of molecular mechanisms in the matter of differences in lung tumor biology between women and men is invoking.

Tobacco smoking is still recognized as one of the world's most important health threats. The epidemic of smoking is rising amongst women globally, and female smoking is predicted to double between 2005 and 2025 (Amos *et al.* 2012). Young urbanized women are often among the first to adopt cigarette smoking in low-income countries, and a link between gender empowerment and smoking has been observed (Hitchman and Fong, 2011). Airway exposure to PAH increases the risk of lung cancer in humans, and lung cancer risk partially depend on the proximity to PAH-emission sources (including tobacco smoking) and individual susceptibility (Shen *et al.* 2014). An increased susceptibility to PAH amongst women is especially important to clarify in order to implement these findings in anti-smoking campaigns specifically directed toward girls and women (Amos *et al.* 2012).

Furthermore, sex differences are often inadequately addressed in risk assessment. If and how xenobiotic metabolism is influenced by gender is not always predictable given the current state of knowledge. Differences in gene expression between males and females should be further explored to develop a broader understanding of sex-specific toxicity and carcinogenesis (Vahter *et al.* 2007). It is estimated that the risk of lung cancer could be underestimated by nearly 50 %

if individual susceptibility is not taken into consideration in risk assessments (Shen *et al.* 2014). Furthermore, results from this thesis demonstrate that it is important to consider sex of the cell lines when trying to identify and understand the complexity of cellular mechanisms. Male and female cells are not the same (Dewing *et al.* 2003; Pollitzer, 2013; Shah *et al.* 2014).

6. References

- Adami HO, Persson I, Hoover R, Schairer C, Bergkvist L. Risk of cancer in women receiving hormone replacement therapy. *Int J Cancer*. 1989 Nov 15;44(5):833-9.
- Algranti E, Buschinelli JT, De Capitani EM. Occupational lung cancer. *J Bras Pneumol*. 2010 Nov-Dec;36(6):784-94.
- Amos A, Greaves L, Nichter M, Bloch M. Women and tobacco: a call for including gender in tobacco control research, policy and practice. *Tob Control*. 2012 Mar;21(2):236-43. doi: 10.1136/tobaccocontrol-2011-050280.
- Amr S, Wolpert B, Loffredo CA, Zheng YL, Shields PG, Jones R, Harris CC. Occupation, gender, race, and lung cancer. *J Occup Environ Med*. 2008 Oct;50(10):1167-75. doi: 10.1097/JOM.0b013e31817d3639.
- Bain C, Feskanich D, Speizer FE, Thun M, Hertzmark E, Rosner BA, Colditz GA. Lung cancer rates in men and women with comparable histories of smoking. *J Natl Cancer Inst*. 2004 Jun 2;96(11):826-34.
- Barnes-Ellerbe S, Knudsen KE, Puga A. 2,3,7,8-Tetrachlorodibenzo-p-dioxin blocks androgen-dependent cell proliferation of LNCaP cells through modulation of pRB phosphorylation. *Mol Pharmacol*. 2004 Sep;66(3):502-11.
- Bartsch H, Castegnaro M, Rojas M, Camus AM, Alexandrov K, Lang M. Expression of pulmonary cytochrome P4501A1 and carcinogen DNA adduct formation in high risk subjects for tobacco-related lung cancer. *Toxicol Lett*. 1992 Dec;64-65 Spec No:477-83.
- Bartsch H, Petruzzelli S, De Flora S, Hietanen E, Camus AM, Castegnaro M, Alexandrov K, Rojas M, Saracci R, Giuntini C. Carcinogen metabolism in human lung tissues and the effect of tobacco smoking: results from a case—control multicenter study on lung cancer patients. *Environ Health Perspect*. 1992 Nov;98:119-24.
- Bauer M, Herbarth O, Rudzok S, Schmücking E, Müller A, Aust G, Gräbsch C. Diversity of common alternative splicing variants of human cytochrome P450 1A1 and their association to carcinogenesis. *Int J Oncol*. 2007 Jul;31(1):211-8.
- Benedetti S, Nuvoli B, Catalani S, Galati R. Reactive oxygen species a double-edged sword for mesothelioma. *Oncotarget*. 2015 Jul 10;6(19):16848-65.
- Benhamou S, Lee WJ, Alexandrie AK, Boffetta P, Bouchardy C, Butkiewicz D, Brockmüller J, Clapper ML, Daly A, Dolzan V, Ford J, Gaspari L, Haugen A, Hirvonen A, Husgafvel-Pursiainen K, Ingelman-Sundberg M, Kalina I, Kihara M, Kremers P, Le Marchand L, London SJ, Nazar-Stewart V, Onon-Kihara M, Rannug A, Romkes M, Ryberg D, Seidegard J, Shields P, Strange RC, Stücker I, To-Figueras J, Brennan P, Taioli E. Meta- and pooled analyses of the effects of glutathione S-transferase M1 polymorphisms and smoking on lung cancer risk. *Carcinogenesis*. 2002 Aug;23(8):1343-50.
- Benninghoff AD, Williams DE. The role of estrogen receptor β in transplacental cancer prevention by indole-3-carbinol. *Cancer Prev Res (Phila)*. 2013 Apr;6(4):339-48. doi: 10.1158/1940-6207.CAPR-12-0311.
- Berge G, Mollerup S, Øvrebø S, Hewer A, Phillips DH, Eilertsen E, Haugen A. Role of estrogen receptor in regulation of polycyclic aromatic hydrocarbon metabolic activation in lung. *Lung Cancer*. 2004 Sep;45(3):289-97.

- Bilano V, Gilmour S, Moffiet T, d'Espaignet ET, Stevens GA, Commar A, Tuyl F, Hudson I, Shibuya K. Global trends and projections for tobacco use, 1990-2025: an analysis of smoking indicators from the WHO Comprehensive Information Systems for Tobacco Control. *Lancet*. 2015 Mar 14;385(9972):966-76. doi: 10.1016/S0140-6736(15)60264-1.
- Boffetta P. Human cancer from environmental pollutants: the epidemiological evidence. *Mutat Res*. 2006 Sep 28;608(2):157-62.
- Bollati V, Baccarelli A. Environmental epigenetics. *Heredity (Edinb)*. 2010 Jul;105(1):105-12. doi: 10.1038/hdy.2010.2.
- Boström CE, Gerde P, Hanberg A, Jernström B, Johansson C, Kyrklund T, Rannug A, Törnqvist M, Victorin K, Westerholm R. Cancer risk assessment, indicators, and guidelines for polycyclic aromatic hydrocarbons in the ambient air. *Environ Health Perspect*. 2002 Jun;110 Suppl 3:451-88.
- Bouchardy C, Benhamou S, Schaffar R, Verkooijen HM, Fioretta G, Schubert H, Vinh-Hung V, Soria JC, Vlastos G, Rapiti E. Lung cancer mortality risk among breast cancer patients treated with anti-estrogens. *Cancer*. 2011 Mar 15;117(6):1288-95. doi: 10.1002/ncr.25638.
- Bräuner EV, Andersen CE, Sørensen M, Andersen ZJ, Gravesen P, Ulbak K, Hertel O, Pedersen C, Overvad K, Tjønneland A, Raaschou-Nielsen O. Residential radon and lung cancer incidence in a Danish cohort. *Environ Res*. 2012 Oct;118:130-6. doi: 10.1016/j.envres.2012.05.012.
- Bray F, Sankila R, Ferlay J, Parkin DM. Estimates of cancer incidence and mortality in Europe in 1995. *Eur J Cancer*. 2002 Jan;38(1):99-166.
- Bray F, Ren JS, Masuyer E, Ferlay J. Global estimates of cancer prevalence for 27 sites in the adult population in 2008. *Int J Cancer*. 2013 Mar 1;132(5):1133-45. doi: 10.1002/ijc.27711.
- Brena RM, Costello JF. Genome-epigenome interactions in cancer. *Hum Mol Genet*. 2007 Apr 15;16 Spec No 1:R96-105.
- Brzezińska E, Dutkowska A, Antczak A. The significance of epigenetic alterations in lung carcinogenesis. *Mol Biol Rep*. 2013 Jan;40(1):309-25. doi:10.1007/s11033-012-2063-4.
- Bueno MJ, Pérez de Castro I, Gómez de Cedrón M, Santos J, Calin GA, Cigudosa JC, Croce CM, Fernández-Piqueras J, Malumbres M. Genetic and epigenetic silencing of microRNA-203 enhances ABL1 and BCR-ABL1 oncogene expression. *Cancer Cell*. 2008 Jun;13(6):496-506. doi: 10.1016/j.ccr.2008.04.018.
- Burns TF, Stabile LP. Targeting the estrogen pathway for the treatment and prevention of lung cancer. *Lung Cancer Manag*. 2014 Feb 1;3(1):43-52.
- Buters JT, Mahadevan B, Quintanilla-Martinez L, Gonzalez FJ, Greim H, Baird WM, Luch A. Cytochrome P450 1B1 determines susceptibility to dibenzo[a,l]pyrene-induced tumor formation. *Chem Res Toxicol*. 2002 Sep;15(9):1127-35.
- Butterworth BE, Popp JA, Conolly RB, Goldsworthy TL. Chemically induced cell proliferation in carcinogenesis. *IARC Sci Publ*. 1992;(116):279-305.
- Calin GA, Croce CM. MicroRNA signatures in human cancers. *Nat Rev Cancer*. 2006 Nov;6(11):857-66.

- Carey MA, Card JW, Voltz JW, Arbes SJ Jr, Germolec DR, Korach KS, Zeldin DC. It's all about sex: gender, lung development and lung disease. *Trends Endocrinol Metab.* 2007 Oct;18(8):308-13.
- Cavalieri E, Frenkel K, Liehr JG, Rogan E, Roy D. Estrogens as endogenous genotoxic agents--DNA adducts and mutations. *J Natl Cancer Inst Monogr.* 2000;(27):75-93.
- Chang C, Lee SO, Yeh S, Chang TM. Androgen receptor (AR) differential roles in hormone-related tumors including prostate, bladder, kidney, lung, breast and liver. *Oncogene.* 2014 Jun 19;33(25):3225-34. doi: 10.1038/onc.2013.274.
- Chiba T, Uchi H, Yasukawa F, Furue M. Role of the arylhydrocarbon receptor in lung disease. *Int Arch Allergy Immunol.* 2011;155 Suppl 1:129-34. doi:10.1159/000327499.
- Chlebowski RT, Schwartz AG, Wakelee H, Anderson GL, Stefanick ML, Manson JE, Rodabough RJ, Chien JW, Wactawski-Wende J, Gass M, Kotchen JM, Johnson KC, O'Sullivan MJ, Ockene JK, Chen C, Hubbell FA; Women's Health Initiative Investigators. Oestrogen plus progestin and lung cancer in postmenopausal women (Women's Health Initiative trial): a post-hoc analysis of a randomised controlled trial. *Lancet.* 2009 Oct 10;374(9697):1243-51. doi: 10.1016/S0140-6736(09)61526-9.
- Choi JE, Kang HG, Jang JS, Choi YY, Kim MJ, Kim JS, Jeon HS, Lee WK, Cha SI, Kim CH, Kam S, Jung TH, Park JY. Polymorphisms in telomere maintenance genes and risk of lung cancer. *Cancer Epidemiol Biomarkers Prev.* 2009 Oct;18(10):2773-81. doi: 10.1158/1055-9965.EPI-09-0323.
- Chuang CY, Chang H, Lin P, Sun SJ, Chen PH, Lin YY, Sheu GT, Ko JL, Hsu SL, Chang JT. Up-regulation of osteopontin expression by aryl hydrocarbon receptor via both ligand-dependent and ligand-independent pathways in lung cancer. *Gene.* 2012 Jan 15;492(1):262-9. doi: 10.1016/j.gene.2011.10.019.
- Ciana P, Di Luccio G, Belcredito S, Pollio G, Vegeto E, Tatangelo L, Tiveron C, Maggi A. Engineering of a mouse for the in vivo profiling of estrogen receptor activity. *Mol Endocrinol.* 2001 Jul;15(7):1104-13.
- Conney AH. Induction of microsomal enzymes by foreign chemicals and carcinogenesis by polycyclic aromatic hydrocarbons: G. H. A. Clowes Memorial Lecture. *Cancer Res.* 1982 Dec;42(12):4875-917.
- D'Alessio AC, Szyf M. Epigenetic tête-à-tête: the bilateral relationship between chromatin modifications and DNA methylation. *Biochem Cell Biol.* 2006 Aug;84(4):463-76.
- Davey RA, Grossmann M. Androgen Receptor Structure, Function and Biology: From Bench to Bedside. *Clin Biochem Rev.* 2016 Feb;37(1):3-15.
- De Matteis S, Consonni D, Pesatori AC, Bergen AW, Bertazzi PA, Caporaso NE, Lubin JH, Wacholder S, Landi MT. Are women who smoke at higher risk for lung cancer than men who smoke? *Am J Epidemiol.* 2013 Apr 1;177(7):601-12. doi:10.1093/aje/kws445.
- Denison MS, Nagy SR. Activation of the aryl hydrocarbon receptor by structurally diverse exogenous and endogenous chemicals. *Annu Rev Pharmacol Toxicol.* 2003;43:309-34.
- Deutsch-Wenzel RP, Brune H, Grimmer G, Dettbarn G, Misfeld J. Experimental studies in rat lungs on the carcinogenicity and dose-response relationships of eight frequently occurring environmental polycyclic aromatic hydrocarbons. *J Natl Cancer Inst.* 1983 Sep;71(3):539-44.
- Dewing P, Shi T, Horvath S, Vilain E. Sexually dimorphic gene expression in mouse brain precedes gonadal differentiation. *Brain Res Mol Brain Res.* 2003 Oct 21;118(1-2):82-90.

- Dipple A. DNA adducts of chemical carcinogens. *Carcinogenesis*. 1995 Mar;16(3):437-41.
- Divi RL, Lindeman TL, Shockley ME, Keshava C, Weston A, Poirier MC. Correlation between CYP1A1 transcript, protein level, enzyme activity and DNA adduct formation in normal human mammary epithelial cell strains exposed to benzo[a]pyrene. *Mutagenesis*. 2014 Nov;29(6):409-17. doi: 10.1093/mutage/geu049.
- Dong H, Bonala RR, Suzuki N, Johnson F, Grollman AP, Shibutani S. Mutagenic potential of benzo[a]pyrene-derived DNA adducts positioned in codon 273 of the human P53 gene. *Biochemistry*. 2004 Dec 21;43(50):15922-8.
- Dougherty SM, Mazhawidza W, Bohn AR, Robinson KA, Mattingly KA, Blankenship KA, Huff MO, McGregor WG, Klinge CM. Gender difference in the activity but not expression of estrogen receptors alpha and beta in human lung adenocarcinoma cells. *Endocr Relat Cancer*. 2006 Mar;13(1):113-34.
- DuSell CD, Nelson ER, Wittmann BM, Fretz JA, Kazmin D, Thomas RS, Pike JW, McDonnell DP. Regulation of aryl hydrocarbon receptor function by selective estrogen receptor modulators. *Mol Endocrinol*. 2010 Jan;24(1):33-46. doi: 10.1210/me.2009-0339.
- Ehrlich M. DNA methylation in cancer: too much, but also too little. *Oncogene*. 2002 Aug 12;21(35):5400-13.
- Esser C, Rannug A. The aryl hydrocarbon receptor in barrier organ physiology, immunology, and toxicology. *Pharmacol Rev*. 2015;67(2):259-79. doi:10.1124/pr.114.009001.
- Ferlay J, Soerjomataram I, Dikshit R, Eser S, Mathers C, Rebelo M, Parkin DM, Forman D, Bray F. Cancer incidence and mortality worldwide: sources, methods and major patterns in GLOBOCAN 2012. *Int J Cancer*. 2015 Mar 1;136(5):E359-86. doi: 10.1002/ijc.29210.
- Ferreccio C, Yuan Y, Calle J, Benítez H, Parra RL, Acevedo J, Smith AH, Liaw J, Steinmaus C. Arsenic, tobacco smoke, and occupation: associations of multiple agents with lung and bladder cancer. *Epidemiology*. 2013 Nov;24(6):898-905. doi:10.1097/EDE.0b013e31829e3e03.
- Ferroa P, Catalanob M.G, Dell'Eva R., Fortunatib N., Pfeffera U.,The androgen receptor CAG repeat: a modifier of carcinogenesis. Laboratory of Molecular Biology, National Cancer Research Institute, Largo R. Benzi 10, 16132 Genoa, Italy. *Molecular and Cellular Endocrinology*. 2002 July Vol. 193, Issues 1–2, Pages 109–120.
- Fitzmaurice C, Dicker D, Pain A, Hamavid H, Moradi-Lakeh M, MacIntyre MF, Allen C, Hansen G, Woodbrook R, Wolfe C, Hamadeh RR, Moore A, Werdecker A, Gessner BD, Te Ao B, McMahan B, Karimkhani C, Yu C, Cooke GS, Schwebel DC, Carpenter DO, Pereira DM, Nash D, Kazi DS, De Leo D, Plass D, Ukwaja KN, Thurston GD, Yun Jin K, Simard EP, Mills E, Park EK, Catalá-López F, deVeber G, Gotay C, Khan G, Hosgood HD 3rd, Santos IS, Leasher JL, Singh J, Leigh J, Jonas J, Sanabria J, Beardsley J, Jacobsen KH, Takahashi K, Franklin RC, Ronfani L, Montico M, Naldi L, Tonelli M, Geleijnse J, Petzold M, Shrimme MG, Younis M, Yonemoto N, Breitborde N, Yip P, Pourmalek F, Lotufo PA, Esteghamati A, Hankey GJ, Ali R, Lunevicius R, Malekzadeh R, Dellavalle R, Weintraub R, Lucas R, Hay R, Rojas-Rueda D, Westerman R, Sepanlou SG, Nolte S, Patten S, Weichenthal S, Abera SF, Fereshtehnejad SM, Shiue I, Driscoll T, Vasankari T, Alsharif U, Rahimi-Movaghar V, Vlassov VV, Marcenes WS, Mekonnen W, Melaku YA, Yano Y, Artaman A, Campos I, MacLachlan J, Mueller U, Kim D, Trillini M, Eshrati B, Williams HC, Shibuya K, Dandona R, Murthy K, Cowie B, Amare AT, Antonio CA, Castañeda-Orjuela C, van Gool CH, Violante F, Oh IH, Deribe K, Soreide K, Knibbs L, Kereselidze M, Green M, Cardenas R, Roy N, Tillman T, Li Y, Krueger H, Monasta L, Dey S, Sheikhbahaei S, Hafezi-Nejad N, Kumar GA, Sreeramareddy CT, Dandona L, Wang H, Vollset SE, Mokdad A, Salomon JA, Lozano R, Vos T, Forouzanfar M, Lopez A, Murray C, Naghavi

M. Global Burden of Disease Cancer Collaboration The Global Burden of Cancer 2013. *JAMA Oncol.* 2015 Jul;1(4):505-27. doi: 10.1001/jamaoncol.2015.0735.

Gao YT, Blot WJ, Zheng W, Ershow AG, Hsu CW, Levin LI, Zhang R, Fraumeni JF Jr. Lung cancer among Chinese women. *Int J Cancer.* 1987 Nov 15;40(5):604-9.

Ganti AK, Sahmoun AE, Panwalkar AW, Tendulkar KK, Potti A. Hormone replacement therapy is associated with decreased survival in women with lung cancer. *J Clin Oncol.* 2006 Jan 1;24(1):59-63.

Gautier JC, Lecoecur S, Cosme J, Perret A, Urban P, Beaune P, Pompon D. Contribution of human cytochrome P450 to benzo[a]pyrene and benzo[a]pyrene-7,8-dihydrodiol metabolism, as predicted from heterologous expression in yeast. *Pharmacogenetics.* 1996 Dec;6(6):489-99.

Gelboin HV. Benzo[alpha]pyrene metabolism, activation and carcinogenesis: role and regulation of mixed-function oxidases and related enzymes. *Physiol Rev.* 1980 Oct;60(4):1107-66.

Glozak MA, Seto E. Histone deacetylases and cancer. *Oncogene.* 2007 Aug 13;26(37):5420-32.

Godschalk RW, Moonen EJ, Schilderman PA, Broekmans WM, Kleijnans JC, Van Schooten FJ. Exposure-route-dependent DNA adduct formation by polycyclic aromatic hydrocarbons. *Carcinogenesis.* 2000 Jan;21(1):87-92.

Greenblatt MS, Bennett WP, Hollstein M, Harris CC. Mutations in the p53 tumor suppressor gene: clues to cancer etiology and molecular pathogenesis. *Cancer Res.* 1994 Sep 15;54(18):4855-78.

Grimmer G, Brune H, Dettbarn G, Naujack KW, Mohr U, Wenzel-Hartung R. Contribution of polycyclic aromatic compounds to the carcinogenicity of sidestream smoke of cigarettes evaluated by implantation into the lungs of rats. *Cancer Lett.* 1988 Dec 15;43(3):173-7.

Guibert S, Weber M. Functions of DNA methylation and hydroxymethylation in mammalian development. *Curr Top Dev Biol.* 2013;104:47-83. doi:10.1016/B978-0-12-416027-9.00002-4.

Han W, Pentecost BT, Pietropaolo RL, Fasco MJ, Spivack SD. Estrogen receptor alpha increases basal and cigarette smoke extract-induced expression of CYP1A1 and CYP1B1, but not GSTP1, in normal human bronchial epithelial cells. *Mol Carcinog.* 2005 Nov;44(3):202-11.

Hanahan D, Weinberg RA. The hallmarks of cancer. *Cell.* 2000 Jan 7;100(1):57-70.

Hanahan D, Weinberg RA. Hallmarks of cancer: the next generation. *Cell.* 2011 Mar 4;144(5):646-74. doi: 10.1016/j.cell.2011.02.013.

Harper TA Jr, Morré J, Lauer FT, McQuistan TJ, Hummel JM, Burchiel SW, Williams DE. Analysis of dibenzo[def,p]chrysene-deoxyadenosine adducts in wild-type and cytochrome P450 1b1 knockout mice using stable-isotope dilution UHPLC-MS/MS. *Mutat Res Genet Toxicol Environ Mutagen.* 2015 Apr;782:51-6. doi:10.1016/j.mrgentox.2015.03.007.

Harris CC. Chemical and physical carcinogenesis: advances and perspectives for the 1990s. *Cancer Res.* 1991 Sep 15;51(18 Suppl):5023s-5044s.

Hashim D, Boffetta P. Occupational and environmental exposures and cancers in developing countries. *Ann Glob Health.* 2014 Sep-Oct;80(5):393-411. doi: 10.1016/j.aogh.2014.10.002.

Haugen A. Women who smoke: are women more susceptible to tobacco-induced lung cancer? *Carcinogenesis.* 2002 Feb;23(2):227-9.

- Hawighorst T. [Angiogenesis, lymphangiogenesis, and tumor progression]. *Zentralbl Gynakol.* 2002 Nov;124(11):497-505.
- Hecht SS. Tobacco smoke carcinogens and lung cancer. *J Natl Cancer Inst.* 1999 Jul 21;91(14):1194-210.
- Hecht SS. Tobacco carcinogens, their biomarkers and tobacco-induced cancer. *Nat Rev Cancer.* 2003 Oct;3(10):733-44.
- Heldring N, Pike A, Andersson S, Matthews J, Cheng G, Hartman J, Tujague M, Ström A, Treuter E, Warner M, Gustafsson JA. Estrogen receptors: how do they signal and what are their targets. *Physiol Rev.* 2007 Jul;87(3):905-31.
- Henschke CI, Yip R, Miettinen OS. Women's susceptibility to tobacco carcinogens and survival after diagnosis of lung cancer. International Early Lung Cancer Action Program Investigators, *JAMA.* 2006 Jul 12;296(2):180-4.
- Herman JG, Baylin SB. Gene silencing in cancer in association with promoter hypermethylation. *N Engl J Med.* 2003 Nov 20;349(21):2042-54.
- Hershberger PA, Vasquez AC, Kanterewicz B, Land S, Siegfried JM, Nichols M. Regulation of endogenous gene expression in human non-small cell lung cancer cells by estrogen receptor ligands. *Cancer Res.* 2005 Feb 15;65(4):1598-605.
- Hesterberg TW, Long CM, Bunn WB, Lapin CA, McClellan RO, Valberg PA. Health effects research and regulation of diesel exhaust: an historical overview focused on lung cancer risk. *Inhal Toxicol.* 2012 Jun;24 Suppl 1:1-45. doi:10.3109/08958378.2012.691913.
- Hitchman SC, Fong GT. Gender empowerment and female-to-male smoking prevalence ratios. *Bull World Health Organ.* 2011 Mar 1;89(3):195-202. doi:10.2471/BLT.10.079905.
- Hodek P, Koblihová J, Kizek R, Frei E, Arlt VM, Stiborová M. The relationship between DNA adduct formation by benzo[a]pyrene and expression of its activation enzyme cytochrome P450 1A1 in rat. *Environ Toxicol Pharmacol.* 2013 Nov;36(3):989-96. doi: 10.1016/j.etap.2013.09.004.
- Hosgood HD 3rd, Cawthon R, He X, Chanock S, Lan Q. Genetic variation in telomere maintenance genes, telomere length, and lung cancer susceptibility. *Lung Cancer.* 2009 Nov;66(2):157-61. doi: 10.1016/j.lungcan.2009.02.005.
- Hubaux R, Becker-Santos DD, Enfield KS, Lam S, Lam WL, Martinez VD. Arsenic, asbestos and radon: emerging players in lung tumorigenesis. *Environ Health.* 2012 Nov 22;11:89. doi: 10.1186/1476-069X-11-89.
- Hukkanen J, Pelkonen O, Hakkola J, Raunio H. Expression and regulation of xenobiotic-metabolizing cytochrome P450 (CYP) enzymes in human lung. *Crit Rev Toxicol.* 2002 Sep;32(5):391-411.
- Imataka H, Sogawa K, Yasumoto K, Kikuchi Y, Sasano K, Kobayashi A, Hayami M, Fujii-Kuriyama Y. Two regulatory proteins that bind to the basic transcription element (BTE), a GC box sequence in the promoter region of the rat P-4501A1 gene. *EMBO J.* 1992 Oct;11(10):3663-71.
- International Agency for Research on Cancer, IARC Monographs on the Evaluation of the Carcinogenic Risk of Chemicals in Humans, Polynuclear aromatic compounds, Part 1, Chemical Environmental and Experimental Data. IARC Press, Lyon 1983; Vol 32.

International Agency for Research on Cancer, IARC Monographs on the Evaluation of the Carcinogenic Risks to Humans. Man-Made Mineral Fibers and Radon. IARC Press, Lyon 1988; Vol 43.

International Agency for Research on Cancer, IARC Monographs on the Synergistic Carcinogenic Effects of Tobacco Smoke and Other Carcinogens. IARC Press, Lyon 2004; Vol 83.

International Agency for Research on Cancer, IARC Monographs on the Evaluation of Carcinogenic Risks to Humans, Tobacco Smoke and Involuntary Smoking. IARC Press, Lyon 2004; Vol 83

Ivanova MM, Mazhawidza W, Dougherty SM, Minna JD, Klinge CM. Activity and intracellular location of estrogen receptors alpha and beta in human bronchial epithelial cells. *Mol Cell Endocrinol*. 2009 Jun 16;305(1-2):12-21. doi:10.1016/j.mce.2009.01.021.

Ivanova MM, Mazhawidza W, Dougherty SM, Klinge CM. Sex differences in estrogen receptor subcellular location and activity in lung adenocarcinoma cells. *Am J Respir Cell Mol Biol*. 2010 Mar;42(3):320-30. doi: 10.1165/rcmb.2009-0059OC.

Izzotti A, Calin GA, Arrigo P, Steele VE, Croce CM, De Flora S. Downregulation of microRNA expression in the lungs of rats exposed to cigarette smoke. *FASEB J*. 2009 Mar;23(3):806-12. doi: 10.1096/fj.08-121384.

Jana NR, Sarkar S, Ishizuka M, Yonemoto J, Tohyama C, Sone H. Cross-talk between 2,3,7,8-tetrachlorodibenzo-p-dioxin and testosterone signal transduction pathways in LNCaP prostate cancer cells. *Biochem Biophys Res Commun*. 1999 Mar 24;256(3):462-8.

Jana NR, Sarkar S, Ishizuka M, Yonemoto J, Tohyama C, Sone H. Role of estradiol receptor-alpha in differential expression of 2,3,7,8-tetrachlorodibenzo-p-dioxin-inducible genes in the RL95-2 and KLE human endometrial cancer cell lines. *Arch Biochem Biophys*. 1999 Aug 1;368(1):31-9.

Jelinic P, Shaw P. Loss of imprinting and cancer. *J Pathol*. 2007 Feb;211(3):261-8.

Jemal A, Thun MJ, Ries LA, Howe HL, Weir HK, Center MM, Ward E, Wu XC, Ehemann C, Anderson R, Ajani UA, Kohler B, Edwards BK. Annual report to the nation on the status of cancer, 1975-2005, featuring trends in lung cancer, tobacco use, and tobacco control. *J Natl Cancer Inst*. 2008 Dec 3;100(23):1672-94. doi:10.1093/jnci/djn389.

Jensen E V, Jacobson H I. 1960. Fate of steroid estrogens in target tissues. In *Biological Activities of Steroids in Relation to Cancer*, ed G Pincus, E Vollmer pp.161 - 74.1960, New York: Academic.

Johnsen AR, Wick LY, Harms H. Principles of microbial PAH-degradation in soil. *Environ Pollut*. 2005 Jan;133(1):71-84.

Jorgensen JS, Nilson JH. AR suppresses transcription of the LHBeta subunit by interacting with steroidogenic factor-1. *Mol Endocrinol*. 2001 Sep;15(9):1505-16.

Kadzhoian AV, Shevchenko AI. Effect of high dose tamoxifen therapy in conservative treatment of patients with III-IV stages of non-small cell lung cancer. *Lik Sprava*. 2014 Jan-Feb;(1-2):103-10.

Kaiser U, Hofmann J, Schilli M, Wegmann B, Klotz U, Wedel S, Virmani AK, Wollmer E, Branscheid D, Gazdar AF, Havemann K. Steroid-hormone receptors in cell lines and tumor biopsies of human lung cancer. *Int J Cancer*. 1996 Jul 29;67(3):357-64.

Kawai H, Ishii A, Washiya K, Konno T, Kon H, Yamaya C, Ono I, Minamiya Y, Ogawa J. Estrogen receptor alpha and beta are prognostic factors in non-small cell lung cancer. *Clin Cancer Res*. 2005 Jul 15;11(14):5084-9.

- Kawai H. Estrogen receptors as the novel therapeutic biomarker in non-small cell lung cancer. *World J Clin Oncol*. 2014 Dec 10;5(5):1020-7. doi: 10.5306/wjco.v5.i5.1020.
- Kim JH, Sherman ME, Curriero FC, Guengerich FP, Strickland PT, Sutter TR. Expression of cytochromes P450 1A1 and 1B1 in human lung from smokers, non-smokers, and ex-smokers. *Toxicol Appl Pharmacol*. 2004 Sep 15;199(3):210-9.
- Kiyohara C, Ohno Y. Sex differences in lung cancer susceptibility: a review. *Gend Med*. 2010 Oct;7(5):381-401. doi: 10.1016/j.genm.2010.10.002.
- Kizu R, Okamura K, Toriba A, Kakishima H, Mizokami A, Burnstein KL, Hayakawa K. A role of aryl hydrocarbon receptor in the antiandrogenic effects of polycyclic aromatic hydrocarbons in LNCaP human prostate carcinoma cells. *Arch Toxicol*. 2003 Jun;77(6):335-43.
- Kleiner HE, Vulimiri SV, Hatten WB, Reed MJ, Nebert DW, Jefcoate CR, DiGiovanni J. Role of cytochrome p4501 family members in the metabolic activation of polycyclic aromatic hydrocarbons in mouse epidermis. *Chem Res Toxicol*. 2004 Dec;17(12):1667-74.
- Koch DC, Jang HS, O'Donnell EF, Punj S, Koppurapu PR, Bisson WH, Kerkvliet NI, Kolluri SK. Anti-androgen flutamide suppresses hepatocellular carcinoma cell proliferation via the aryl hydrocarbon receptor mediated induction of transforming growth factor- β 1. *Oncogene*. 2015 Dec 10;34(50):6092-104. doi:10.1038/onc.2015.55.
- Kojima M, Sekimoto M, Degawa M. A novel gender-related difference in the constitutive expression of hepatic cytochrome P4501A subfamily enzymes in Meishan pigs. *Biochem Pharmacol*. 2008 Mar 1;75(5):1076-82.
- Koo LC, Ho JH. Worldwide epidemiological patterns of lung cancer in nonsmokers. *Int J Epidemiol*. 1990;19 Suppl 1:S14-23.
- Kouri RE, McKinney CE, Slomiany DJ, Snodgrass DR, Wray NP, McLemore TL. Positive correlation between high aryl hydrocarbon hydroxylase activity and primary lung cancer as analyzed in cryopreserved lymphocytes. *Cancer Res*. 1982 Dec;42(12):5030-7.
- Kouzarides T. Chromatin modifications and their function. *Cell*. 2007 Feb 23;128(4):693-705.
- Kreuzer M, Heinrich J, Kreienbrock L, Rosario AS, Gerken M, Wichmann HE. Risk factors for lung cancer among nonsmoking women. *Int J Cancer*. 2002 Aug 20;100(6):706-13.
- Kuiper GG, Enmark E, Peltö-Huikko M, Nilsson S, Gustafsson JA. Cloning of a novel receptor expressed in rat prostate and ovary. *Proc Natl Acad Sci U S A*. 1996 Jun 11;93(12):5925-30.
- Kwekel JC, Desai VG, Moland CL, Vijay V, Fuscoe JC. Sex differences in kidney gene expression during the life cycle of F344 rats. *Biol Sex Differ*. 2013 Jul 31;4(1):14. doi: 10.1186/2042-6410-4-14.
- Kyrtopoulos SA. Biomarkers in environmental carcinogenesis research: striving for a new momentum. *Toxicol Lett*. 2006 Mar 15;162(1):3-15.
- Laaksonen M, Rahkonen O, Karvonen S, Lahelma E. Socioeconomic status and smoking: analysing inequalities with multiple indicators. *Eur J Public Health*. 2005 Jun;15(3):262-9.
- Langevin SM, Kratzke RA, Kelsey KT. Epigenetics of lung cancer. *Transl Res*. 2015 Jan;165(1):74-90. doi: 10.1016/j.trsl.2014.03.001.

- Lawless MW, O'Byrne KJ, Gray SG. Oxidative stress induced lung cancer and COPD: opportunities for epigenetic therapy. *J Cell Mol Med*. 2009 Sep;13(9A):2800-21. doi: 10.1111/j.1582-4934.2009.00845.x. Review.
- Lewis DF. Homology modelling of human cytochromes P450 involved in xenobiotic metabolism and rationalization of substrate selectivity. *Exp Toxicol Pathol*. 1999 Jul;51(4-5):369-74.
- Lin P, Chang JT, Ko JL, Liao SH, Lo WS. Reduction of androgen receptor expression by benzo[alpha]pyrene and 7,8-dihydro-9,10-epoxy-7,8,9,10-tetrahydrobenzo[alpha]pyrene in human lung cells. *Biochem Pharmacol*. 2004 Apr 15;67(8):1523-30.
- Lin RT, Takahashi K, Karjalainen A, Hoshuyama T, Wilson D, Kameda T, Chan CC, Wen CP, Furuya S, Higashi T, Chien LC, Ohtaki M. Ecological association between asbestos-related diseases and historical asbestos consumption: an international analysis. *Lancet*. 2007 Mar 10;369(9564):844-9.
- Lin TM, Ko K, Moore RW, Simanainen U, Oberley TD, Peterson RE. Effects of aryl hydrocarbon receptor null mutation and in utero and lactational 2,3,7,8-tetrachlorodibenzo-p-dioxin exposure on prostate and seminal vesicle development in C57BL/6 mice. *Toxicol Sci*. 2002 Aug;68(2):479-87.
- Liu MM, Albanese C, Anderson CM, Hilty K, Webb P, Uht RM, Price RH Jr, Pestel RG, Kushner PJ. Opposing action of estrogen receptors alpha and beta on cyclin D1 gene expression. *J Biol Chem*. 2002 Jul 5;277(27):24353-60.
- London SJ, Yuan JM, Coetzee GA, Gao YT, Ross RK, Yu MC. CYP1A1 I462V genetic polymorphism and lung cancer risk in a cohort of men in Shanghai, China. *Cancer Epidemiol Biomarkers Prev*. 2000 Sep;9(9):987-91.
- Lothar SA, Harding GA, Musto G, Navaratnam S, Pitz MW. Antiestrogen use and survival of women with non-small cell lung cancer in Manitoba, Canada. *Horm Cancer*. 2013 Oct;4(5):270-6. doi: 10.1007/s12672-013-0149-7.
- Lubin JH, Boice JD Jr, Edling C, Hornung RW, Howe GR, Kunz E, Kusiak RA, Morrison HI, Radford EP, Samet JM, et al. Lung cancer in radon-exposed miners and estimation of risk from indoor exposure. *J Natl Cancer Inst*. 1995 Jun 7;87(11):817-27.
- Lutz WK. A true threshold dose in chemical carcinogenesis cannot be defined for a population, irrespective of the mode of action. *Hum Exp Toxicol*. 2000 Oct;19(10):566-8.
- Ma H, Zhou Z, Wei S, Liu Z, Pooley KA, Dunning AM, Svenson U, Roos G, Hosgood HD 3rd, Shen M, Wei Q. Shortened telomere length is associated with increased risk of cancer: a meta-analysis. *PLoS One*. 2011;6(6):e20466. doi:10.1371/journal.pone.0020466.
- Mačak-Šafranko Z, Sobočanec S, Sarić A, Balog T, Sverko V, Kušić B, Marotti T. Cytochrome P450 gender-related differences in response to hyperoxia in young CBA mice. *Exp Toxicol Pathol*. 2011 May;63(4):345-50. doi: 10.1016/j.etp.2010.02.009.
- Mah V, Seligson DB, Li A, Márquez DC, Wistuba II, Elshimali Y, Fishbein MC, Chia D, Pietras RJ, Goodglick L. Aromatase expression predicts survival in women with early-stage non small cell lung cancer. *Cancer Res*. 2007 Nov 1;67(21):10484-90.
- Markowitz S. Asbestos-related lung cancer and malignant mesothelioma of the pleura: selected current issues. *Semin Respir Crit Care Med*. 2015 Jun;36(3):334-46. doi: 10.1055/s-0035-1549449.
- Márquez-Garbán DC, Chen HW, Fishbein MC, Goodglick L, Pietras RJ. Estrogen receptor signaling pathways in human non-small cell lung cancer. *Steroids*. 2007 Feb;72(2):135-43.

- Márquez-Garbán DC, Chen HW, Goodglick L, Fishbein MC, Pietras RJ. Targeting aromatase and estrogen signaling in human non-small cell lung cancer. *Ann N Y Acad Sci.* 2009 Feb;1155:194-205. doi: 10.1111/j.1749-6632.2009.04116.x.
- Marshall AL, Christiani DC. Genetic susceptibility to lung cancer--light at the end of the tunnel? *Carcinogenesis.* 2013 Mar;34(3):487-502. doi:10.1093/carcin/bgt016.
- Martinez VD, Vucic EA, Becker-Santos DD, Gil L, Lam WL. Arsenic exposure and the induction of human cancers. *J Toxicol.* 2011;2011:431287. doi:10.1155/2011/431287.
- Massaro D, Massaro GD. Estrogen receptor regulation of pulmonary alveolar dimensions: alveolar sexual dimorphism in mice. *Am J Physiol Lung Cell Mol Physiol.* 2006 May;290(5):L866-70.
- Matthews J, Wihlén B, Heldring N, MacPherson L, Helguero L, Treuter E, Haldosén L-A, Gustafsson J-Å. Co-planar 3,3',4,4',5-pentachlorinated biphenyl and non-co-planar 2,2',4,6,6'-pentachlorinated biphenyl differentially induce recruitment of oestrogen receptor α to aryl hydrocarbon receptor target genes *Biochem J.* 2007 September 1; 406(Pt 2): 343–353. doi: 10.1042/BJ20070585.
- McFadyen MC, Murray GI. Cytochrome P450 1B1: a novel anticancer therapeutic target. *Future Oncol.* 2005 Apr;1(2):259-63.
- McKay JD, Hung RJ, Gaborieau V, Boffetta P, Chabrier A, Byrnes G, Zaridze D, Mukeria A, Szeszenia-Dabrowska N, Lissowska J, Rudnai P, Fabianova E, Mates D, Bencko V, Foretova L, Janout V, McLaughlin J, Shepherd F, Montpetit A, Narod S, Krokan HE, Skorpen F, Elvestad MB, Vatten L, Njølstad I, Axelsson T, Chen C, Goodman G, Barnett M, Loomis MM, Lubiński J, Matyjasik J, Lener M, Oszutowska D, Field J, Liloglou T, Xinarianos G, Cassidy A; EPIC Study, Vineis P, Clavel-Chapelon F, Palli D, Tumino R, Krogh V, Panico S, González CA, Ramón Quirós J, Martínez C, Navarro C, Ardanaz E, Larrañaga N, Kham KT, Key T, Bueno-de-Mesquita HB, Peeters PH, Trichopoulou A, Linseisen J, Boeing H, Hallmans G, Overvad K, Tjønneland A, Kumle M, Riboli E, Zelenika D, Boland A, Delepine M, Foglio M, Lechner D, Matsuda F, Blanche H, Gut I, Heath S, Lathrop M, Brennan P. Lung cancer susceptibility locus at 5p15.33. *Nat Genet.* 2008 Dec;40(12):1404-6. doi: 10.1038/ng.254.
- McLemore TL, Adelberg S, Liu MC, McMahan NA, Yu SJ, Hubbard WC, Czerwinski M, Wood TG, Storeng R, Lubet RA, et al. Expression of CYP1A1 gene in patients with lung cancer: evidence for cigarette smoke-induced gene expression in normal lung tissue and for altered gene regulation in primary pulmonary carcinomas. *J Natl Cancer Inst.* 1990 Aug 15;82(16):1333-9.
- Mehta R. The potential for the use of cell proliferation and oncogene expression as intermediate markers during liver carcinogenesis. *Cancer Lett.* 1995 Jun 29;93(1):85-102.
- Mikkonen L, Pihlajamaa P, Sahu B, Zhang FP, Jänne OA. Androgen receptor and androgen-dependent gene expression in lung. *Mol Cell Endocrinol.* 2010 Apr 12;317(1-2):14-24. doi: 10.1016/j.mce.2009.12.022.
- Mitsudomi T. Molecular epidemiology of lung cancer and geographic variations with special reference to EGFR mutations. *Transl Lung Cancer Res.* 2014 Aug;3(4):205-11. doi: 10.3978/j.issn.2218-6751.2014.08.04.
- Mollerup S, Ryberg D, Hewer A, Phillips DH, Haugen A. Sex differences in lung CYP1A1 expression and DNA adduct levels among lung cancer patients. *Cancer Res.* 1999 Jul 15;59(14):3317-20.
- Mollerup S, Jørgensen K, Berge G, Haugen A. Expression of estrogen receptors alpha and beta in human lung tissue and cell lines. *Lung Cancer.* 2002 Aug;37(2):153-9.

Mollerup S, Berge G, Baera R, Skaug V, Hewer A, Phillips DH, Stangeland L, Haugen A. Sex differences in risk of lung cancer: Expression of genes in the PAH bioactivation pathway in relation to smoking and bulky DNA adducts. *Int J Cancer*. 2006 Aug 15;119(4):741-4.

Moorthy B, Chu C, Carlin DJ. Polycyclic aromatic hydrocarbons: from metabolism to lung cancer. *Toxicol Sci*. 2015 May;145(1):5-15. doi: 10.1093/toxsci/kfv040.

Monostory K, Pascussi JM, Kóbori L, Dvorak Z. Hormonal regulation of CYP1A expression. *Drug Metab Rev*. 2009;41(4):547-72. doi: 10.1080/03602530903112284.

Motoyama K, Inoue H, Takatsuno Y, Tanaka F, Mimori K, Uetake H, Sugihara K, Mori M. Over- and under-expressed microRNAs in human colorectal cancer. *Int J Oncol*. 2009 Apr;34(4):1069-75.

Murray GI, Melvin WT, Greenlee WF, Burke MD. Regulation, function, and tissue-specific expression of cytochrome P450 CYP1B1. *Annu Rev Pharmacol Toxicol*. 2001;41:297-316.

Nakamura H, Ando K, Shinmyo T, Morita K, Mochizuki A, Kurimoto N, Tatsunami S. Female gender is an independent prognostic factor in non-small-cell lung cancer: a meta-analysis. *Ann Thorac Cardiovasc Surg*. 2011;17(5):469-80.

Nagy-Mignotte H, Guillem P, Vesin A, Toffart AC, Colonna M, Bonnetterre V, Brichon PY, Brambilla C, Brambilla E, Lantuejoul S, Timsit JF, Moro-Sibilot D; Multidisciplinary Thoracic Oncology Group at Grenoble University Hospital. Primary lung adenocarcinoma: characteristics by smoking habit and sex. *Eur Respir J*. 2011 Dec;38(6):1412-9. doi: 10.1183/09031936.00191710.

National Center for Biotechnology Information, updated on 17-Mar-2016, PubMed Gene AHR; Gene ID: 196, access date 27-Mar-2016. <http://www.ncbi.nlm.nih.gov/gene/196>.

National Center for Biotechnology Information, updated on 20-Mar-2016, PubMed Gene AR; Gene ID: 367, access date 27-Mar-2016. <http://www.ncbi.nlm.nih.gov/gene/367>.

National Center for Biotechnology Information, updated on 20-Mar-2016, PubMed Gene CYP1A1; Gene ID: 1543, access date 27-Mar-2016. <http://www.ncbi.nlm.nih.gov/gene/1543>.

National Center for Biotechnology Information, updated on 13-Mar-2016, PubMed Gene CYP1B1; Gene ID: 1545, access date 27-Mar-2016. <http://www.ncbi.nlm.nih.gov/gene/1545>.

National Center for Biotechnology Information, updated on 20-Mar-2016, PubMed Gene ESR1; Gene ID: 2099, access date 27-Mar-2016. <http://www.ncbi.nlm.nih.gov/gene/2099>.

National Center for Biotechnology Information, updated on 23-Mar-2016, PubMed Gene ESR2; Gene ID: 2100, access date 27-Mar-2016. <http://www.ncbi.nlm.nih.gov/gene/2100>.

National Institute of Health, National Cancer Institute, Cancer drugs approved by the Food and Drug Administration (FDA) for lung cancer, updated 15-Mars-2016, access date 12-April-2016. <http://www.cancer.gov/about-cancer/treatment/drugs/lung#1>.

National Institute of Health, National Cancer Institute, U.S. SEER Cancer Statistics Review 1975-2011, Lung and Bronchus. Updated 17-December-2014, access date 04-May-2016. http://seer.cancer.gov/archive/csr/1975_2011/.

Naujokas MF, Anderson B, Ahsan H, Aposhian HV, Graziano JH, Thompson C, Suk WA. The broad scope of health effects from chronic arsenic exposure: update on a worldwide public health problem. *Environ Health Perspect*. 2013 Mar;121(3):295-302. doi: 10.1289/ehp.1205875.

Nebert DW. The Ah locus: genetic differences in toxicity, cancer, mutation, and birth defects. *Crit Rev Toxicol.* 1989;20(3):153-74.

Nebert DW, Dalton TP, Okey AB, Gonzalez FJ. Role of aryl hydrocarbon receptor-mediated induction of the CYP1 enzymes in environmental toxicity and cancer. *J Biol Chem.* 2004 Jun 4;279(23):23847-50.

Ng M, Freeman MK, Fleming TD, Robinson M, Dwyer-Lindgren L, Thomson B, Wollum A, Sanman E, Wulf S, Lopez AD, Murray CJ, Gakidou E. Smoking prevalence and cigarette consumption in 187 countries, 1980-2012. *JAMA.* 2014 Jan 8;311(2):183-92. doi: 10.1001/jama.2013.284692.

Ninomiya F, Yokohira M, Kishi S, Nakano Y, Yamakawa K, Inoue T, Kuno T, Imaida K. Gender-dependent effects of gonadectomy on lung carcinogenesis. 4-(methylnitrosamino)-1-(3-pyridyl)-1-butanone (NNK) in female and male A/J mice. *Oncol Rep.* 2013 Dec;30(6):2632-8. doi: 10.3892/or.2013.2759.

Novello S, Baldini E. Women and lung cancer. *Ann Oncol.* 2006 Mar;17 Suppl 2:ii79-82.

Ochieng J, Nangami GN, Ogunkua O, Miousse IR, Koturbash I, Odero-Marah V, McCawley LJ, Nangia-Makker P, Ahmed N, Luqmani Y, Chen Z, Papagerakis S, Wolf GT, Dong C, Zhou BP, Brown DG, Colacci AM, Hamid RA, Mondello C, Raju J, Ryan EP, Woodrick J, Scovassi AI, Singh N, Vaccari M, Roy R, Forte S, Memeo L, Salem HK, Amedei A, Al-Temaimi R, Al-Mulla F, Bisson WH, Eltom SE. The impact of low-dose carcinogens and environmental disruptors on tissue invasion and metastasis. *Carcinogenesis.* 2015 Jun;36 Suppl 1:S128-59. doi: 10.1093/carcin/bgv034.

O'Connor RJ, Wilkins KJ, Caruso RV, Cummings KM, Kozlowski LT. Cigarette characteristic and emission variations across high-, middle- and low-income countries. *Public Health.* 2010 Dec;124(12):667-74. doi:10.1016/j.puhe.2010.08.018.

Oliveira PA, Colaço A, Chaves R, Guedes-Pinto H, De-La-Cruz P LF, Lopes C. Chemical carcinogenesis. *An Acad Bras Cienc.* 2007 Dec;79(4):593-616.

O'Lone R, Frith MC, Karlsson EK, Hansen U. Genomic targets of nuclear estrogen receptors. *Mol Endocrinol.* 2004 Aug;18(8):1859-75.

Ono-Ogasawara M, Smith TJ. Diesel exhaust particles in the work environment and their analysis. *Ind Health.* 2004 Oct;42(4):389-99.

Page TJ, O'Brien S, Holston K, MacWilliams PS, Jefcoate CR, Czuprynski CJ. 7,12-Dimethylbenz[a]anthracene-induced bone marrow toxicity is p53-dependent. *Toxicol Sci.* 2003 Jul;74(1):85-92.

Palackal NT, Burczynski ME, Harvey RG, Penning TM. Metabolic activation of polycyclic aromatic hydrocarbon trans-dihydrodiols by ubiquitously expressed aldehyde reductase (AKR1A1). *Chem Biol Interact.* 2001 Jan 30;130-132(1-3):815-24.

Palackal NT, Lee SH, Harvey RG, Blair IA, Penning TM. Activation of polycyclic aromatic hydrocarbon trans-dihydrodiol proximate carcinogens by human aldo-keto reductase (AKR1C) enzymes and their functional overexpression in human lung carcinoma (A549) cells. *J Biol Chem.* 2002 Jul 5;277(27):24799-808.

Patel JD, Bach PB, Kris MG. Lung cancer in US women: a contemporary epidemic. *JAMA.* 2004 Apr 14;291(14):1763-8.

Patel JD. Lung cancer in women. *J Clin Oncol.* 2005 May 10;23(14):3212-8.

Patrone C, Cassel TN, Pettersson K, Piao YS, Cheng G, Ciana P, Maggi A, Warner M, Gustafsson JA, Nord M. Regulation of postnatal lung development and homeostasis by estrogen receptor beta. *Mol Cell Biol.* 2003 Dec;23(23):8542-52.

Pelkonen O, Nebert DW. Metabolism of polycyclic aromatic hydrocarbons: etiologic role in carcinogenesis. *Pharmacol Rev.* 1982 Jun;34(2):189-222.

Penaloza CG, Estevez B, Han DM, Norouzi M, Lockshin RA, Zakeri Z. Sex-dependent regulation of cytochrome P450 family members Cyp1a1, Cyp2e1, and Cyp7b1 by methylation of DNA. *FASEB J.* 2014 Feb;28(2):966-77. doi: 10.1096/fj.13-233320.

Perera FP, Weinstein IB. Molecular epidemiology and carcinogen-DNA adduct detection: new approaches to studies of human cancer causation. *J Chronic Dis.* 1982;35(7):581-600.

Perera FP, Santella RM, Brenner D, Poirier MC, Munshi AA, Fischman HK, Van Ryzin J. DNA adducts, protein adducts, and sister chromatid exchange in cigarette smokers and nonsmokers. *J Natl Cancer Inst.* 1987 Sep;79(3):449-56.

Piccardo MT, Stella A, Valerio F. Is the smokers exposure to environmental tobacco smoke negligible? *Environ Health.* 2010 Jan 29;9:5. doi: 10.1186/1476-069X-9-5.

Pirie K, Peto R, Green J, Reeves GK, Beral V; Million Women Study Collaborators. Lung cancer in never-smokers. *Int J Cancer.* 2016 Mar 8. doi: 10.1002/ijc.30084.

Pitot HC, Dragan YP. Facts and theories concerning the mechanisms of carcinogenesis. *FASEB J.* 1991 Jun;5(9):2280-6.

Poirier MC, Beland FA. DNA adduct measurements and tumor incidence during chronic carcinogen exposure in animal models: implications for DNA adduct-based human cancer risk assessment. *Chem Res Toxicol.* 1992 Nov-Dec;5(6):749-55.

Poirier MC. Chemical-induced DNA damage and human cancer risk. *Nat Rev Cancer.* 2004 Aug;4(8):630-7.

Pollitzer E. Biology: Cell sex matters. *Nature.* 2013 Aug 1;500(7460):23-4. doi: 10.1038/500023a.

Portal-Nuñez S, Shankavaram UT, Rao M, Datrice N, Atay S, Aparicio M, Camphausen KA, Fernández-Salguero PM, Chang H, Lin P, Schrupp DS, Garantziotis S, Cuttitta F, Zudaire E. Aryl hydrocarbon receptor-induced adrenomedullin mediates cigarette smoke carcinogenicity in humans and mice. *Cancer Res.* 2012 Nov 15;72(22):5790-800. doi: 10.1158/0008-5472.CAN-12-0818.

Quint LE. Staging non-small cell lung cancer. *Cancer Imaging.* 2007 Oct 22;7:148-59.

Rades D, Setter C, Dahl O, Schild SE, Noack F. The prognostic impact of tumor cell expression of estrogen receptor- α , progesterone receptor, and androgen receptor in patients irradiated for nonsmall cell lung cancer. *Cancer.* 2012 Jan 1;118(1):157-63. doi: 10.1002/cncr.26282.

Ramchandran K, Patel JD. Sex differences in susceptibility to carcinogens. *Semin Oncol.* 2009 Dec;36(6):516-23. doi: 10.1053/j.seminoncol.2009.09.005.

Ricci MS, Toscano DG, Mattingly CJ, Toscano WA Jr. Estrogen receptor reduces CYP1A1 induction in cultured human endometrial cells. *J Biol Chem.* 1999 Feb 5;274(6):3430-8.

- Richter PA, Li AP, Polzin G, Roy SK. Cytotoxicity of eight cigarette smoke condensates in three test systems: comparisons between assays and condensates. *Regul Toxicol Pharmacol*. 2010 Dec;58(3):428-36. doi: 10.1016/j.yrtph.2010.08.009.
- Rifkind AB. CYP1A in TCDD toxicity and in physiology-with particular reference to CYP dependent arachidonic acid metabolism and other endogenous substrates. *Drug Metab Rev*. 2006;38(1-2):291-335.
- Risom L, Møller P, Loft S. Oxidative stress-induced DNA damage by particulate air pollution. *Mutat Res*. 2005 Dec 30;592(1-2):119-37.
- Rivera MP. Lung cancer in women: differences in epidemiology, biology, histology, and treatment outcomes. *Semin Respir Crit Care Med*. 2013 Dec;34(6):792-801. doi: 10.1055/s-0033-1358550.
- Robertson KD. DNA methylation and human disease. *Nat Rev Genet*. 2005 Aug;6(8):597-610.
- Rodriguez A, Viscardi RM, Torday JS. Fetal androgen exposure inhibits fetal rat lung fibroblast lipid uptake and release. *Exp Lung Res*. 2001 Jan -Feb;27(1):13-24.
- Rodriguez C, Spencer Feigelson H, Deka A, Patel AV, Jacobs EJ, Thun MJ, Calle EE. Postmenopausal hormone therapy and lung cancer risk in the cancer prevention study II nutrition cohort. *Cancer Epidemiol Biomarkers Prev*. 2008 Mar;17(3):655-60. doi: 10.1158/1055-9965.EPI-07-2683.
- Roemer E, Dempsey R, Schorp MK. Toxicological assessment of kretek cigarettes: Part 1: background, assessment approach, and summary of findings. *Regul Toxicol Pharmacol*. 2014 Dec;70 Suppl 1:S2-14. doi: 10.1016/j.yrtph.2014.11.015.
- Routledge PA, Shand DG. Presystemic drug elimination. *Annu Rev Pharmacol Toxicol*. 1979;19:447-68.
- Ruano-Ravina A, Rodríguez MC, Cerdeira-Caramés S, Barros-Dios JM. Residential radon and lung cancer. *Epidemiology*. 2009 Jan;20(1):155-6. doi: 10.1097/EDE.0b013e31818ef498.
- Ruiz-Hernandez A, Kuo CC, Rentero-Garrido P, Tang WY, Redon J, Ordovas JM, Navas-Acien A, Tellez-Plaza M. Environmental chemicals and DNA methylation in adults: a systematic review of the epidemiologic evidence. *Clin Epigenetics*. 2015 Apr 29;7(1):55. doi: 10.1186/s13148-015-0055-7.
- Safe S, Kim K. Nuclear receptor-mediated transactivation through interaction with Sp proteins. *Prog Nucleic Acid Res Mol Biol*. 2004;77:1-36.
- Safe S, Lee SO, Jin UH. Role of the aryl hydrocarbon receptor in carcinogenesis and potential as a drug target. *Toxicol Sci*. 2013 Sep;135(1):1-16. doi: 10.1093/toxsci/kft128.
- Sagredo C, Øvrebø S, Haugen A, Fujii-Kuriyama Y, Baera R, Botnen IV, Mollerup S. Quantitative analysis of benzo[a]pyrene biotransformation and adduct formation in Ahr knockout mice. *Toxicol Lett*. 2006 Dec 15;167(3):173-82.
- Sagredo C, Mollerup S, Cole KJ, Phillips DH, Uppstad H, Øvrebø S. Biotransformation of benzo[a]pyrene in Ahr knockout mice is dependent on time and route of exposure. *Chem Res Toxicol*. 2009 Mar 16;22(3):584-91. doi:10.1021/tx8003664.
- Samet JM, Avila-Tang E, Boffetta P, Hannan LM, Olivo-Marston S, Thun MJ, Rudin CM. Lung cancer in never smokers: clinical epidemiology and environmental risk factors. *Clin Cancer Res*. 2009 Sep 15;15(18):5626-45. doi:10.1158/1078-0432.CCR-09-0376.

Sanada N, Gotoh Y, Shimazawa R, Klinge CM, Kizu R. Repression of activated aryl hydrocarbon receptor-induced transcriptional activation by 5 α -dihydrotestosterone in human prostate cancer LNCaP and human breast cancer T47D cells. *J Pharmacol Sci.* 2009 Mar;109(3):380-7.

San Jose C, Cabanillas A, Benitez J, Carrillo JA, Jimenez M, Gervasini G. CYP1A1 gene polymorphisms increase lung cancer risk in a high-incidence region of Spain: a case control study. *BMC Cancer.* 2010 Aug 30;10:463. doi: 10.1186/1471-2407-10-46.

Schabath MB, Wu X, Vassilopoulou-Sellin R, Vaporciyan AA, Spitz MR. Hormone replacement therapy and lung cancer risk: a case-control analysis. *Clin Cancer Res.* 2004 Jan 1;10(1 Pt 1):113-23.

Schrenk D. Impact of dioxin-type induction of drug-metabolizing enzymes on the metabolism of endo- and xenobiotics. *Biochem Pharmacol.* 1998 Apr 15;55(8):1155-62.

Schwartz AG, Ray RM, Cote ML, Abrams J, Sokol RJ, Hendrix SL, Chen C, Chlebowski RT, Hubbell FA, Kooperberg C, Manson JE, O'Sullivan MJ, Rohan T, Stefanick ML, Wactawski-Wende J, Wakelee H, Simon MS. Hormone Use, Reproductive History, and Risk of Lung Cancer: The Women's Health Initiative Studies. *J Thorac Oncol.* 2015 Jul;10(7):1004-13. doi: 10.1097/JTO.0000000000000558.

Shah K, McCormack CE, Bradbury NA. Do you know the sex of your cells? *Am J Physiol Cell Physiol.* 2014 Jan 1;306(1):C3-18. doi: 10.1152/ajpcell.00281.2013.

Sharma D, Newman TG, Aronow WS. Lung cancer screening: history, current perspectives, and future directions. *Arch Med Sci.* 2015 Oct 12;11(5):1033-43. doi: 10.5114/aoms.2015.54859.

Shen H, Yuan Y, Sun J, Gao W, Shu YQ. Combined tamoxifen and gefitinib in non-small cell lung cancer shows antiproliferative effects. *Biomed Pharmacother.* 2010 Feb;64(2):88-92. doi: 10.1016/j.biopha.2009.06.010

Shen H, Tao S, Liu J, Huang Y, Chen H, Li W, Zhang Y, Chen Y, Su S, Lin N, Xu Y, Li B, Wang X, Liu W. Global lung cancer risk from PAH exposure highly depends on emission sources and individual susceptibility. *Sci Rep.* 2014 Oct 9;4:6561. doi: 10.1038/srep06561.

Shimada T, Oda Y, Gillam EM, Guengerich FP, Inoue K. Metabolic activation of polycyclic aromatic hydrocarbons and other procarcinogens by cytochromes P450 1A1 and P450 1B1 allelic variants and other human cytochromes P450 in *Salmonella typhimurium* NM2009. *Drug Metab Dispos.* 2001 Sep;29(9):1176-82.

Shimada T, Fujii-Kuriyama Y. Metabolic activation of polycyclic aromatic hydrocarbons to carcinogens by cytochromes P450 1A1 and 1B1. *Cancer Sci.* 2004 Jan;95(1):1-6.

Shimada T, Guengerich FP. Inhibition of human cytochrome P450 1A1-, 1A2-, and 1B1-mediated activation of procarcinogens to genotoxic metabolites by polycyclic aromatic hydrocarbons. *Chem Res Toxicol.* 2006 Feb;19(2):288-94.

Shimada T. Xenobiotic-metabolizing enzymes involved in activation and detoxification of carcinogenic polycyclic aromatic hydrocarbons. *Drug Metab Pharmacokinet.* 2006 Aug;21(4):257-76.

Siegfried JM, Stabile LP. Estrogenic steroid hormones in lung cancer. *Seminv Oncol.* 2014 Feb;41(1):5-16. doi: 10.1053/j.seminoncol.2013.12.009.

Siegfried JM. Women and lung cancer: does oestrogen play a role? *Lancet Oncol.* 2001 Aug;2(8):506-13.

- Slatore CG, Chien JW, Au DH, Satia JA, White E. Lung cancer and hormone replacement therapy: association in the vitamins and lifestyle study. *J Clin Oncol.* 2010 Mar 20;28(9):1540-6. doi: 10.1200/JCO.2009.25.9739.
- Spitz MR, Wei Q, Dong Q, Amos CI, Wu X. Genetic susceptibility to lung cancer: the role of DNA damage and repair. *Cancer Epidemiol Biomarkers Prev.* 2003 Aug;12(8):689-98.
- Stabile LP, Davis AL, Gubish CT, Hopkins TM, Luketich JD, Christie N, Finkelstein S, Siegfried JM. Human non-small cell lung tumors and cells derived from normal lung express both estrogen receptor alpha and beta and show biological responses to estrogen. *Cancer Res.* 2002 Apr 1;62(7):2141-50.
- Stabile LP, Siegfried JM. Sex and gender differences in lung cancer. *J Gend Specif Med.* 2003;6(1):37-48.
- Stabile LP, Dacic S, Land SR, Lenzner DE, Dhir R, Acquafondata M, Landreneau RJ, Grandis JR, Siegfried JM. Combined analysis of estrogen receptor beta-1 and progesterone receptor expression identifies lung cancer patients with poor outcome. *Clin Cancer Res.* 2011 Jan 1;17(1):154-64. doi: 10.1158/1078-0432.CCR-10-0992.
- Statistics Norway, Smoking habits, updated 14-January-2016, access date 04-May-2016, <https://www.ssb.no/helse/statistikker/royk/aar/2016-01-14>.
- Stockwell HG, Armstrong AW, Leaverton PE. Histopathology of lung cancers among smokers and nonsmokers in Florida. *Int J Epidemiol.* 1990;19 Suppl 1:S48-52.
- Swenberg JA, Fryar-Tita E, Jeong YC, Boysen G, Starr T, Walker VE, Albertini RJ. Biomarkers in toxicology and risk assessment: informing critical dose-response relationships. *Chem Res Toxicol.* 2008 Jan;21(1):253-65.
- Taioli E, Wynder EL. Re: Endocrine factors and adenocarcinoma of the lung in women. *J Natl Cancer Inst.* 1994 Jun 1;86(11):869-70.
- Tan MH, Li J, Xu HE, Melcher K, Yong EL. Androgen receptor: structure, role in prostate cancer and drug discovery. *Acta Pharmacol Sin.* 2015 Jan;36(1):3-23. doi:10.1038/aps.2014.18.
- Tarantini A, Maître A, Lefèbvre E, Marques M, Rajhi A, Douki T. Polycyclic aromatic hydrocarbons in binary mixtures modulate the efficiency of benzo[a]pyrene to form DNA adducts in human cells. *Toxicology.* 2011 Jan 11;279(1-3):36-44. doi: 10.1016/j.tox.2010.09.002.
- Tekpli X, Zienolddiny S, Skaug V, Stangeland L, Haugen A, Mollerup S. DNA methylation of the CYP1A1 enhancer is associated with smoking-induced genetic alterations in human lung. *Int J Cancer.* 2012 Oct 1;131(7):1509-16. doi: 10.1002/ijc.27421.
- Thomas A, Chen Y, Yu T, Jakopovic M, Giaccone G. Trends and Characteristics of Young Non-Small Cell Lung Cancer Patients in the United States. *Front Oncol.* 2015 May 26;5:113. doi: 10.3389/fonc.2015.00113.
- Thomsen JS, Wang X, Hines RN, Safe S. Restoration of aryl hydrocarbon (Ah) responsiveness in MDA-MB-231 human breast cancer cells by transient expression of the estrogen receptor. *Carcinogenesis.* 1994 May;15(5):933-7.
- Thum T, Erpenbeck VJ, Moeller J, Hohlfeld JM, Krug N, Borlak J. Expression of xenobiotic metabolizing enzymes in different lung compartments of smokers and nonsmokers. *Environ Health Perspect.* 2006 Nov;114(11):1655-61.

- Thun M, Peto R, Boreham J, Lopez AD. Stages of the cigarette epidemic on entering its second century. *Tob Control*. 2012 Mar;21(2):96-101. doi:10.1136/tobaccocontrol-2011-050294.
- Torre LA, Siegel RL, Jemal A. Lung Cancer Statistics. *Adv Exp Med Biol*. 2016;893:1-19. doi: 10.1007/978-3-319-24223-1_1.
- Townsend EA, Miller VM, Prakash YS. Sex differences and sex steroids in lung health and disease. *Endocr Rev*. 2012 Feb;33(1):1-47. doi: 10.1210/er.2010-0031.
- Travis WD, Brambilla E, Noguchi M, Nicholson AG, Geisinger K, Yatabe Y, Ishikawa Y, Wistuba I, Flieder DB, Franklin W, Gazdar A, Hasleton PS, Henderson DW, Kerr KM, Petersen I, Roggli V, Thunnissen E, Tsao M. Diagnosis of lung cancer in small biopsies and cytology: implications of the 2011 International Association for the Study of Lung Cancer/American Thoracic Society/European Respiratory Society classification. *Arch Pathol Lab Med*. 2013 May;137(5):668-84. doi: 10.5858/arpa.2012-0263-RA.
- Tsay JJ, Tchou-Wong KM, Greenberg AK, Pass H, Rom WN. Aryl hydrocarbon receptor and lung cancer. *Anticancer Res*. 2013 Apr;33(4):1247-56.
- Tsuchiya Y, Nakajima M, Kyo S, Kanaya T, Inoue M, Yokoi T. Human CYP1B1 is regulated by estradiol via estrogen receptor. *Cancer Res*. 2004 May 1;64(9):3119-25.
- Uno S, Dalton TP, Derkenne S, Curran CP, Miller ML, Shertzer HG, Nebert DW. Oral exposure to benzo[a]pyrene in the mouse: detoxication by inducible cytochrome P450 is more important than metabolic activation. *Mol Pharmacol*. 2004 May;65(5):1225-37.
- Uno S, Dalton TP, Dragin N, Curran CP, Derkenne S, Miller ML, Shertzer HG, Gonzalez FJ, Nebert DW. Oral benzo[a]pyrene in Cyp1 knockout mouse lines: CYP1A1 important in detoxication, CYP1B1 metabolism required for immune damage independent of total-body burden and clearance rate. *Mol Pharmacol*. 2006 Apr;69(4):1103-14.
- Vahter M, Gochfeld M, Casati B, Thiruchelvam M, Falk-Filippson A, Kavlock R, Marafante E, Cory-Slechta D. Implications of gender differences for human health risk assessment and toxicology. *Environ Res*. 2007 May;104(1):70-84.
- van Loon AJ, Kant IJ, Swaen GM, Goldbohm RA, Kremer AM, van den Brandt PA. Occupational exposure to carcinogens and risk of lung cancer: results from The Netherlands cohort study. *Occup Environ Med*. 1997 Nov;54(11):817-24.
- Vasudevan S, Tong Y, Steitz JA. Switching from repression to activation: microRNAs can up-regulate translation. *Science*. 2007 Dec 21;318(5858):1931-4.
- Veglia F, Loft S, Matullo G, Peluso M, Munnia A, Perera F, Phillips DH, Tang D, Autrup H, Raaschou-Nielsen O, Tjønneland A, Vineis P; Genair-EPI Investigators. DNA adducts and cancer risk in prospective studies: a pooled analysis and a meta-analysis. *Carcinogenesis*. 2008 May;29(5):932-6. doi: 10.1093/carcin/bgm286.
- Villeneuve PJ, Mao Y. Lifetime probability of developing lung cancer, by smoking status, Canada. *Can J Public Health*. 1994 Nov-Dec;85(6):385-8.
- Vineis P, Veglia F, Benhamou S, Butkiewicz D, Cascorbi I, Clapper ML, Dolzan V, Haugen A, Hirvonen A, Ingelman-Sundberg M, Kihara M, Kiyohara C, Kremers P, Le Marchand L, Ohshima S, Pastorelli R, Rannug A, Romkes M, Schoket B, Shields P, Strange RC, Stucker I, Sugimura H, Garte S, Gaspari L, Taioli E. CYP1A1 T3801 C polymorphism and lung cancer: a pooled analysis of 2451 cases and 3358 controls. *Int J Cancer*. 2003 May 1;104(5):650-7.

- Vineis P, Husgafvel-Pursiainen K. Air pollution and cancer: biomarker studies in human populations. *Carcinogenesis*. 2005 Nov;26(11):1846-55.
- Wakelee HA, Chang ET, Gomez SL, Keegan TH, Feskanich D, Clarke CA, Holmberg L, Yong LC, Kolonel LN, Gould MK, West DW. Lung cancer incidence in never smokers. *J Clin Oncol*. 2007 Feb 10;25(5):472-8.
- Wang CK, Chang H, Chen PH, Chang JT, Kuo YC, Ko JL, Lin P. Aryl hydrocarbon receptor activation and overexpression upregulated fibroblast growth factor-9 in human lung adenocarcinomas. *Int J Cancer*. 2009 Aug 15;125(4):807-15. doi: 10.1002/ijc.24348.
- Watanabe K, Emoto N, Hamano E, Sunohara M, Kawakami M, Kage H, Kitano K, Nakajima J, Goto A, Fukayama M, Nagase T, Yatomi Y, Ohishi N, Takai D. Genome structure-based screening identified epigenetically silenced microRNA associated with invasiveness in non-small-cell lung cancer. *Int J Cancer*. 2012 Jun 1;130(11):2580-90. doi: 10.1002/ijc.26254.
- Watson PA, Chen YF, Balbas MD, Wongvipat J, Socci ND, Viale A, Kim K, Sawyers CL. Constitutively active androgen receptor splice variants expressed in castration-resistant prostate cancer require full-length androgen receptor. *Proc Natl Acad Sci U S A*. 2010 Sep 28;107(39):16759-65. doi: 10.1073/pnas.1012443107.
- Wei R, DeVilbiss FT, Liu W. Genetic Polymorphism, Telomere Biology and Non-Small Lung Cancer Risk. *J Genet Genomics*. 2015 Oct 20;42(10):549-61. doi:10.1016/j.jgg.2015.08.005.
- Wei SJ, Chang RL, Wong CQ, Bhachech N, Cui XX, Hennig E, Yagi H, Sayer JM, Jerina DM, Preston BD, et al. Dose-dependent differences in the profile of mutations induced by an ultimate carcinogen from benzo[a]pyrene. *Proc Natl Acad Sci U S A*. 1991 Dec 15;88(24):11227-30.
- Weickert CS, Miranda-Angulo AL, Wong J, Perlman WR, Ward SE, Radhakrishna V, Straub RE, Weinberger DR, Kleinman JE. Variants in the estrogen receptor alpha gene and its mRNA contribute to risk for schizophrenia. *Hum Mol Genet*. 2008 Aug 1;17(15):2293-309. doi: 10.1093/hmg/ddn130.
- Weinberg OK, Marquez-Garban DC, Fishbein MC, Goodglick L, Garban HJ, Dubinett SM, Pietras RJ. Aromatase inhibitors in human lung cancer therapy. *Cancer Res*. 2005 Dec 15;65(24):11287-91.
- Whitlock JP Jr. Induction of cytochrome P4501A1. *Annu Rev Pharmacol Toxicol*. 1999;39:103-25.
- Wilson CL, Bodnar JA, Brown BG, Morgan WT, Potts RJ, Borgerding MF. Assessment of dioxin and dioxin-like compounds in mainstream smoke from selected US cigarette brands and reference cigarettes. *Food Chem Toxicol*. 2008 May;46(5):1721-33. doi: 10.1016/j.fct.2008.01.009.
- Wilson CM, McPhaul MJ. A and B forms of the androgen receptor are expressed in a variety of human tissues. *Mol Cell Endocrinol*. 1996 Jun 18;120(1):51-7.
- Wolfe AR, Smith TJ, Meehan T. Benzo[a]pyrene diol epoxide forms covalent adducts with deoxycytidylic acid by alkylation at both exocyclic amino N(4) and ring imino N-3 positions. *Chem Res Toxicol*. 2004 Apr;17(4):476-91.
- Wong PS, Vogel CF, Kokosinski K, Matsumura F. Arylhydrocarbon receptor activation in NCI-H441 cells and C57BL/6 mice: possible mechanisms for lung dysfunction. *Am J Respir Cell Mol Biol*. 2010 Feb;42(2):210-7. doi: 10.1165/rcmb.2008-0228OC.
- Wormke M, Stoner M, Saville B, Walker K, Abdelrahim M, Burghardt R, Safe S. The aryl hydrocarbon receptor mediates degradation of estrogen receptor alpha through activation of proteasomes. *Mol Cell Biol*. 2003 Mar;23(6):1843-55.

Wu AH, Yu MC, Thomas DC, Pike MC, Henderson BE. Personal and family history of lung disease as risk factors for adenocarcinoma of the lung. *Cancer Res.* 1988 Dec 15;48(24 Pt 1):7279-84.

Yan M, Chen X, Wang S, Li Y. Expression of ER and AR in lung cancer. *Zhongguo Fei Ai Za Zhi.* 2008 Feb 20;11(1):126-9. doi:10.3779/j.issn.1009-3419.2008.01.027.

Yanagida A, Sogawa K, Yasumoto KI, Fujii-Kuriyama Y. A novel cis-acting DNA element required for a high level of inducible expression of the rat P-450c gene. *Mol Cell Biol.* 1990 Apr;10(4):1470-5.

Yang IA, Holloway JW, Fong KM. Genetic susceptibility to lung cancer and co-morbidities. *J Thorac Dis.* 2013 Oct;5 Suppl 5:S454-62. doi:10.3978/j.issn.2072-1439.2013.08.06.

Yang SH, Mechanic LE, Yang P, Landi MT, Bowman ED, Wampfler J, Meerzaman D, Hong KM, Mann F, Dracheva T, Fukuoka J, Travis W, Caporaso NE, Harris CC, Jen J. Mutations in the tyrosine kinase domain of the epidermal growth factor receptor in non-small cell lung cancer. *Clin Cancer Res.* 2005 Mar 15;11(6):2106-10.

Yang XR, Wacholder S, Xu Z, Dean M, Clark V, Gold B, Brown LM, Stone BJ, Fraumeni JF Jr, Caporaso NE. CYP1A1 and GSTM1 polymorphisms in relation to lung cancer risk in Chinese women. *Cancer Lett.* 2004 Oct 28;214(2):197-204.

Yeh S-D, Yang P-C, Lu H-H, Chang C, Wu C-W. Targeting androgen receptor as a new potential therapeutic approach to battle tobacco carcinogens-induced non-small cell lung cancer From 2012 Sino-American Symposium on Clinical and Translational Medicine (SAS-CTM) Shanghai, China. 27-29 June 2012

Yeh WL, Shioda K, Coser KR, Rivizzigno D, McSweeney KR, Shioda T. Fulvestrant-induced cell death and proteasomal degradation of estrogen receptor α protein in MCF-7 cells require the CSK c-Src tyrosine kinase. *PLoS One.* 2013 Apr 4;8(4):e60889. doi: 10.1371/journal.pone.0060889.

Youlden DR, Cramb SM, Baade PD. The International Epidemiology of Lung Cancer: geographical distribution and secular trends. *J Thorac Oncol.* 2008 Aug;3(8):819-31. doi: 10.1097/JTO.0b013e31818020eb.

Young RP, Hopkins RJ, Hay BA, Epton MJ, Mills GD, Black PN, Gardner HD, Sullivan R, Gamble GD. Lung cancer susceptibility model based on age, family history and genetic variants. *PLoS One.* 2009;4(4):e5302. doi: 10.1371/journal.pone.0005302.

Zheng W, Jefcoate CR. Steroidogenic factor-1 interacts with cAMP response element-binding protein to mediate cAMP stimulation of CYP1B1 via a far upstream enhancer. *Mol Pharmacol.* 2005 Feb;67(2):499-512.

